











SMITHSONIAN INSTITUTION UNITED STATES NATIONAL MUSEUM

Bulletin 125

NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA

BY

FERDINAND CANU

Versailles, France

AND

RAY S. BASSLER

Washington, District of Columbia



WASHINGTON GOVERNMENT PRINTING OFFICE 1923

ADVERTISEMENT.

The scientific publications of the United States National Museum consist of two series, the *Proceedings* and the *Bulletins*.

The Proceedings, the first volume of which was issued in 1878, are intended primarily as a medium for the publication of original, and usually brief, papers based on the collections of the National Museum, presenting newly-acquired facts in zoology, geology, and anthropology, including descriptions of new forms of animals, and revisions of limited groups. One or two volumes are issued annually and distributed to libraries and scientific organizations. A limited number of copies of each paper, in pamphlet form, is distributed to specialists and others interested in the different subjects as soon as printed. The date of publication is printed on each paper, and these dates are also recorded in the tables of contents of the volumes.

The Bulletins, the first of which was issued in 1875, consist of a series of separate publications comprising chiefly monographs of large zoological groups and other general systematic treatises (occasionally in several volumes), faunal works, reports of expeditions, and catalogues of type-specimens, special collections, etc. The majority of the volumes are octavos, but a quarto size has been adopted in a few instances in which large plates were regarded as indispensable.

Since 1902 a series of octavo volumes containing papers relating to the botanical collections of the Museum, and known as the *Contributions from the National Herbarium*, has been published as bulletins.

The present work forms No. 125 of the Bulletin series.

WILLIAM DEC. RAVENEL,
Administrative Assistant to the Secretary,
In charge of the United States National Museum.

Washington, D. C., May 14, 1923.

TABLE OF CONTENTS.

	Page.
Introduction	rage.
Catalogue of papers of American Later Tertiary and Quaternary Bryozoa.	2
Lists of North American Later Tertiary and Quaternary Bryozoan faunas.	4
Systematic descriptions	15
Order Ctenostomata Busk	15
Family Terehriporidae Busk, 1839.	15
Genus Terebripora D'Orbigny, 1839	15
Genus Spathipora Fischer, 1866.	16
Order Cheilostomata Busk.	17
Suborder Anasca Levinsen.	17
Division I. Malacostega Levinsen.	17.
Family Electrinidae D'Orbigny, 1851.	17
Genus Electra Lamouroux, 1816.	17
Genus Pyripora D'Orbigny, 1852.	19
Genus Mystriopora Lang, 1915.	19
Genus Membranipora Blainville, 1834	20
Group Membraniporae Canu and Bassler, 1917.	24
Section 1. No ovicell.	24
Genus Membraniporina Levinsen, 1909.	24
Genus Conopeum Norman, 1903.	26
Genus Cupuladria Canu and Bassler, 1919.	28
Genus Acanthodesia Canu and Bassler, 1919.	30
Section II. Membraniporae with endozooecial ovicell.	35
Genus Vibracellina Canu and Bassler, 1917.	35
Genus Membrendoecium Canu and Bassler, 1917	36
Genus Ogivalina Canu and Bassler, 1917.	37
Genus Hincksina Norman, 1903.	38
Section III. Ovicell hyperstomial, always closed by the operculum.	39
Genus Membraniporidra Canu and Bassler, 1917.	39
Section IV. Ovicell never closed by the operculum.	39
Genus Alderina Norman, 1903	39
Genus Callopora, Gray, 1848.	40
Genus Amphiblestrum Gray, 1848.	44
Genus Ramphonotus Norman, 1894.	46
Genus Cauloramphus Norman, 1903.	48
Family Aeteidae Smitt, 1867.	49
Genus Aetea Lamouroux, 1812.	49
Family Chaperiidae Jullien, 1888.	49
Genus Chaperia Jullien, 1881	49
Family Farciminariidae Busk, 1852.	55
Genus Nellia Busk, 1852.	55
Family Opesiulidae Jullien, 1888.	56
Genus Floridina Jullien, 1881	56
Genus Velumella Canu and Bassler, 1917.	58
Genus Micropora Gray, 1848.	58
Genus Selenaria Busk, 1854	59

Systematic descriptions - Continued	
Order Cheilostomata Busk - Continued.	Page.
Family Thalamoporellidae Levinsen, 1909.	
Genus Manzonella Jullien, 1888.	. 60
Genus Woodipora Jullien, 1888.	. 61
Genus Thalamoporella Hincks, 1887.	. 61
Family Steganoporellidae Levinsen, 1909	. 62
Genus Steganoporella Smitt, 1873.	62
Family Aspidostomidae Canu, 1908.	. 64
Genus Odontionella Canu and Bassler, 1917.	. 64
Genus Foraminella Levinsen, 1909.	. 66
Genus Labiopora Levinsen, 1909	. 67
Family Calmonsides new family	- 107
Family Calpensiidae, new family.	. 67
Genus Diplodidymia Reuss, 1869.	. 67
Genus Hemiseptella Levinsen, 1909.	. 69
Genus Cupularia Lamouroux, 1821	. 75
Genus Calpensia Jullien, 1888.	. 82
Genus Verminaria Jullien, 1888.	. 83
Genus Microporina Levinsen, 1909.	. 83
Genus Corynostylus Canu and Bassler, 1919	. 84
Family Cellariidae Hincks, 1880	. 85
Genus Cellaria Authors.	. 85
Genus Erina Canu, 1908.	. 87
Genus Melicerita Milne-Erlwards, 1836	87
Genus Euginoma Jullien, 1882	5.7
The Costulae (Family Cribrilinidae Hincks, 1880)	87
Genus Cribrilina Gray, 1848.	87
Genus Tuellina Jullien, 1886	80
Genus Metracolposa Canu and Bassler, 1917	(19)
ramily Hippothoidae Levinsen, 1909.	0.0
Genus Hippothoa (Lamouroux 1821) Hineks, 1880	69
Genus Trypostega Levinsen, 1909	0.5
ramny Escharellidae Levinsen, 1909.	0.5
Group 1. Schizoporellae Canu and Bassler, 1917	05
Genus Schizoporella Hincks, 1880.	95
Genus Arthropoma Levinsen, 1909	Ð≈
Genus Dakaria Jullien, 1903	D=
Genus Lacerna Jullien, 1888.	197
Genus Stephanosella Canu and Bassler, 1917.	99
Genus Stylopoma Levinsen, 1909.	99
Genus Schizopodrella Canu and Bassler, 1917.	101
Genus Schizolavella Canu and Bassler, 1920.	104
Genus Schizomavella Canu and Bassler, 1920.	108
Genus Gemelliporella Canu and Bassler, 1920.	109
Group 2. Microporellae Canu and Bassler, 1917.	110
Genus Fenestrulina Jullien, 1888.	113
Genus Wigrouperlle Hinele 1987	113
Genus Microporella Hincks, 1877.	117
Genus Diporula Hineks, 1879.	126
Genus Ellipsopora, new subgenus.	127
Cichus i Iusualiininia (Tia), 1548	
rends canoportia Neviani, 1895	
TORUS INVESTIGATION INTO	
Choup 5. http://opolae.cana.and.hassier.lai/	
Circle 111/1/04/01/11/11/11/11/11/11/11/11/11/11/11/11/	
Genus improdipiosia Cand, 1816	
Genus Impromenena Canu and Rassier 1917	
trends rippoporena cand and Bassier, 1920	132
Genus Lepralia Johnston, 1847.	100

vstematic descriptions—Continued.	
Order Cheilostomata Busk—Continued.	
Family Escharellidae Levinsen, 1909—Continued.	Page.
Group 4 Peristomellae Capu and Bassler, 1917	134
Comma Trypometalla Canu and Bassler, 1920	134
Group 5 Divers genera	135
Conus Cyclocolposa Canu and Bassler, 1920	135
Genus Cycloneriella Canu and Bassler, 1920	137
Conus Cyclicopora Hincks 1884	138
Ganus Aimulosia Jullien, 1888	139
Family Furystamollidae Levinsen, 1909	14I
Ganna Furustomella Levinsen, 1909	142
Family Stomachetosellidae Canu and Bassler, 1917	142
Genus Leiosella Canu and Bassler, 1917.	142
Family Smittinidac Levinsen, 1909.	143
Genus Smittina Norman, 1903.	143
Genus Porella Gray, 1848.	147
Genus Porella Gray, 1848	149
Genus Rhamphostomella Lorenz, 1886.	151
Genus Cystisella Canu and Bassler, 1917.	152
Genus Cystisella Canu and Bassier, 1917	152
Family Reteporidae Smitt, 1867.	152
Genus Retepora Imperato, 1599	154
Genus Phidolopora Gabb and Horn, 1862	155
Genus Rhynchozoon Hincks, 1891.	158
Family Adeonidae Jullien, 1903.	158
Genus Adeona (Lamouroux, 1816) Levinsen, 1909.	160
Genus Bracebridgia MacGillivray, 1886	160
Genus Laminopora Michelin, 1842.	161
Genus Adeonellopsis MacGillivray, 1886.	162
Genus Anarthropora Smitt, 1867.	162
Family Hippopodinidae Levinsen, 1909.	162
Genus Metrarabdotos Canu, 1914	
Genus Hippaliosina Canu, 1918.	. 167
Genus Tremogasterina Canu, 1911	
Genus Cheiloporina, new genus	. 170
Family Tulucellariidae Busk, 1884	
Conne Tubucellaria D'Orbigny, 1852	
B Dhylaotallidae Canu and Bassler, 1917	· T(,
Conver Phylocially Hineks 1880	. 210
Canua Lagoninora Hincks 1877	
Carrie Wastigenhora Hincks 1880	
Converte Tomorbio Iullion 1889	
Come Cyanida antha Levinsen, 1909	
Benily Colloporides Rusk 1852	- 1
Conva Holoporalla Waters, 1909	-
Conve Schigmonora MacGillivray, 1888	. 10
Conuc Costaggia Noviani, 1895	
Converted Torminula Jullien, 1882	- 1
Carrie Collopora Linnaeus, 1767	. 10
E-wile Meriogoidae Smitt 1868	
Conve Marriozoum Donati 1750	
Come Myriogodla Levinsen 1909	
Danilar Onlitulinoridae new family	
Conus Ratonora Reuss 1867	
Genus Orbitulipora Stoliczka, 1861	. 18

Systematic descriptions- Continued.	
Order Cheilostomata Busk—Continued.	
Family Orbituliporidae, new family—Continued.	Page.
Genus Stichoporina Stoliczka, 1861.	189
Genus Sphaerophora Haswell, 1881	190
Genus Schizorthosecos Canu and Bassler, 1917.	190
Genus Fedora Jullien, 1882	190
Genus Mamillopora Smitt, 1873.	191
Order Cyclostomata Busk	193
Family Heteroporidae Pergens and Meunier, 1886	193
Genus Ceriopora Goldfuss, 1827.	193
Family Diastoporidae Gregory, 1899.	193
Genus Proboscina Audouin, 1826.	193
Genus Berenicea Lamouroux, 1821.	194
Genus Atelesopora, new genus	194
Family Mecynoeciidae Canu, 1918.	195
Genus Entalophora Lamouroux, 1821	195
Family Oncousoeciidae Canu, 1918.	195
Genus Filisparsa D'Orbigny, 1853.	195
Family Crisiidae Johnston, 1847	196
Genus Crisia Lamouroux, 1816.	196
Family Tubuliporidae Johnston, 1838.	197
Genus Tubulipora Lamarck, 1816.	197
Genus Idmonea Lamouroux, 1821.	198
Genus Crisina D'Orbigny, 1852	200
Family Theonoidae Busk, 1859	201
Genus Theonoa Lamouroux, 1821.	201
Family Diaperoeciidae Canu, 1918.	201
Genus Stathmepora Canu and Bassier, 1922.	201
Genus Diaperoecia Canu, 1918.	201
Family Lichenoporidae Smitt, 1866.	203
Genus Lichenopora Defrance, 1823.	203
Family Tretocycloeciidae Canu, 1918	203
Genus Tretocycloecia Canu, 1918.	206
Genus Psilosolen Canu and Bassler, 1922.	
	207
Bibliography of literature on Bryozoa since 1899.	209
Appendix.	244
Explanation of plates.	245
Index	293

LIST OF ILLUSTRATIONS.

TEXT FIGURES.

	707	18
Fig.	I. Genera of the family Electrinidae D'Orbigny, 1851	30
	a a la Jania Cany and Rassler 1919	50
	70	51
	TV-1 1000 Loringon 1909	60
		61
	and the Manager of th	63
	a to the Champaporollidge Levinsen 1909	65
	a de la familla Appidogtómidae Canii, 1908	66
	a continue of the Converse of Reselver 1917	68
	the Colmongidae now family	69
	Total 1/1 : David 1960	72
	TT 11. I	76
	7 7 1 Town account 1001	85
	and the College Handle 1880	101
	~ 0. 1 T. Junear 1000	109
	a st 1 H Class and Decelor 1920	112
	3 3 3 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	114
	to a Million and loo	116
	11 T.111 am 1000	118
	11. Trimales 1977	125
		127
	T311'	127
		128
		129
		141
		153
	and the state of t	161
		166
	and the state of t	
	7 1 1 Taringon 1000	
	ng 1 11 Common 1000	
	37. Genus Orbitulipora Stoliczka, 1861. 38. Genus Stichoporina Stoliczka, 1861.	



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

By Ferdinand Canu,
Of Versailles, France,
AND
RAY S. BASSLER,
Of Washington, District of Columbia.

INTRODUCTION.

The present volume contains the results of researches upon the Post-Oligocene fossil bryozoa of North America and forms the concluding part of our studies upon the Tertiary and Quaternary faunas, those of the Eocene and Oligocene epochs having been published in 1920 under the title of North American Early Tertiary Bryozoa. The present work, like the companion volume on the Early Tertiary faunas, was undertaken under the joint anspices of the United States Geologica Survey and the United States National Museum. Almost without exception all the type specimens described and illustrated in the present volume are contained in the paleontological collections of the United States National Museum.

The authors are deeply indebted to Dr. Charles D. Walcott, Secretary of the Smithsonian Institution, and Mr. W. deC. Ravenel, Administrative Assistant in charge United States National Museum, who have arranged for the publication of the work and have extended various courtesies to us during its preparation. Dr. T. Wayland Vanghan, of the United States Geological Survey, has likewise spared no effort in assisting us to bring the work to a successful conclusion and we are greatly indebted to him, as well as to other members of the Federal Survey, particularly Mr. Wendell C. Mansfield, Mr. R. D. Mesler, and Mr. I. B. Milner.

We are under many obligations to Mr. F. Julius Fohs, chief geologist of the Humphreys Mexia Oil Company at Mexia. Texas, who has very generously assisted us financially in the preparation and illustration of the volume, and who has thus shown his appreciation of the value of the bryozoa in stratigraphic and economic work.

Through several grants from the Marsh Fund of the National Academy of Sciences and from the American Association for the Advancement of Science we have been able to carry on supplementary studies of other fossil and recent bryozoan faunas which were quite necessary in the preparation of this volume. For this assistance we are highly grateful, as we have thus been enabled to pursue our researches on a larger scale and to secure more definite and lasting results.

Our hearty thanks are due to Dr. F. C. Clark, of Los Angeles, California, and Dr. Ralph Arnold, of Pasadena, California, who have supplied us with practically all of the faunas described from the Pleistocene of California.

The methods of study of the bryozoa and various other subjects relating to them in general have been described in our Early Tertiary volume, to which the reader is referred. We have, however, added to the present volume a bibliography of bryozoan literature published since 1899, the list of literature up to this date being contained in the work by Nickles and Bassler of 1900.2

Although the present work deals primarily with North American Post-Oligocene fossil Bryozoa, the descriptions of a few small faunas from the Oligocene rocks of the West Indies and of several miscellaneous Eocene species are added for various reasons.

CATALOGUE OF PAPERS OF AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

[A few papers by Cann, Ulrich, and Bassler, published since 1900, are not listed here because their contents are reproduced in the present work.]

CONRAD, T. A. Observations on the Secondary and Tertiary formations of the southern Atlantic States, by James T. Hodge. With an appendix by T. A. Conrad. (Amer. Journ. Sci., ser. 1. vol. 41, 1841, pp. 344-348.)

The bryozoa in the paper are:

Lunulites denticulata, p. 348.

Luxulites depressa, p. 348.

1845.

Lonsdale, William. Report on the Corals from the Tertiary formations of North America. (Quart. Journ. Geol. Soc. London, vol. 1, 1845, pp. 495-509.)

Heteropora? tortilis, new species, p. 500. Miocene: Williamsburg, Petersburg.

Escharina tumidula, new species, p. 502. Miocene: Petersburg.

Lunulites denticulata Conrad, p. 503. Miocene: Petersburg.

Cellepora informata, new species, p. 505. Miocene: Petersburg, Virginia. Cellepora umbilicata, new species, p. 507. Miocene: Petersburg.

Cellepora quadrangularis, new species, p. 508. Miocene: Williamsburg, Evergreen.

Cellepora similis, new species, p. 509. Miocene: Williamsburg.

CONRAD, T. A. Note on the Miocene and Post-Pliocene deposits of California, with descriptions of two new fossil corals. (Proc. Acad. Nat. Sci. Philadelphia, vol. 7, 1855, p. 441.)

Idmonea californica, p. 441. Miocene: Santa Barbara, California.

Lichenopora californica, p. 441. Miocene: Santa Barbara, California.

1857.

TUOMEY, M., and HOLMES, F. S. Pleiocene Fossils of South Carolina. Charleston, 1857, xvi+ 152 pp., 30 pls.

The bryozoa are:

Lunulites denticulata Conrad. p. 11, pl. 4, figs. 1-5. Darlington district, South Carolina.

Cellepora tessellata, new species, p. 13, pl. 4, fig. 7. Giles Bluff, Peedee River.

Cellepora radiata, new species, p. 13, pl. 4, fig. 8. Smith's, Goose Creek.

Cellepora depressa, new species, p. 14, pl. 4, fig. 9. Smith's, Goose Creek,

Membranipora lacinia, new species, p. 14, pl. 4, fig. 10. Smith's, Goose Creek.

² A Synopsis of American Fossil Bryozoa, including Bibliography and Synonymy. Bull. No. 173, United States Geological Survey.

Reptocelleporaria informata (Lonsdale), p. 15, pl. 4, figs. 11, 12. Darlington district.

Reptocelleporaria similis (Lonsdale), p. 16, pl. 4, figs. 13, 14. Darlington district, South Carolina; Petersburg, Virginia.

Heteropora tortilis Lonsdale, p. 16, pl. 4, figs. 15, 16. Smith's, Goose Creek.

EMMONS, EBENEZER. Report of the North Carolina Geological Survey. Raleigh, 1858. Paleontology, pp. 193-314.

Lunulites denticulata, p. 311, figs. 248, 249. Miocene: Beds on Neuse and Cape Fear rivers.

Lunulites oblongus, new species, p. 312, figs. 252, 253. Locality not given.

Discoporella umbellata, p. 312, figs. 254, 255. Locality not given.

1860.

HOLMES, Francis S. Postpliocene fossils of South Carolina. Charleston, 1860. xii+v+122 pp., 28 pls.

The bryozoa described are:

Reptocelleporaria informata (Lonsdale), p. 6, pl. 1, fig. 5. Charleston, South Carolina. Lunulites denticulata Conrad, p. 6, pl. 2, figs. 4-4b. Simmons'; Abbapoola; St. Andrew's.

1862

GABB, WILLIAM M., and HORN, GEORGE H. Monograph of the fossil Polyzoa of the Secondary and Tertiary formations of North America. (Journ. Acad. Nat. Sci. Philadelphia, ser. 2, vol. 5, 1862, pp. 111-178, pls. 19-21.)

Eschara? fragilissima, new species, p. 118, pl. 19, 3. Miocene: St. Mary's River, Maryland.

Lunulites oblonga Emmons, p. 121. Miocene(?): North Carolina.

Cellepora tumidula D'Orbigny, p. 127. Miocene: Petersburg, Virginia.

Cellepora formosa Tuomey and Holmes, p. 129. Miocene (?Pliocene): Darlington district, South Carolina.

Cellepora tessellata Tuomey and Holmes, p. 129. Miocene (?Pliocene): Giles Bluff, Peedee River, South Carolina.

Cellepora radiata Tuomey and Holmes, p. 129. Miocene (?Pliocene): Goose Creek, South Carolina. Cellopora de pressa Tuomey and Holmes, p. 129. Miocene (?Pliocene): Goose Creek, South Carolina. Cellepora californiensis, new species, p. 130, pl. 19, fig. 12. Postpliocene: Santa Barbara, California. Cellepora bellerophon, new species, p. 130, pl. 19, fig. 13. Postpliocene: Santa Barbara, California. Reptocelleporaria informata D'Orbigny, p. 132. Miocene: Petersburg, Virginia; and South Carolina. Reptocelleporaria quadrangularis D'Orbigny, p. 132. Locality not given.

Reptocelle poraria similis D'Orbigny, p. 133. Miocene (?Pliocene): Virginia and South Carolina. Reptescharella hermannii, new species, p. 137, pl. 19, fig. 20. Postpliocene: Santa Barbara, California. Reptescharella plana, new species, p. 137, pl. 19, fig. 19. Postpliocene: Santa Barbara, California. Phidolopora labiata, new species, p. 138, pl. 19, fig. 21. Postpliocene: Santa Barbara, California. Ennallipora quadrangularis, new species, p. 141, pl. 20, fig. 24. Miocene: Petersburg, Virginia.

Discoporella denticulata (Conrad), p. 142, pl. 20, fig. 25. Miocene: Generally distributed from New Jersey to South Carolina.

Reptoporina eustomata, new species, p. 144, pl. 20, fig. 26. Postpliocene: Santa Barbara, California. Multiporina umbilicata (Lonsdale), p. 145, pl. 20, fig. 27. Miocene: Petersburg, Virginia. Reptescharellina disparilis, new species, p. 147, pl. 20, fig. 29. Postpliocene: Santa Barbara, California. Reptescharellina? hermannii, new species, p. 147, pl. 20, fig. 30. Postpliocene: Santa Barbara,

California. Reptescharellina cornuta, new species, p. 147, pl. 20, fig. 31. Postpliocene: Santa Barbara, California. Siphonella multipora, new species, p. 154, pl. 20, fig. 38. Postpliocene: Santa Barbara, California. Membranipora sexpunctata, new species, p. 159, pl. 20, fig. 44. Miocene or Eocene: Locality unknown. Membranipora speciosa (Gabb and Horn), p. 159, pl. 20, fig. 45. Miocene(?): Chiriqui, Central America.

Membranipora californica, new species, p. 160, pl. 20, fig. 46. Postpliocene: Santa Barbara, California.
Membranipora barbarensis, new species, p. 160, pl. 20, fig. 47. Postpliocene: Santa Barbara, California.

Reptoftustrella tubulata, new species, p. 162, pl. 20, fig. 51. With Membranipora sexpunctata. Locality unknown, probably from the Virginia Miocene.

Pyriflustrella tuberculum D'Orbigny, p. 163. Locality not given.

Idmonca californica Conrad, p. 168, pl. 21, fig. 56. Postpliocene: Santa Barbara, California. Scmitubigera tuba, new species, p. 169, pl. 21, fig. 57. Postpliocene: Santa Barbara, California. Entalophora punctulata, new species, p. 171, pl. 21, fig. 61. Postpliocene: Santa Barbara, California. Crisina serrata, new species, p. 174, pl. 21, fig. 66. Postpliocene: Santa Barbara, California. Lichenopora california Conrad, p. 176, pl. 21, fig. 68. Postpliocene: Santa Barbara, California. Multicrescis tortilis (Lonsdale), p. 178. Miocene Virginia and South Carolina.

LISTS OF NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOAN FAUNAS.

[e.=common; v. e.=very common; r.=rare; v. r.=very rare.]

OLIGOCENE

```
Oligocene (Antigua formation): Rifle butts, Antigua, Leeward Islands (loc. No. 6854)-
     Floridina fusifera Canu and Bassler, 1919. (v. r.)
     Floridina pyripora Canu and Bassler, 1919. (r.)
    Hippoporina lata Smitt, 1862. (r.)
    Porella bella Busk, 1860. (r.)
    Puellina radiata carolinensis Gabb and Horn, 1862. (r.)
Oligocene (Antigua formation): Carlisle marl pit, Antigua. Leeward Islands (loc. No. 6873)-
    Calpensia impressa Moll, 1803. (r.)
Oligocene (Anguilla formation): Southwest side of Crocus Bay and vicinity, Anguilla, Leeward Islands-
    Callopora dumerilli Savigny-Audouin 1826. +r.)
    Floridina pyripora Canu and Bassler, 1919. (r.)
    Holoporella albirostris Smitt, 1872. (r.)
    Ogivalina mutabilis Canu and Bassler, 1919. (r.)
    Puellina radiata carolinensis Gabb and Horn, 1862. (r.
    Stephanosella biaperta Michelin, 1842. (r.)
    Stylopoma spongites Pallas, 1766. (r.)
Oligocene (Emperador limestone): one-third mile north of west of Empire, Panama Canal Zone (loc.
  No. 6016)-
    Holoporella albirostris Smitt, 1872. (r.)
    Ogivalina mutabilis Canu and Bassler, 1919. (r.)
                                       LOWER MIOCENE FAUNAS.
Lower Miocene (Bowden horizon): Rio Cana, Santo Domingo-
```

```
Lower Miocene (Bowden horizon): Rio Cana, Santo Domingo—
Cupuladria canariensis Busk, 1859. (r.)

Mamillopora tuberosa Canu and Bassler, 1919. (c.)

Metrarabdotos colligatum Canu and Bassler, 1919. (c.)

Thalamoporella biperforata Canu and Bassler, 1919. (c.)

Lower Miocene (Bowden horizon): Rio Gurabo, Santo Domingo—
Cupuladria canariensis Busk, 1859. (c.)

Cupuladria umbellata Defrance, 1823. (c.)

Labiopora miocenica Canu and Bassler, 1919. (r.)

Mamillopora tuberosa Canu and Bassler, 1919. (c.)

Thalamoporella biperforata Canu and Bassler, 1919. (c.)

Thalamoporella granulata Levinsen, 1909. (r.)
```

```
Lower Miocene (Bowden horizon): Santo Domingo-
    Cupuladria biporosa, new species. (r.)
    Laminopora miocenica, new species. (v. r.)
    Palmiecllaria c/r. inermis Jullien. (r.)
    Rhunchozoon curtum, new species, (v. r.)
    Stylopoma magniporosa, new species. (r.)
    Tremogasterina truncatorostris, new species. (v. r.)
Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo-
    Acanthodesia savarti forma monilifera Canu and Bassler, 1919. (r.)
    Adeona porosa Canu and Bassler, 1919. (r.)
    Aimulosia brevis, new species. (r.)
    Corynostylus ellipticus Canu and Bassler, 1919. (r.)
     Corynostylus labiatus Canu and Bassler, 1919. (c.)
     Cupuladria canariensis Busk, 1859. (r.)
     Cupularia umbellata Defrance, 1823. (c.)
     Diaperoecia milneana D'Orbigny, 1839. (r.)
     Hemiseptella lata Canu and Bassler, 1919. (v. r.)
     Hippomenella infratelum Canu and Bassler, 1919. (v. r.)
     Mamillopora tuberosa Canu and Bassler, 1919. (c.)
     Membranipora vaughani Canu and Bassler, 1919. (v. r.)
     Metrarabdotos colligatum Canu and Bassler, 1919. (c.)
     Nellia oculata Busk, 1852. (v. c.)
     Rhynchozoon vaughani Canu and Bassler, 1919. (v. r.)
     Schizopodrella (?) mutabilis Canu and Bassler. 1919. (r.)
     Smittina (?) brevis Canu and Bassler, 1919. (r.)
     Steganoporella parvicella Canu and Bassler, 1919. (c.)
      Thalamoporella biperforata Canu and Bassler, 1919. (c.)
 Lower Miocene (Bowden marl): Bowden, Jamaica—
      Aeanthodesia savarti forma texturata Reuss, 1847. (v. c.)
      Adeona heckeli Reuss, 1847. (r.)
      Aimulosia brevis, new species. (c.)
      Bracebridgia deformis Canu and Bassler, 1919. (r.)
      Callopora dumerillii Savigny-Audouin, 1826. (r.)
      Conopeum lacroirii Busk, 1872. (r.)
      Conopeum ovale, new species. (v. r.)
      Cupuladria canariensis Busk, 1859. (r.)
      Cupularia umbellata Defrance, 1823. (c.)
      Cycloperiella rubra, new species. (v. r.)
      Gemelliporella punctata Canu and Bassler, 1919. (r.)
      Hemiseptella grandicella, new species. (r.)
      Hippodiplosia baccata Canu and Bassler, 1920. (r.)
      Holoporella albirostris Smitt, 1872. (v. c.
      Holoporella hemispherica, new species. (r.)
       Mamillopora tuberosa Canu and Bassler, 1919. (c.)
       Mastigophora granulosa, new species. (v. r.)
       Membranipora osburni, new species. (r.)
       Membranipora tenella Ilincks, 1880. (r.)
       Membrendoeeium parvicapitatum, new species. (r.)
       Metrarabdotos lucrymosum Canu and Bassler, 1919. (c.)
       Rhamphostomella granulosa, new species. (v. r.)
       Rhamphostomella laticella Canu and Bassler, 1919. (v. 1.)
       Rhynchozoon verruculatum Smitt, 1872. (r.)
       Schizopodrella unicornis Johnston, 1847. (r.)
       Smittina ophidiana Waters, 1878. (v. r.)
       Steganoporella parvicella Canu and Bassler, 1919. (c.)
```

```
Lower Miocene (Bowden marl): Bowden, Jamaica—Continued.
     Stephanosella biaperta Michelin, 1841. (r.)
     Stylopoma minuta, new species. (r.)
     Terebripora clongata, new species. (r.)
     Terebripora sincfilum, new species. (r.)
     Thalamoporella biperforata Canu and Bassler, 1919. (c.)
Lower Miocene (Gatun formation): Banana River, Costa Rica-
     Cupuladria canariensis Busk, 1859. (r.)
     Cupularia umbellata Defrance, 1823. (c.)
     Mamillopora tuberosa Canu and Bassler, 1919. (c.)
Lower Miocene (Chipola marl): Chipola River, Calhoun County, Florida-
     Adeonellopsis coccinella, new species. (r.)
     Callopora dumerilii Savigny-Andonin, 1826. (r.)
     Cupularia umbellata Defrance, 1823. (c.)
    Gemelliporella vorax, new species. (r.)
     Leiosella cdax, new species. (r.)
                                        MIOCENE OF MARYLAND.
Miocene (Choptank formation): Cordova, Maryland-
    Acanthodesia savarti forma bifoliata Ulrich and Bassler, 1904. (c.)
     Microporella bifoliata Ulrich and Bassler, 1904. (r.)
Miocene (Choptank formation): Governor Run, Maryland-
    Acanthodesia oblongula Ulrich and Bassler, 1904. (r.)
    Schizopodrella unicornis Johnston, 1847. (r.)
Miocene (Choptank formation): Greensboro, Maryland-
    Holoporella massalis Ulrich and Bassler, 1904.
Miocene (Choptank formation): Pawpaw Point, Maryland-
    Conopeum? nitidulum Ulrich and Bassler, 1904. (r.)
Miocene (Choptank formation): Dover Bridge, Maryland-
    Acanthodesia oblongula Ulrich and Bassler, 1904. (r.)
    Conopeum germanum Ulrich and Bassler, 1904. (v. r.)
    Retepora doverensis Ulrich and Bassler, 1904. (c.)
    Schizopodrella doverensis Ulrich and Bassler, 1904. (r.)
Miocene (Choptank formation): Jones Wharf, Maryland-
    Acanthodesia oblongula Ulrich and Bassler, 1904. (r.)
    Acanthodesia savarti forma bifoliata Ulrich and Bassler, 1904. (c.)
    Chaperia caminosa Ulrich and Bassler. (r.)
    Crisina striatopora Ulrich and Bassler, 1904. (r.)
    Microporella ciliata Linnaeus, 1759. (r.)
    Retepora doverensis Ulrich and Bassler, 1904. (c.)
    Schizoporella cumulata Ulrich and Bassler. (r.)
    Schizoporella latisinuata Ulrich and Bassler, 1904. (r.)
Miocene (Calvert formation): Reed's, Maryland-
    Acanthodesia oblongula Ulrich and Bassler, 1904. (c.)
    Cellepora cribrosa Ulrich and Bassler, 1904. (v. r.)
    Conopeum lacroixii Busk, 1852. (r.)
    Cyclicopora mansfieldi, new species. (v. r.)
    Membranipora fossulifera Ulrich and Bassler, 1904. (v. r.)
    Ogivalina parvula Ulrich and Bassler, 1904. (r.)
    Porella convoluta Ulrich and Bassler, 1904. (r.)
    Porella punetata Ulrich and Bassler, 1904. (v. r.)
Miocene (Calvert formation): Plum Point, Maryland-
    Acanthodesia oblongula Ulrich and Bassler, 1904. (c.)
    Cellepora maculata Ulrich and Bassler, 1904. (c.)
    Holoporella massalis Ulrich and Bassler, 1904. (r.)
```

```
Miocene (Calvert formation): 1 mile south of Parkers Creek, Calvert County, Maryland-
    Callopora parvirostris, new species. (v. r.)
    Cribrilina ligulata, new species. (v. r.)
Miocene (Calvert formation): Chesapeake Beach, Maryland-
    Acanthodesia oblongula Ulrich and Bassler, 1904. (c.)
    Holoporella massalis Ulrich and Bassler, 1904. (r.)
Miocene (St. Mary's formation): St. Mary's River, Maryland-
     Cupularia denticulata Conrad, 1841. (r.)
     Hemiseptella fistula Ulrich and Bassler, 1904. (v. r.)
     Holoporella massalis Ulrich and Bassler, 1904. (r.)
     Lepralia montifera Ulrich and Bassler, 1904. (v. r.)
     Theonoa glomerata Ulrich and Bassler, 1904. (r.)
Miocene (St. Mary's formation): Cove Point, Maryland-
     Amphiblestrum constrictum Ulrich and Bassler, 1904. (r.)
     Conopeum germanum Ulrich and Bassler, 1904. (v. r.)
     Cribrilina punctata Hassall, 1841. (r.)
     Idmonea expansa Ulrich and Bassler, 1904. (r.)
     Membranipora tuberimargo, new species. (v. r.)
     Microporella ciliata Linnaeus, 1759. (r.)
     Porella reversa Ulrich and Bassler, 1904. (r.)
     Ramphonotus agellus Ulrich and Bassler, 1904. (r.)
     Schizoporella cumulata Ulrich and Bassler, 1904. (r.)
                                           MIOCENE OF VIRGINIA.
 Miocene (St. Mary's formation): Bowlers wharf, 18 miles above Urbana, Middlesex County, Virginia-
     Acanthodesia savartii forma typiea. (r.)
     Conopeum lacroixii Busk, 1852. (r.)
      Membraniporina baccata, new species. (r.)
     Ramphonotus agellus Ulrich and Bassler, 1904. (r.)
     Schizopodrella unicornis Johnston, 1847. (r.)
     Spathipora longicauda, new species. (v. r.)
 Miocene (Yorktown formation): Yorktown, Virginia-
      Cellepora maculata Ulrich and Bassler, 1904. (r.)
      Dakaria grandis, new species. (r.)
      Floridina regularis, new species. (r.)
      Gemelliporella vorax, new species. (r.)
      Hippaliosina rostrigera Smitt, 1872. (c.)
      Hippoporella spinosa, new species. (r.)
      Holoporella (?) cchinata, new species. (v. r.)
      Membranipora flabellata Canu, 1906. (r.)
      Microporella fissurifera, new species. (v. r.)
      Ramphonotus asperus, new species. (r.)
      Stylopoma spongites Pallas, 1766. (c.)
      Terebripora parvicella, new species. (r.)
      Tretoeycloecia tortilis Lonsdale, 1845. (c.)
  Miocene (Yorktown formation): 1 mile northeast and 1 mile west of Suffolk, Virginia-
      Acanthodesia savarti forma typica. (r.)
      Atelesopora reptans, new species. (r.)
      Ceriopora virginiana, new species. (v. r.)
      Cyclopericlla rubra, new species.
      Gemelliporella vorax, new species. (r.)
      Hippaliosina rostrigera Smitt, 1872. (c.)
       Membranipora flabellata Canu, 1906. (r.)
       Microporella fissurifera, new species. (r.)
       Ramphonotus asperus, new species. (r.)
```

```
Upper Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia-
     Acanthodesia rectangularia, new species. (v. r.)
      Adeona heckeli Reuss. (c)
     Amphiblesteum constrictum Ulrich and Bassler, 1904. (r.)
     Atelesopora reptans, new species. (r.)
     Berenicea flabellum? Reuss, 1847. (r.)
     Callopora lanccolata, new species. (v. r.)
     Cycloperiella rubra, new species. (c.)
     Dakaria torquata D'Orbigny, 1839. (v. r.)
     Gemelliporella vorax, new species. (r.)
     Hippuliosina rostrigera Smitt, 1872. (c.)
     Hippodiplosia bigibbera, new species. (r.)
     Membraniporina vincularina, new species. (v r.
     Microporella fissurifera, new species. (r.)
     Ramphonotus asperus, new species. (r)
     Stylopoma spongites Pallas, 1766, (c.)
     Tretocycloecia tortilis Lonsdale, 1845. (c.)
 Miocene (Yorktown formation): 1 mile west of Fort Nonsense, Gloucester County, Virginia-
     Atelesopora reptans, new species. (r.)
     Cyclocopora spinifera, new species. (r.)
     Gemelliporella vorax, new species. (r.)
     Microporella fissurifera, new species. (r.)
     Ramphonotus asperus, new species. (r.)
     Schizopodrella unicornis Johnston, 1842. (r.)
     Trypostega venusta Norman, 1864. (r.)
Miocene (Yorktown formation): 1 mile south of Cash, Gloucester County, Virginia-
     Atclesopora reptans, new species. (r.)
     Cyclocolposa spinifera, new species. (r.)
     Ramphonotus asperus, new species. (r.)
Miocene (Yorktown formation): 2 miles southwest of Cash, Gloucester County, Virginia-
     Cyclocol posa spinifera, new species. (r.)
     Cyclopericlla rubra, new species. (c.)
     Dakaria torquata D'Orbigny, 1839. (r.)
     Ramphonotus asperus, new species. (r.)
     Stylopoma spongites Pallas, 7766. (c.)
     Tretocycloccia avellana, new species. (r)
Miocene (Yorktown formation): Near Powcan, King and Queen County, Virginia (loc. No. 8205)—
     Adeona heckeli Reuss, 1847. (r.)
     Berenicea flabellum? Reuss. (r.)
     Cycloperiella rubra, new species. (r.)
     Cyclocolposa spinifera, new species. (r.)
    Gemelliporella vorax, new species. (r.)
Miocene (Yorktown formation): Beulahland, King and Queen County, Virginia (loc. No. 8229)-
    Atelesopora reptans, new species. (r.)
    Berenicca flabellum? Reuss, 1847. (r.)
    Callopora lancrolata, new species. (r.)
    Cyclocolposa spinifera, new species. (r.)
    Cyclopericlla rubra, new species. (r.)
    Ramphonotus asperus, new species. (r.)
    Schizopodrella unicornis Johnston, 1847. (r.)
Miocene (Yorktown formation): One-third mile north of Macedonia Church, Essex County, Virginia
  Toc. No. 8208 -
     Atelesopora reptans, new species. Ir.
     Microporella fissurifera, new species. (v. r.
    Ramphonotus asperus, new species. r.
```

```
Miocene (Yorktown): York River, Virginia-
    Cupularia denticulata Conrad, 1841. (r.)
    Hemiseptella filimargo, new species. (v. r.)
    Holoporella rostrifera, new species. (v. r.)
Miocene (Yorktown formation): Williamsburg, Virginia-
    Cupularia denticulata Conrad, 1841. (r.)
    Tretocycloecia tortilis, Lonsdale, 1845. (c.)
Miocene (Yorktown formation): 11 miles southwest of Reed's Ferry, Virginia-
    Cyclicopora? mansfieldi, new species. (y. r.)
                                      MIOCENE OF NORTH CAROLINA.
Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina-
    Adeona heckeli Reuss, 1847. (r.)
    Aimulosia aculeata, new species. (c.)
    Alderina cesticella, new species. (r.)
    Atelesopora reptans, new species. (r.)
    Cellepora maculata Ulrich and Bassler, 1904. (r.)
    Cupularia denticulata Conrad, 1841. (r.)
    Cupularia umbellata De France.
     Cycloperiella rubra, new species. (c.)
     Floridina minuta, new species. (r.)
     Floridina regularis, new species. (c.)
    Gemelliporella vorax, new species. (c.)
     Hippaliosina rostrigera Smitt, 1872. (c.)
     Puellina radiata-scripta Reuss.
     Schizopodrella unicornis Johnson, 1847.
     Stylopoma spongites Pallas, 1766. (c.)
     Terebripora parvicella, new species. (r.)
     Velumella clongata, new species. (r.)
     Vibracellina pusilla, new species. (r.)
     l'ibracellina simplex, new species. (r.)
 Miocene: Kuhns, Carteret County, North Carolina-
     Lagenipora brevicollis, new species. (r.)
     Membranipora fossulifera Ulrich and Bassler, 1904. (r.)
     Membranipora spiculata, new species. (r.)
     Microporella ciliata Linnaeus, 1759. (r.)
     Porella reversa Ulrich and Bassler ,1904. (r.)
     Pyripora brevicauda, new species.
 Miocene (Duplin marl): Lake Waccamaw, North Carolina-
     Schizopodrella floridina ()sburn, 1914. (r.)
 Miocene (Duplin marl): Snow Hill, Green County, North Carolina-
      Membranipora flabellata Canu, 1906. (r.)
 Miocene (Duplin marl): 2½ miles northwest of Chocowinity, North Carolina-
      Membranipora flabellata Canu, 1906. (r.)
 Miocene (Duplin): 10 miles south of Greenville, North Carolina-
     Cyclopericlla rubra, new species. (c.)
     Microporella fissurifera, new species. (r.)
      Tretocycloecia avellana, new species. (r.)
 Miocene (Duplin marl): 28 miles northwest of Wilmington, North Carolina-
      Holoporella parvula, new species. (r.)
      Holoporella subturrita, new species. (v. r.)
 Miocene (Duplin marl): Wilmington, North Carolina-
      Adeona heckeli Reuss, 1847. (c.)
      Aimulosia aculeata, new species. (r.)
      Callopora dumerilii Savigny-Audouin, 1826. (r.
             12184-23-Bull, 125--2
```

```
Miocene (Duplin marl): Wilmington, North Carolina-Continued.
    Cellepora minuta, new species. (r.)
    Cupularia denticulata Conrad, 1841. (v. c.)
    Cupularia doma D'Orbigny, 1851.
    Cupularia umbellata Defrance, 1823. (c.)
    Cycloperiella rubra, new species. (c.)
    Floridina regularis, new species. (c.)
    Gemelliporella vorax, new species. (c.)
    Hemiseptella rectangulata, new species. (r.)
    Hippoporina gibbosa, new species. (r.)
    Holoporella orbifera, new species. (r.)
    Holoporella subturrita, new species. (r.)
    Lacerna mucronata Smitt, 1872. (v. r.)
    Membrendoecium grande, uew species. (v. r.)
    Proboscina mesleri, new species. (r.)
    Puellina radiata forma scripta Reuss, 1847. (r.)
    Schizopodrella floridina Osburn, 1915. (v. r.)
    Schizopodrella unicornis Johnston, 1847. (c.)
    Smittina trispinosa Johnston, 1838. (c.)
    Stephanosella biaperta Michelin, 1841. (r.)
    Stylopoma spongites Pallas, 1766. (c.)
    Trypostega venusta Norman, 1864. (v. r.)
                             MIOCENE OF SOUTH CAROLINA, FLORIDA, AND CUBA
Miocene (Duplin marl): Muldrows Mills, 5 miles south of Mayville, Sumter County, South Carolina-
    Aimulosia aculeata, new species. (r.)
    Atelesopora reptans, new species. (r.)
    Cellepora maculata Ulrich and Bassler, 1904. (r.)
    Gemelliporella vorax, new species. (r.)
    Schizopodrella unicornis Johnston, 1847. (v. r.)
    Stylopoma spongites Pallas, 1766. (r.)
Miocene (Duplin marl): Darlington, South Carolina-
    Microporella hexagona, new species. (r.)
    Stylopoma spongites Pallas, 1766. (r.)
Miocene: Giles Bluff, Peedee River, South Carolina-
    Microporella tessellata Tuomey and Holmes.
Miocene: Smith's, Goose Creek, South Carolina—
    Hemiseptella lacinia Tuomey and Holmes.
     Tretocycloecia tortilis Lonsdale, 1845.
Miocene: Charleston, South Carolina-
    Hemiseptella granulosa, new species. (v. r.)
Miocene Harvey's Mills, Leon County, Florida-
    Aimulosia aculeata, new species. (r.)
    Cellepora maculata Ulrich and Bassler, 1904. (v. r.)
    Cyclocolposa tenuiparietis, new species. (r.)
     Holoporella orbifera, new species. (r.)
Miocene: Santiago, Cuba-
    Cribrilina cuspidata, new species. (v. r.)
Miocene (Choctawhatchee marl): Jackson's Bluff, Ocklockonee River, Leon County, 25 miles southwest
  of Tallahassee, Florida-
    Acanthodesia savarti forma delicatula Busk, 1859. (r.)
    Amphiblestrum tenuiparietis, new species. (r.)
    Arthropoma cornuta, new species. (r.)
     Cellepora maculata Ulrich and Bassler, 1904. (v. r.)
     Chapcria parvispina, new species. (v. r.)
     Cupuladria canariensis Busk, 1859. (r.)
```

```
Miocene (Choetawhatchee marl): Jaekson's Bluff, Ocklockonee River, Leon County, 25 miles south-
  west of Tallahassee, Florida-Continued.
    Cupularia denticulata Conrad, 1841. (c.)
    Gemelliporella asper, new species. (r.)
    Gemelliporella vorax, new species. (r.)
    Hippoporella? papulifera, new species. (v. r.)
    Hippoparina (?) vestita, new species. (r.)
     Idmonea planula, new species. (v. r.)
     Membraniporidra parca, new species. (v. r.)
     Membraniporina vincularina, new species. (r.)
     Membrendoccium parvicapitatum, new species. (r.)
     Microporella ciliata Linnaeus, 1759. (r.)
     Puellina innominata Couch, 1844. (v. r.)
     Schizopodrella floridina Osburn, 1914. (v. r.)
     Schizopodrella pusilla, new species. (r.)
     Steganoporella magnilabris Busk, 1854. (v. r)
     Tremogasterina horrida, new species. (r.)
     Vibracellina simplex, new species. (v. r.)
                                 PLIOCENE OF SOUTH CAROLINA AND FLORIDA.
 Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina-
     Aimulosia aculeata, new species. (r.)
     Amphiblestrum constrictum Ulrich and Bassler, 1904. (r.)
     Cellepora minuta, new species. (r.)
     Cupularia denticulata Conrad, 1841. (v. c.)
     Cyclocolposa perforata, new species. (v. c.)
     Cyclopericlla rubra, new species. (c.)
     Dakaria parviporosa, new species. (v. r.)
     Electra monostachys Busk, 1875. (r.)
     Floridina parvicella, new species. (v. r.)
     Gemelliporella as per, new species. (v.)
     Gemelliporella vorax, new species. (c.)
     Hemiscptella planulata, new species. (r.)
     Holoporella bicornis, new species. (r.)
     Holoporella orbifera, new species. (c.)
      Membranipora lacroixii Audouin, 1826. (r.)
      Membrendoecium grande, new species. (r.)
      Metrarabdotos auriculatum, new species. (c.)
      Microporella tessellata Tuomey and Holmes, 1857. (r.)
      Puellina crassilabiata, new species. (r.)
     Schizopodrella aculeata, new species. (v. r.)
     Schizopodrella marginata, new species. (r.)
      Smittina trispinosa Johnston, 1838. (r.)
      Spathipora longirima, new species. (r.)
      Tretocycloecia avellana, new species. (v. r.)
 Pliocene (Caloosahatchie marl): Shell Creek, De Soto County, Florida-
      Adeona heckeli Reuss, 1847. (c.)
      Aimulosia brevis, new species. (r.)
      Aimulosia radiata, new species. (v. r.)
      Gemelliporella vorax, new species. (r.)
      Metrarabdotos auriculatum, new species. (v. c.)
      Rhynchozoon verruculatum Smitt, 1872. (r.)
      Schismopora brevincisa, new species. (c.)
      Schizoporella doverensis Ulrich and Bassler, 1904. (v. r.)
      Smittina malcoposita, new species. (v. r.)
```

```
Pliocene (Caloosahatchie marl); Shell Creek, De Soto County, Florida-Continued.
    Smittina trispinosa Johnston, 1838, 1c.
    Stylopoma spongites Pallas, 1766. (c.)
     Vibracellina pusilla, new species. (v. r.)
    Holoporella albirostris Smitt. (v. c.)
Pliocene (Caloosahatchie marl): Monroe County, Florida
    Cupuladria canariensis Busk, 1859. (r.)
    Cupularia denticulata Conrad, 1841. (v. c.)
    Gemelliporella vorax, new species. (c.)
    Hippaliosina rostrigera Smitt, 1872. (r.)
    Holoporella albirostris Smitt, 1872. (v. c.)
     Metrarabdotos auriculatum, new species. (v. c.)
    Schizopodrella unicornis Johnston, 1847. (r.)
    Smittina trispinosa Johnston, 1838. (c.)
    Steganoporella magnilabris Busk, 1854. (v. r.)
    Stylopoma spongites Pallas, 1766. (c.)
                                    PLEISTOCENE OF THE ATLANTIC COAST.
Pleistocene: Simmons Bluff, Yonges Island, Charleston County, South Carolina-
    Hemiseptella tuberosa, new species. (r.)
    Rhynchozoon verruculatum Smitt, 1872. (v. r.)
    Schizopodrella unicornis Johnston, 1847. r.
    Smittina trispinosa Johnston, 1838. (c.)
    Stylopoma spongites Pallas, 1766. (r.)
Pleistocene or Recent: Vero, Florida-
    Acanthodesia savarti forma texturata Reuss, 1847. (r.)
    Acanthodesia savarti forma typica. (c.)
    Smittina maleposita, new species. (r.)
    Smittina trispinosa Johnston, 1838. r.)
    Stephanosella biaperta Michelin, 1842. r.:
    Stylopoma spongites Pallas, 1766. (r.)
Pleistocene: Daytona, Florida-
    Schizopodrella unicornis Johnston, 1847. (r.)
    Smittina trispinosa Johnston, 1838. (r.)
Pleistocene: Los Angeles, California-
    Cellaria diffusa Robertson, 1905. Ir.
    Cellaria mandibulata Hincks, 1882. (v. c.)
Hippothoa hyalina Liunaeus, 1768. (r.)
    Stephanosella biaperta Michelin, 1842. r.)
Pleistocene: Mount Hope, Panama Canal Zone-
    Callopora filum Jullien, 1903. (r.)
    Callopora guernei Jullien, 1903. v. r.
    Cyclicopora multilamellosa, new species. r.)
    Dakaria cherreuxi Jullien, 1903. r.)
    Hippoporella costulata, new species. v. r.
    Hippoporina pusilla, new species. v. r. +
    Holoporella aviculifera, new species. r.)
    Holoporella mucronata, new species. V. T.
    Holoporella turrita Smitt, 1873, (v. r.)
    Mastigophora pesanseris Smitt, 1872. (r.)
    Membranipora osburni, new species. r.;
    Porclla (Palmicellaria) costulata, new species. r.)
    Rhynchozoon (?) levigatum, new species. v.r.
    Schizoporella magniporosa, new species. v. r.
    Stylopoma projecta, new species. r.
    Stylopoma spongites Pallas, 1766. (r.)
```

PLEISTOCENE OF THE PACIFIC COAST.

```
Pleistocene: Santa Barbara, California-
    Callopora crassospina, new species. (r.)
    Callopora horrida Hincks, 1880. (r.)
    Callopora (?) multipora Gabb and Horn, 1862.
    Cauloramphus porosus, new species. (r.)
    Cauloramphus triangularis, new species. (r.)
    Chaperia galeata Busk, 1852. (v. r.)
    Conopeum barbarensis Gabb and Horn, 1862. (r.)
    Crisia serrata Gabb and Horn, 1862. (v. c.)
    ('ystisella ariculifera, new species. (r.)
    Electra monostachys Busk, 1875. (r.)
    Fenestrulina malusi Savigny-Audouin, 1826. (c.)
    Filisparsa clarki, new species. (r.)
    Filisparsa clarki, var. parvula, new variety. (r.)
    Hincksina multispinata, new species. (v. r.)
    Hippothoa hyalina Linnaens, 1768. r.)
    Hippothoa hyalina, var. rugosa, new variety. (v. r.)
    Holoporella umbonata, new species. (r.)
    Idmonea californica D'Orbigny, 1852. (v. c.)
    Lichenopora californica Conrad, 1855. (r.)
    Lichenopora hispida Fleming, 1828. (r.)
     Membraniporina californica Gabb and Horn.
     Metracolposa mucronata, new species. (r.)
     Micropora coriacca Esper, 1794. (r.)
     Microporella californica Hincks, 1882. (c.)
     Microporella ciliata Linnaeus, 1759. (r.)
     Microporella customata Gabb and Horn, 1862.
     Microporella heermani Gabb and Horn, 1862.
     Microporella umbonata Hincks, 1882. (v. r.)
     Microporella vibraculifera Hincks, 1882. (v. r.)
     Phidolopora labiata Gabb and Horn, 1862. (v. c.)
     Poretla collifera Robertson, 1908. (c.)
     Psilosolen capitiferar, new species. (v. c.)
     Puellina heermanni Gabb and Horn, 1862. (v. r.)
    Puellina radiata forma scripta Renss, 1847. (r.)
    Ramphonotus multispinatus, new species. (r.)
    Schismopora abrupta, new species. (r.)
    Schizolavella vulgaris Moll, 1803. (v. r.)
    Schizomarella longirostrata Hincks, 1853. (r.)
    Smittina californicusis Robertson, 1908. (r.)
     Stathmepora flabellata, new species. (c.)
     Tubucellaria punctata Gabb and Horn, 1862. (v. c.)
     Tubucellaria punctata, var. minor, new variety. (r.)
     Tubulipora fasciculifera Hincks, 1884. (r.)
     Tubulipora tuba Gabb and Horn, 1862. (r.)
Pleistocene: Santa Monica, California.
     Callopora circumclathrata Hincks, 1881. (r.)
     Cellaria diffusa Robertson, 1905. (r.)
     Cellaria fissurifera, new species. (r.)
     Costazzia robertsoniae, new species. (r.)
     Crisia serrata Gabb and Horn, 1862. (v. c.)
     Cupularia robertsoniae, new species. (r.)
     Cyclicopora (?) gigantea, new species. (r.)
     Cystisella ariculifera, new species. (r.)
     Diaperoccia flabellata, new species. (r.)
```

```
Pleistocene: Santa Monica, California—Continued.
     Entalophora fasciculifera, new species. (r.)
     Fencstrulina porosa, new species. (r.)
    Hincksina quadrispinosa, new species. (r.)
    Hippothoa hyalina Linnaeus, 1768. (r.)
    Holoporella umbonata, new species. (v. r.)
    Idmonea californica D'Orbigny, 1852. (r.)
    Idmonea dispar, new species. (c.)
    Lepralia cribrosa (?) Maplestone, 1900. (v. r.)
    Lichenopora radiata Savigny-Audouin, 1826. (r.)
    Lichenopora verrucaria Fabricius, 1780. (v. r.)
    Membranipora tuberculata Bose, 1802. (r.)
    Microporella californica Hineks, 1882. (r.)
    Microporella ciliata Linnaeus, 1759. (r.)
    Microporella eustomata Gabb and Horn, 1862. (r.)
    Microporella gibbera, new species. (v. r.)
    Microporella heermanni Gabb and Horn, 1862. (r.)
    Microporella umbonata Hineks, 1882. (r.)
    Microporella vibraculifera Hincks, 1883. (r.)
    Mystriopora (?) areblata, new species. (v. r.)
    Phidolopora labiata Gabb and Horn, 1862. (c.)
    Phidolopora pacifica Robertson, 1908. (r.)
    Phylactella spinosissima, var. major Hincks, 1884. (r.)
    Porella collifera Robertson, 1908. (r.)
    Porella cyclopea, new species. (r.)
    Puellina radiata forma rarccosta Reuss, 1847. (r.)
    Rhynchozoon grandicella, new species. (r.)
    Schismopora lanceolata, new species. (r.)
    Smittina discoidea, new species. (c.)
    Smittina grandicella, new species. (r.)
    Smittina porifera Hineks, 1884. (r.)
    Stathmepora flabellata, new species. (r.)
    Stephanosella biaperta Michelin, 1841. (r.)
    Trypematella papulifera, new species. (r.)
    Tubucellaria punctata Gabb and Horn, 1862. (r.)
    Tubucellaria punctata, var. minor, new variety. (1)
    Tubulipora tuba Gabb and Horn, 1862. (r.)
Pleistocene: Dead Mans Island, off San Pedro, California.-
    Diaperoccia flabellata, new species. (v. r.)
    Entalophora fasciculifera, new species. (v. r.)
    Eurystomella bilabiata Hincks, 1884. (r.)
    Fenestrulina malusi Savigny-Audouin, 1826. (v. r.)
    Hippothoa hyalina Linnaeus, 1768. (r.)
    Idmonea californica D'Orbigny, 1852. (v. c.)
    Idmonea clarki, new species. (r.)
    Lagenipora spinulosa Hincks, 1881. (r.)
    Microporella californica Hineks, 1884. (r.)
    Microporella ciliata Linnaeus, 1759. (r.)
    Microporella umbonata Hincks, 1882. (v. r.)
    Microporella vibraculifera Hincks, 1883. (r.)
    Phidolopora labiata Gabb and Horn, 1862. (r.)
    Phidolopora pacifica Robertson, 1908. (r.)
    Psilosolen capitiferax, new species. (r.)
    Stephanosella biaperta Michelin, 1841. (v. r.)
    Terebripora pacifica, new species (r.) (San Pedro.)
    Tubucellaria punctata Gabb and Horn, 1862. (r.)
    Tubulipora tuba Gabb and Horn, 1862. (r.)
```

SYSTEMATIC DESCRIPTIONS.

Class BRYOZOA Ehrenberg.

Order CTENOSTOMATA Busk.

Family TEREBRIPORIDAE Busk, 1839.

Genus TEREBRIPORA D'Orbigny, 1839.

For description see Bulletin 106, United States National Museum, p. 842.)

TEREBRIPORA PARVICELLA, new species.

Plate 27, figs. 15, 16.

Description.—The canalicules are very thin and branch almost at a right angle. The zooecia are very small, arranged somewhat obliquely on the canalicules.

Occurrence.—Miocene (Yorktown formation): Yorktown, Virginia (rare). Miocene (Duplin marl): Natural Wells, 2 miles southwest of Magnolia, North Carolina (rare).

Holotype.—Cat. No. 68389, U.S.N.M.

TEREBRIPORA SINEFILUM, new species.

Plate 3, figs. 14, 15.

Description.—The canalicules are very thin; they branch obliquely and disappear on the adult zooecia. The young zooecia are small and united among themselves; the old zooecia are large and not united.

Structure.—Our specimens show no zooecia intact and the perforations alone are visible. The cavity containing the cellule alone persists. It is quite remarkable that the canalicules disappear when the cavities are deep. These two sorts of perforations are moreover visible on the same zoarium.

Occurence.—Miocene (Bowden marl); Bowden Jamaica (rare).

Cotypes.—Cat. No. 68390, U.S.N.M.

TEREBRIPORA ELONGATA, new species.

Plate 3, figs. 16, 17.

We know this species only from the perforations left in shells and are therefore not able to give a complete description. The canalicules branch almost at a right angle. The distance between the zooecia is equal to their length. The zooecia are much larger than those of *Terebripora falunica* Fischer, 1866, which is found in Europe at the corresponding geological horizon.

Occurence.—Miocene (Bowden marl): Bowden, Jamaica (rare).

Holotype.—Cat. No. 68391, U.S.N.M.

TEREBRIPORA PACIFICA, new species.

Plate 46, fig. 13.

This species is known only from the perforations left by it, but these are so distinct that it can easily be recognized. The zooecia appear small, pyriform approaching each other from one to two times their length. The ternary axes are

deprived of zooecia but they bear unicellular ramifications. In this character this new species is close to *Terebripora irregularis* D'Orbigny, 1839, but it differs from it in that the distance between the zooecia is much less than five or six times their length.

Occurrence.—Pleistocene: San Pedro, California (rare).

Holotype.—Cat. No. 68392. U.S.N.M.

Genus SPATHIPORA Fischer, 1866.

SPATHIPORA LONGICAUDA, new species.

Plate 27, figs. 12,13.

Description.—The principal axes are linear and do not bear zooccia; they intersect each other at about an angle of 90°. The zooccia are attached to them and are arranged alternately; they are thin, elongated, fusiform and are provided with a long peduncle; the angle of insertion is about 45°.

Affinities.—This species differs from Spathipora scrtum Fischer, 1863, wide spread in the Helvetian faluns of France, in the great length of the peduncle of insertion.

Occurrence.—Miocene (St. Marys formation): 18 miles above Urbana, Middlesex County, Virginia (very rare).

Holotype.—Cat. No. 68393, U.S.N.M.

SPATHIPORA LONGIRIMA, new species.

Plate 47, fig. 3.

Description.—The canalicules are delicate, rectilinear, branching at an angle varying from 45° to 90°. The zooccia are conical, attached directly to the canalicules, without a peduncle. The apertura is terminal, orbicular, with a very long rimule prolonged over almost all the frontal.

Affinities.—This species differs from Spathipora cucullata in which the form is identical, in its very long rimule. It differs from Spathipora longicauda in the absence of a peduncle to the zooccia.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw, River, Horry County South Carolina (rare).

Holotype.—Cat. No. 68394, U.S.N.M.

SPATHIPORA CUCULLATA, new species.

Plate 27, fig. 14.

Description.—The canalicules are delicate, deeply buried in the surface of shells; the ramifications are little oblique. The zooecia are conical, in the form of a cornet of paper; their apertura is at the level of the shell, and their peduncles, attached to a canalicule, are deeply buried and very short.

Affinities.—Of the preceding species, Spathipora longicauda, we know only the perforations left by the zooecia. In the present one, on the contrary, the zooecium is entire because it is entirely embedded in the shell substance. We have no knowledge of the mechanism employed by this species in perforating the shell.

Occurence. Miocene (Yorktown formation): Beulahland, King and Queen County, Virginia (rare).

Holotype.—Cat. No. 68395, U.S.N.M.

Order CHEILOSTOMATA Busk.

Suborder Anasca Levinsen.

Division I. MALACOSTEGA Levinsen

Family ELECTRINIDAE D'Orbigny, 1851.

Genus ELECTRA Lamouroux, 1816.

(For description, see Bulletin 106, U. S. National Museum, p. 76.)

ELECTRA MONOSTACHYS Busk, 1854.

Plate 29, figs. 1-3.

1854. Membranipora monostachys Busk, Catalogue marine Polyzoa in British Museum, pt. 2, Cheilestomata, p. 61, pl. 70, figs. 1-4.

1859. Membranipora monostachys Busk, Monograph fossil Polyzoa of the Crag. Publications Paleontographical Society, London, vol. 14, p. 31, pl. 2, fig. 2.

1880. Membranipora monostachys HINCKS, British Marine Polyzoa, p. 131, pl. 17, figs. 3, 4; pl. 18, figs. 1-4.

1889. Membranipora monostachys Jelly, A Synonymic Catalogue of Marine Bryozoa, p. 155 (Cites bibliography).

1896. Membranipora monostachys Calvet, Bryozoaires: Résultats scientifiques de la Campagne du "Cauden" dans le Golfe de Gascogne, Annales de l'Université de Lyon, vol. 26, p. 253.

1900. Membranipora monostachys Waters. Bryozoa from Franz-Josef Land, Journal Linnean Society London, vol. 28, p. 59, pl. 8, fig. 3.

1904. Membranipora monostachys Calver, Bryozoen: Ergebnisse der Hamberger Magalhaensiche Sammelreise, p. 13.

1911. Membranipora monostachys Guérin-Ganiver, Bryozoaires de l'expedition des Jacques Cartier, Bulletin de l'Institut oceanigraphique, no. 207, p. 9.

1912. Electra monostachys Nordcaard, Revision av Universitetsmuseets samling ar norske Bryozoer, Kgl. norske Videnskabers Selskabs Skriften, 1911, no. 3, pp. 1-19.

1912. Membranipora monostachys Osburn, The Bryozoa of the Woods Hole Region, Bulletin Bureau Fisheries, vol. 30, 1910, p. 227, pl. 22, fig. 29; pl. 39, fig. 87.

1914. Electra monostachys Canu, Contribution a l'etude des Bryozoaires fossiles, Les Bryozoaires du Stampien, XIV, Bulletin Société Géologique de France, ser. 4, vol. 14, p. 147, pl. 4,

Measurements.—Opesia | ho = 0.26-0.34 mm. Zooecia | Lz = 0.40-0.48 mm. | Lz = 0.40-0.26 mm.

Variations.—Our specimens agree with the figures given by Hincks in 1880 and by Busk in 1859. The variety with large gymnocyst noted by Waters in 1900 probably constitutes a distinct species, perhaps identical with Membranipora arcuata Canu, 1908, from the Post-Pampean of Argentina.

This species extends scarcely beyond the Tropics and it has remained fixed in the Frigid and Temperate Zones. Its occurrence in the Phocene of South Carolina indicates a great contraction of the equatorial zone and the position of the Tropic of Cancer toward the thirty-third parallel. Other species confirm this phenomenon.

There are 10 tentacles according to Waters.

Occurrence.-Pliocene (Waccamaw marl): Waccamaw River, Horry County. South Carolina (rare). Pleistocene: Santa Barbara, California (very rare): Waies Bluff, near Cornfield Harbor, St. Marys County, Maryland (rare).

Geological distribution.—Stampian, environs of Paris (Cann); Helvetian of Touraine (Collection Canu); Astian of England (Busk).

Habitat.—Arctic Ocean: Franz Josef Land (16-205 meters) (Waters), Barentz Sea. Eastern Atlantic: English Channel, North Sea off Germany, Denmark, England, Norway (19-24 meters), Gulf of Gascony (17-180 meters), mouth of the Tagus, Cape Verde Islands. Western Atlantic: Woods Hole (3-30 meters), Panama.

This species lives principally at the mouth of streams and large rivers.

Plesiotypes.—Cat. Nos. 68396-68398, U.S.N.M.

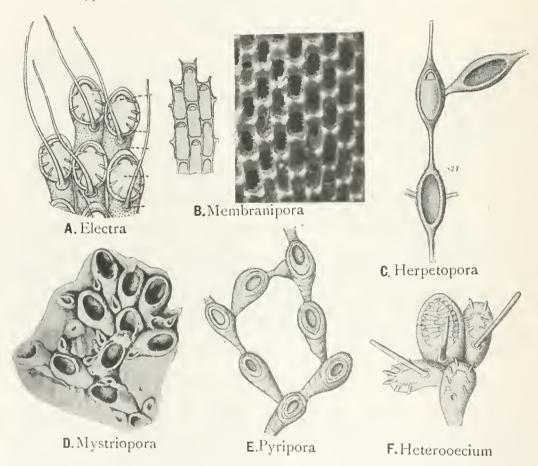


Fig. 1.—Genera of the family Electrinidae D'Orbigny, 1851.

- A. Electra Lamouroux, 1816. E. pilosa Linnaeus, 1758, X 46. Recent.
- B. Membranipora Blainville, 1834. M. membranacca Linnaeus, 1758, X 15 (left) and M. tuberculata Bosc, 1802, X 20. Recent.
 - C. Herpetopora Lang, 1914. H. anglica Lang, 1914, × 27. Cretaceous (after Lang, 1914).
 - D. Mystriopora Lang, 1915. M. möckleri Lang, 1915, × 26. Cretaceous of England.
- E. Pyripora D'Orbigny, 1852. P. catchularia Jameson, 1914, × 25. Recent. (A.E., after Hincks, 1880.)
- F. Heterooecium Hincks, 1892. H. amplectens Hincks, 1892, \times 55. Recent. (After Levinsen, 1909.)

Genus PYRIPORA D'Orbigny, 1852.

(For description see Bulletin 106, U. S. National Museum, p. 78.)

PYRIPORA BREVICAUDA, new species.

Plate 9, figs. 2-4.

Description.—The zoarium encrusts Cellepore bryozoa. The zooecia are pyriform, little narrowed behind, elongated, distinct, separated by a furrow; the gymnocyst is convex and very short. The opesium is elliptical or oval, anterior.

Measurements.3--Opesia $\begin{cases} ho = 0.20 \text{ mm.} \\ lo = 0.12 \text{ mm.} \end{cases}$

Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.22 \text{ mm.} \end{cases}$

Variations.—This small species is of a disconcerting irregularity, especially since the substratum itself is very irregular. The calcified zooecia bear a large elliptical pore. The heterozooecia are numerous and generally fusiform. The zoarial expansions appear to be flabelliform.

Occurrence. - Miocene: Kuhns, Carteret County, North Carolina (rare).

Cotypes.—Cat. No. 68399, U.S.N.M.

Genus MYSTRIOPORA Lang, 1915.

1915. Mystriopora Lang, New Uniserial Cretaceous Cheilostome Polyzoa, Geological Magazine ser. 6, vol. 2, p. 502.

MYSTRIOPORA (?) AREOLATA, new species.

Plate 33, figs. 1, 2.

Description.—The zoarium is incrusting. The zooecia are distinct, separated by a deep furrow, elongated, pyriform; the gymnocyst is short, convex, smooth, much narrowed. The mural rim is thin, garnished all around with hollow spines; the opesium is large, oval, anterior; between the zooecia are some zooeciules whose opesium is also garnished with some spines. The zooecia are separated from each other by rectangular areolar spaces. Dietellae are present.

Measurements.3—Opesia $\begin{cases} ho = 0.45 \text{ mm.} \\ lo = 0.25 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.50 \text{ mm.} \end{cases}$

Affinities.—The Cretaceous genus Mystriopora Lang, 1915, shows also zoo-eciules (although the English authors thought them to be more in the nature of avicularia) and areal spines, but it did not have areolar spaces between the zooecia. Our specimens appear, therefore, to belong to a new genus, although we prefer to employ Lang's name provisionally because the rarity of specimens has not permitted us to make a tangential section in order to discover the dietellae and to establish the nature of the areolar spaces. We have observed cases of double and triple regeneration of the polypide.

This species differs from Membranipora pedunculata Hincks, 1881, also provided with zooeciules, by the absence of spines and by the presence of arcolar spaces.

Occurrence.—Pleistocene: Santa Monica (Long Wharf Canyon), California (rare).

Cotypes.—Cat. No. 68400, U.S.N.M.

In the citation of measurements ho is the length and lo the width of the opesia, Lz and lz similarly the length and width of the zooecia, Lv and lv the same for the vibraculum, Lon and lon for the onychocellaria, ha and la for the apertura, etc.

Genus MEMBRANIPORA Blainville, 1834.

For description see Bulletin 106, U. S. National Museum, p. 77.)

MEMBRANIPORA FLABELLATA Canu, 1904.

Plate 10, figs, 10-14.

1904. Membranipora flabellata Canu. Les Bryozoaires du Patagonien echelle des Bryozoaires pour les terrains tertiares, Memoires Société Géologique de France, Paleontologie, vol. 12, p. 7, pl. 1, fig. 8.

1908. Membranipora flabellata Canu, Iconographie des Bryozoaires fossiles de l'Argentine, Anales del Museo Nacional de Buenos Aires, vol 17, p. 249, pl. 1, figs. 1-5.

$$\label{eq:loss_energy} \begin{array}{lll} \textit{Measurements.} - \text{Opesia} \begin{vmatrix} ho = 0.26 \text{ mm.} \\ lo = 0.16 \text{ mm.} \\ \end{array} \quad \text{Zooecia} \begin{vmatrix} Lz = 0.36 - 0.40 \text{ mm.} \\ lz = 0.26 \text{ mm.} \\ \end{array}$$

Structure.—The zoarium incrusts shells and algae over large surfaces forming many superposed lamellae. The tubercles of the interzooecial angles are well preserved only on the inner lamellae, the outermost lamellae being deprived of them. The zooecia are very deep, the mural rim is quite thick, and the opesium is crenulated. The superior lamellae are formed of zooecia developed around pseudoancestrulae. The latter do not arise from the development of a larva, but they are produced by the gemmation of an inferior zooecium.

Variations.—One of our species from Yorktown, Virginia, is exactly similar to the typical Argentina specimens, but our other examples present variations not noted in the types. The variations are due evidently to the diversity of conditions under which the species existed, as its geographic distribution appears very great.

The tubercles are very inconstant (fig. 10) and entire colonies are deprived of them (fig. 12). The young zooecia have thin mural rims (fig. 12) which gives them an aspect similar to that of Acanthodesia savarti Andouin, 1826, but old zooecia are normal and there are never large zooecia giving rise to new rows. Sometimes the two distal tubercles are joined together (fig. 13). The tubercles are often replaced by interopesial cavities (fig. 11) on the much expanded zoaria, but the presence of normal tuberose zooecia reveals the true nature of these sorts of specimens.

Affinities.—The exterior aspect much recalls Acanthodesia oblongula Ulrich and Bassler, 1904, but the present species differs in its smaller measurements (ho < 0.30 mm.) in the absence of opesial spicules and in its multilamellar colonies.

Occurrence.—Miocene (Yorktown formation): Yorktown, Bellfield and Suffolk, Virginia (rare). Miocene (Duplin marl): 2½ miles northwest of Chocowinity and at Snow Hill, North Carolina (rare).

Geological distribution.—Patagonian, Pampean and Post Pampean of Argentina Canu).

Plesiotype.—Cat. Nos. 68401, 68402, U.S.N.M.

MEMBRANIPORA FOSSI LIFERA Ulrich and Bassler, 1904.

Plate 9, figs. 6, 7,

1904. Membranipora fossulifera Ulrich and Bassler, Bryozoa, Maryland Geological Survey, Miocene, p. 408, pl. 110, fig. 1.

The original description is as follows:

Description.—Zoarium forming a thin expansion upon foreign bodies. Zooecia oblong, subquadrate, sometimes obscurely hexagonal, arranged in regular longitudinal and diagonally intersecting rows, with about 11 in 5 mm., measuring lengthwise, 9 to 10 in 3 mm., diagonally, and 11 to 13 of the longi-

tudinal rows in 3 mm. Opesium elongate oval, generally about twice as long as wide. Walls nearly always a little less than half the width of the opesium, with a median channel, the ring-like elevation enclosing the opesium uniformly elevated except across the anterior end where it is higher and obliquely arched and elevated beneath, probably to form a cover for an opecium, and is usually surmounted by a transverse rib terminating at each end in a small rounded prominence. Rarely the space of an ordinary zooecium is taken up by a cell having a thicker wall and a smaller aperture varying from elongate to nearly circular, while in one instance, a small cell with an oblique opening, narrowed distally, is wedged in between three zooecia.

Measurements.—Opesia
$$\begin{cases} ho = 0.36 \text{ mm.} \\ lo = 0.20 \text{ mm.} \end{cases}$$
 Zooccia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.32 \text{ mm.} \end{cases}$

Variations.—Our micrometric measurements represent the average; in reality as in all the Membranipores the variations are considerable. The mural rim is always somewhat wider in the proximal portion; the separating furrow is deep: the opesium is finely crenulated. The two distal tubercles are placed generally on the termen of the mural rim, but frequently they are attached to the exterior slope and appear there as inserted in the interzooccial angles. We have not yet discovered regenerated zooccia.

We present a new photograph of the type of this species which brings out the surface ornament and especially the tubercles more clearly. The species has resemblance to both *Conopeum* and *Membranipora*, but the apparent absence of the two impressions on the dorsal wall of the zooecium, characteristic of *Conopeum*, and the presence of the spines of *Membranipora* cause us to refer the species to *Membranipora* in its restricted sense.

Occurrence.—Miocene (Calvert formation): Reed's, Maryland (very rare). Miocene: Kuhns, Carteret County, North Carolina (rare).

Plesiotype.—Cat. No. 68403, U.S.N.M.

MEMBRANIPORA SPICULATA, new species.

Plate 9, fig. 1.

Description.—The zoarium encrusts species of Cellepora. The zooccia are distinct, separated by a deep furrow, elliptical, elongated; the mural rim is thin, salient, flat, bearing two distal tuberosities and two pairs of lateral spicules which are conical and very salient. The opesium is of the same form as the zooccium.

$$\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{vmatrix} ho = 0.32 \text{ mm.} \\ lo = 0.18 \text{ mm.} \end{vmatrix} \\ \textit{Zooecia} \begin{vmatrix} Lz = 0.40 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{vmatrix}$$

Affinities.—In its lateral spicules this species has much resemblance to Spiralaria denticulata Busk, 1852, but it differs from it in the absence of an endozooecial ovicell and in the presence of two lateral pairs of spicules. Hincksina serrata MacGillivray, 1881, also has a similar aspect, but in the present species we have not observed either the ovicell or interzooecial avicularia. The spicules are rarely erect; they are more or less curved in the form of a claw. The distal tuberosities are often transformed into spicules. It is therefore rather probable that this species bears an ovicell. But a single specimen has been found.

Occurrence.—Miocene: Kuhns, Carteret County, North Carolina (rare). Holotype.—Cat. No. 68404, U.S.N.M.

MEMBRANIPORA LACROIXII Audouin, 1826.

Plate 29, fig. 4.

1826. Flustra lacroixii Audouin, Explication des planches de Polypes de l'Egypte. . . . In Savigny's Description de L'Egypte, Historie Naturelle, vol. 1, 1809, pt. 3, p. 240, pl. 10, fig. 9 (not Busk, Hincks, Canu).

$$\label{eq:measurements} \begin{split} \textit{Measurements}. &-\text{Ordinary zooeeia: Opesia} \begin{bmatrix} ho = 0.40 \text{ mm.} \\ lo = 0.22 \text{ mm.} \\ zooeeia \end{bmatrix} \\ & \begin{aligned} \text{Zooeeia} \begin{bmatrix} Lz = 0.50 \text{ mm.} \\ lz = 0.32 - 0.34 \text{ mm.} \end{aligned} \end{split}$$

First zooecia of a row:

Opesia
$$|ho = 0.40 \text{ mm.}$$

 $lo = 0.24-0.28 \text{ mm.}$
 $Looecia | Lz = 0.50-0.56 \text{ mm.}$
 $Lz = 0.40 \text{ mm.}$

Variations.—The mural rim is flat and finely striated. There is a constant tubercle at each interzooecial angle. The zooecium giving rise to two or more series is much larger. Our specimen is exactly like Audouin's figure of 1826.

Affinities.—Years ago Waters noted that the Membranipora lacroixi of Busk and Hincks was probably not Audouin's species. Canu, sharing this opinion, has carefully revised the synonymy.⁴ The unexpected discovery in America of Andouin's species confirms therefore the idea of both Waters and Canu. There is no necessity of a new name, since our specimen appears to indicate Membranipora more than Conopeum.

This species differ from Membranipora fossulifera Ulrich and Bassler, 1904, in the presence of tubercles at the interzooecial angles. It is much larger than Biflustra savarti Smitt, 1872, which belongs moreover to another genus.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare).

Habitat.—Red Sea or Mediterranean (Audouin). Plesiotype.—Cat. No. 68405, U.S.N.M.

MEMBRANIPORA TUBERCULATA Bosc, 1802.

Plate 33, figs. 3-5.

1802. Flustra tuberculata Bosc, Histoire naturelle des Vers, vol. 3, p. 118.

1839. Flustra tchuelca D'Orbiony, Voyage dans l'Amerique-Méridionale, vol. 5, pt. 4, Zoophytes, p. 17, pl. 8, fig. 10-14.

1858. Membranipora tuberculata Busk, Zoophytology: On some Madeiran Polyzoa, Quarterly Journal of Microscopical Science, vol. 6, p. 126, pl. 18, fig. 4.

1898. Membranipora tehuelea Waters, Observations on Membraniporidae, Journal of the Linnean Society, Zoology, vol. 26, p. 674, pl. 48, fig. 6-8.

1908. Membranipora tehuelca Robertson, The incrusting Cheilostomatons Bryozoa of the West Coast of North America, University of California Publications, Zoology, vol. 4, no. 5, p. 265, pl. 15, fig. 16, 17; pl. 16, fig. 18.

1909. Membranipora tuberculata Norman, The Polyzoa of Madeira and neighboring Islands, Journal Linnean Society London, Zoology, vol. 30, p. 286.

See North American Early Tertiary Bryozon, p. 89.

1911. Membranipora tehuclea Guerin-Ganivet, Contributions a l'etude des Bryozoaires des cotes Armoricaines II, Bryozoaires provenant de la rade de Brest, et recueilles par les freres Crouan, Travaux scientifiques du Laboratoire de Zoologie de Concarneau, vol. 3, fasc. 5, p. 6, fig. 3.

1912. Membranipora tehuelea Osburn, The Bryozoa of the Woods Hole Region, Bulletin Bureau

Fisheries, vol. 30, 1910, p. 231, pl. 24, fig. 40.

We agree with Norman that this species is indeed that of Bose, who indicated it as very abundant on the algae of the Sargossa Sea. These algae, swept along by the equatorial current, have transported to America a considerable number of European species. This particular one has made the tour of America. One of our fossil specimens still retains its ectocyst and its corneous opercular valve.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (rare).

Habitat.—Eastern Atlantic off France, Senegal, Madeira, and Angola. Western Atlantic: United States, Chagos Isles, Pernambuco, Rio Janeiro, Patagonia. Pacific: Kursachee and California.

Plesiotype.—Cat. No. 68406, U.S.N.M.

MEMBRANIPORA VAUGHANI Cann and Bassler, 1919.

Plate 2, fig. 1.

1919. Membranipora vaughani Canu and Bassler, Geology and paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 77, pl. 2, fig. 1.

Description.—The zoarium incrusts shells. The zooccia are little distinct, united among themselves by their mural rim, elongated, hexagonal; the mural rim is flat, granular, everywhere of equal width. The opesium is elliptical or oval, finely crenulated. Between the zooccia at the angles of junction large rounded tubercles occur.

Measurements.—Opesia
$$box{lo = 0.32 - 0.36 \text{ mm.}}{lo = 0.22 - 0.24 \text{ mm.}}$$
 Zooecia $box{lz = 0.40 - 0.44 \text{ mm.}}{lz = 0.32 \text{ mm.}}$

Affinities.—A single specimen has been found and its zooecia are somewhat deformed by the substratum. The species differs from Membranipora tuberculata Busk, 1859, from the English Crag in its much larger micrometric measurements. It differs from Membranipora tuberculata Bosc, 1802 (not Busk, 1859) in the much more reduced and very different form of its interzooecial tuberosites.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (very rare).

Holotype.—Cat. No. 68407, U.S.N.M.

MEMBRANIPORA(?) TUBERIMARGO, new species.

Plate 9, fig. 12.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a deep furrow, elongated, elliptical. The mural rim is thin, rounded, a little enlarged at the base; it bears one pair of distal and two pairs of lateral tuberosities. The opesium is large, anterior, elliptical.

Variations.—Our micrometric measurements have been made as far as possible from the ancestrula on our single specimen. The opesium of the small ancestrular zooccia measures only 0.30 by 0.20 mm. The number and size of the tubercles is very irregular. They are hollow.

This species differs from Membranipora osburni in its larger dimensions, and in

the presence of two pairs of lateral tuberosities.

Occurrence.—Miocene (St. Mary's formation): Cove Point, Maryland (very rare).

Holotype.—Cat. No. 68408, U.S.N.M.

MEMHRANIPORA OSBURNI, new species.

Plate 46, figs. 11, 12.

Description.—The zoarium incrusts sponges. The zooecia are distinct, separated by a furrow, elongated hexagonal; the mural rim is thin, rounded, very finely crenulated, salient, ornamented with two distal tubercles; the proximal cryptocyst is concave and very small. The opesium is elliptical.

Measurements.—Opesia
$$\begin{cases} ho = 0.30 \text{ mm.} \\ lo = 0.20 \text{ mm.} \end{cases}$$
 Zooecia
$$\begin{cases} Lz = 0.35 \text{ mm.} \\ lz = 0.28 \text{ mm.} \end{cases}$$

Affinities.—The micrometric measurements are smaller than Membranipora vanghani, but the species is quite close to Membranipora tuberculata Busk, 1859, in its two distal tubercles. It is distinguished from the latter by its cryptocyst placed only in the proximal portion of the zooecium, by its larger micrometric measurements (Lz = 0.35 mm. and not 0.30 mm.), and in its much thinner mural rim.

We dedicate this interesting species to Dr. Raymond C. Osburn, of the Ohio State University, in honor of his important researches upon American recent bryozon.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare). Miocene (Bowden marl): Bowden, Jamaica (rare).

Holotype.—Cat. No. 68409, U.S.N.M.

Group MEMBRANIPORAE Canu and Bassler, 1917.

SECTION I. NO OVICELL.

Genus MEMBRANIPORINA Levinsen, 1909.

(For description see Bulletin 106, U. S. National Museum, p. 94.)

MEMIIRANIPORINA TENELLA Hincks, 1880.

Plate 5, figs. 10, 11.

1880. Membranipora tenella Hincks, Contributions History Marine Polyzoa, Foreign Membraniporina II, Annals Magazine Natural History, ser. 5, vol. 6, p. 376, pl. 16, fig. 7.

1889. Membranipora tenella Jelly, Synonymic catalogue of recent marine Bryozoa, p. 167.

1919. Callopora tenella Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa,
Publication of the Carnegie Institution at Washington, No. 291, p. 81, pt. 5, fig. 10.

Affinities. This species is quite easily recognized by its very thin mural rim and especially by its small tubercles arranged more or less symmetrically on the gymnocyst. These tubercles appear to be hollow.

The species may perhaps be confused with Callopora dumerilli Savigny-Audouin, 1826, in which the exterior aspect is quite similar; it differs from it in the presence of nodules instead of avicularia placed on the gymnocyst (and not between the opesia).

Occurrence.—Lower Miocene (Bowden marl): Bowden, Jamaica (rare).

Habitat.—Florida (Hincks).

Plesiotypes.—Cat. No. 68410, U.S.N.M.

MEMBRANIPORINA BACCATA, new species.

Plate 9, fig. 14.

Description.—The zoarium is incrusting, unilamellar. The zooecia are distinct, separated by a deep furrow, much elongated, elliptical or irregular. The mural rim is thin, regular, salient, rounded, covered with large granules. The opesium has the same form as the zooecium.

Measurements.—Opesia $\begin{cases} ho = 0.50 \text{ mm.} \\ lo = 0.22 \text{ mm.} \end{cases}$

Zooecia $\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.35 \text{ mm.} \end{cases}$

Variations.—The zooecia giving rise to new series are much larger; they are numerous, and when the zoarium is not perfectly regular the smaller zooecia which they emit have not sufficient place for development and thus become deformed.

We have noted traces of spicules, so that this species could possibly belong to the genus Acanthodesia. However, it is imprudent to pronounce on this point from a single specimen. The ensemble of the large granules of the mural rims give to this species the aspect of a chain of pearls.

Occurrence.-Miocene (St. Mary's formation): Bowler's wharf, 18 miles above

Urbana, Middlesex County, Virginia (rare).

Holotype.—Cat. No. 68411, U.S.N.M.

MEMBRANIPORINA CALIFORNICA Gabb and Horn, 1862.

Plate 33, fig. 6.

1862. Membranipora californica Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences of Philadelphia, ser. 2, vol. 5, p. 160, pl. 20, fig. 46.

The original description is as follows:

Colony encrusting, cellules arranged in lines, not always in regular quincunx. Cellules pyriform. sometimes continued behind, often abruptly truncated. Opening large, occupying two-thirds or more of the cellules; varying from rounded triangular to oval, usually narrowest in front, never sharply acuminate. Surface sometimes rounded about the oval opening, at others carinate, midway between the aperture and the margin of the cellule, from which line the surface slopes downwards, convexly in both directions. Behind the opening the surface is convex, often narrow. No abortive cellules nor ovarian vescicles were observed.

We have been unable to find specimens of this species and we therefore reproduce the original description and figures for the benefit of future students.

Occurrence.-Pleistocene: Santa Barbara, California.

12184-23-Bull, 125-3

MEMBRANIPORINA VINCULARINA, new species.

Plate 9, figs. S-10.

Description.—The zoarium is free, vincular, formed of two lamellae back to back, composed of four to eight longitudinal rows of zooecia. The zooecia are elliptical, distinct, separated by a deep furrow; the mural rim is wide, rounded, tuberose somewhat wider in the proximal portion; the opesium is regular, elongated, elliptical.

Measurements.—Opesia
$$\begin{cases} ho = 0.24-0.32 \text{ mm.} \\ lo = 0.14-0.16 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.40-0.50 \text{ mm.} \\ lz = 0.30-0.36 \text{ mm.} \end{cases}$

Affinities.—We have not yet discovered an ovicell on any of our specimens. The quadrangular zoaria have much resemblance to Farcimia, but their zooccia are identical with those of the multiserial colonies.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (rare). Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia (very rare).

Cotypes.—Cat. Nos. 68412, 68413, U.S.N.M.

Genus CONOPEUM Norman, 1903.

(For description see Bulletin 106, U.S. National Museum, p. 86.)

CONOPEUM LACROIXII Busk, 1852.

Plate 1, fig. 6.

1919. Conopeum lacroixii Canu and Bassler, Monograph Early Tertiary Bryozoa of North America, Bull. 106, U. S. National Museum, p. 89, pl. 13, fig. 9. (Bibliography, history and distribution.)

$$\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \left\{ \begin{array}{l} \textit{ho} = 0.40 \text{ mm.} \\ \textit{lo} = 0.20 \text{ mm.} \end{array} \right. \quad \text{Zooecia} \left\{ \begin{array}{l} \textit{Lz} = 0.50 \text{ mm.} \\ \textit{lz} = 0.28 \text{ mm.} \end{array} \right.$$

Although we have had the good fortune to discover the true *Membranipora lacroixii* Audouin, 1826 (see p. 22, pl. 29, fig. 4), it is evident that the present species ought henceforth to be classed under Busk's name of 1852. We have no new observations to add to our previous description of this species, for the Miocene specimens are very rare.

Occurrence.—Miocene (St. Mary's formation): Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia (rare). Miocene (Calvert formation): Reeds, Maryland (rare). Miocene (Bowden marl): Bowden, Jamaica (rare).

Plesiotype.—Cat. No. 68414, U.S.N.M.

CONOPEUM OVALE Canu and Bassler, 1919.

Plate 5, fig. 6.

1919. Conopeum ovale Canu and Bassler, Geology and Paleontology of the West Indies Bryozoa, Pub. Carnegie Institution of Washington, No. 291, p. 77, pl. 5, fig. 6.

Description.—The zoarium incrusts shells. The zooccia are distinct, clongated, oval, the point above, separated by a deep furrow; the mural rim is thin, oblique, flat; the opesium is entire, oval. There are two orbicular impressions at the base of the zooccia. The interopesial cavities are irregular and rare.

Measurements.—Opesia
$$\begin{cases} ho = 0.30 \text{ mm.} \\ lo = 0.15 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.35 \text{ mm.} \\ lz = 0.25 \text{ mm.} \end{cases}$

Affinitics.—The primoscrial zooccia are small and calcified. The figured specimen alone has been found. We know no equatorial species with which to compare the present one.

Occurrence.—Lower Miocene (Bowden marl): Bowden, Jamaica (very rare). Holotype.—Cat. No. 68415, U.S.N.M.

CONOPEUM GERMANUM Ulrich and Bassler, 1904.

Plate 10, fig. 8.

1904. Membranipora germana Ulrich and Bassler, Bryozoa: Maryland Geological Survey, Miocene, p. 410, pl. 111, fig. 29.

The original description is as follows:

Zoarium forming a delicate crust upon foreign bodies, the largest seen being less than 1 cm, in diameter. Zooecia shallow, arranged in curved radiating lines in which about 6 occur in 3 mm.; measuring transversely, 11 to 12 of the rows in the same space. Opesia large, more or less elongate-ovate, the length and width usually as 3 is to 2, separated laterally from their neighbors by about half their width, enclosed by a ring-like thickening formed by a furrow separating adjoining zooecia. At somewhat irregular intervals, the interzooecial space widens and is occupied by a rounded cell that may have lodged some kind of avicularium. These cells vary greatly in size but are always considerably smaller than the true zooecia. Occasionally the front margin of the zooecium is more elevated than the rest of the circumference. No ovicells observed.

The description quoted above gives all the essential features of this species.

Occurrence.—Miocene (St. Mary's formation): Cove Point, Maryland (very rare). Miocene (Choptank formation): Dover Bridge, Maryland (very rare). Holotype.—Cat. No. 68416, U.S.N.M.

CONOPEUM? NITIDULUM Ulrich and Bassler, 1904.

Plate 9, fig. 5.

1904. Membranipora nitidula Ulrich and Bassler, Bryozoa: Maryland Geological Survey, Miocene, p. 412, pl. 112, fig. 1.

The original description is as follows:

Zoarium apparently erect, bifoliate. Zooecia oblong, subquadrate, the length twice the width, arranged rather regularly in longitudinal and diagonally intersecting series, rarely four, usually five in three mm. lengthwise, about seven diagonally, and ten or eleven transversely in the same space. Opesia elongate-elliptical, separated transversely by an obtusely ridge-shaped wall generally equalling about half their width; longitudinal interspaces about twice as great as the transverse, medially ridged with a crescentic ovicellar excavation below (that is, in front of each opesium) and usually a small pore-like depression at each end of the ridge. Very minute spines or granules on inner slope of walls.

$$\label{eq:loss} \begin{array}{ll} \textit{Measurements.} - \text{Opesia} \begin{cases} ho = 0.45 \text{ mm.} \\ lo = 0.22 - 0.25 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.25 - 0.30 \text{ mm.} \end{cases}$$

Affinities.—A restudy of the type specimen shows that the original illustration of the species was inverted. At the bottom of each zooccium there is a small flat cryptocyst more or less developed. The interopesial spaces are triangular. The opesium is finely crenulated.

This species is very close to *Conopeum lacroixii* Authors, but it differs in its larger micrometric measurements and in the presence of a small proximal cryptocyst. Nevertheless, as the figured example alone has been found, we can not affirm the constancy of this latter characteristic.

Occurence.—Miocene (Choptank formation): Pawpaw Point, Maryland (rare). Holotype.—Cat. No. 68417, U.S.N.M.

CONOPEUM BARBARENSIS Gabb and Horn, 1862.

Plate 33, figs. 7-9.

1862. Membranipora barbarensis Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences of Philadelphia, ser. 2. p. 60, pl. 20, fig. 47.

Description.—The zoarium incrusts bryozoa. The zooecia are distinct, irregular, elliptical or oval; the mural rim is thin, flat, trenchant; the opesium is large, elliptical or oval. The axis of the avicularium is always in the axis of the proximal zooecium.

Measurements.—Opesia $\begin{cases} ho = 0.32 \text{ mm.} \\ lo = 0.16 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.28 \text{ mm.} \end{cases}$

Affinities.—There are two muscular impressions as in the genus Conopeum. The avicularium (?) is not constant; it only appears from place to place; it is elliptical. We are not certain of its nature for only the figured specimens have been found.

This species differs from *Ellisina* (*Membranipora*) levata Hincks, 1882, from the Queen Charlotte Islands, in its thinner nural rim, and in the avicularium, always placed in the axis of the proximal zooecium (and not in all directions).

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Plesiotypes.—Cat. No. 68418, U.S.N.M.

Genus CUPULADRIA Canu and Bassler, 1919.

1919. Cupuladria Canu and Bassler, The Geology and Paleontology of the West Indies, Bryozoa, Publications Carnegie Institution of Washington, No. 291, p. 77.

No ovicell. The zoarium bears vibracula. No gymnocyst.

Genotype.—Cupuladria (Cupularia) canariensis Busk, 1859. Range: Miocene-Recent.

The genotype does not belong at all to Cupularia as we now understand this genus and as it is defined by its type species. There are neither opesiules nor cryptocyst. Previously, Smitt in 1872 classified the genotype more correctly in Membranipora as this genus was then understood. It can not, however, be maintained in this genus since its significance has been more restricted. We were therefore obliged to create the new genus characterized by the presence of vibracula, although these organs of zoarial adaptation may not logically furnish good generic characters. We only add a letter to the primitive term to modify the long synonymy of this species as little as possible.

This genus differs from *Heliodoma* Calvet, 1907, in the absence of a gymnocyst. The absence of an endozooecial ovicell does not permit it to be confused with either *Vibracellina* Canu and Bassler, 1917, or *Setosellina* Calvet, 1907.

CUPULADRIA CANARIENSIS Busk, 1859.

Plate 1, figs. 7-9.

1859. Cupularia canariensis Busk, Monograph fossil Polyzoa of the Crag, Publications Paleontographical Society, London, vol. 14, p. 87, pl. 13, fig. 2.

1872. Membranipora canariensis Smitt, Floridan Bryozoa, collected by Count L. F. de Pourtales, pt. 1, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 10, No. 11, p. 10.

1908. Cupularia canariensis Canu, Iconographie des Bryozoaires fossiles de L'Argentine, Anales del Museo Nacional de Buenos Aires, vol. 17, p. 275, pl. 5, figs. 8-10 (gives bibliography). 1909. Cupularia guineensis Norman, The Polyzoa of Madeira and neighboring Islands, Journal

Linnean Society London, Zoology, vol. 30, p. 289, pl. 37, figs. 2-6.

1913. Cupularia canariensis Canu, Contributions a l'etude des Bryozoaires fossiles, IV, Pliocene d'Alger, Bulletin Société Géologique de France, ser. 4, vol. 13, pp. 124, 128.

1914. Cupularia guiniensis Osburn, The Bryozoa of the Tortugas Islands, Florida, Publication Carnegie Institution of Washington, No. 182, p. 194.

1916. Cupularia canariensis Canu, Bryozoaires fossiles des Terraines du Sud-Ouest de la France, 9, Aquitanien, Bulletin de la Société Géologique de France, ser. 4 vol. 16, p. 137, pl. 3,

1919. Cupuladria canariensis Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publication Carnegie Institution of Washington, no. 291, p. 78, pl. 1, figs 8-10.

This abundant, well-known fossil and recent species, distinguished by its free, cupuliform zoarium with the zooecia on the convex side having a membraniporoid opesium and vibracula and with the concave side marked by distinct polygonal areas pierced by a few large pores, has been found fossil at number of American localities starting in the Lower Miocene and continuing to the present time. In its geological occurrence it agrees with Cupularia umbellata Defrance, 1823, which likewise begins in the Lower Miocene.

Occurrence.-Lower Miocene (Chipola marl): 1 mile below Baileys Ferry, Chipola River, Florida (rare). Lower Miocene (Bowden horizon): Bowden, Jamaica (rare); Rio Gurabo, Rio Cana, and Cercado de Mao, Santo Domingo (rare); and Port Limon, Costa Rica (common). Lower Miocene (Gatun formation): Banana River, Costa Rica (rare). Miocene (Choctowatchee marl) Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (rare). Pliocene

(Caloosahatchee marl): Monroe County, Florida (rare).

Geological distribution.—Burdigalian of Bordeaux (Canu collection); Helvetian of France (Canu) and Spain (DeAngelis); Tortonian of Austria-Hungary (Reuss) and Italy (Seguenza); Plaisancian of Italy (Manzoni), England (Busk), Spain (DeAngelis) and Algeria (Canu); Astian of Italy (Neviani, Canu); Sicilian of Rhodes (Manzoni) and Italy (Neviani); Quaternary of Italy (Neviani) and Argentina (Canu); Miocene of Australia? (Waters).

Plesiotypes.—Cat. No. 68419-68424, U.S.N.M.

CUPULADRIA BIPOROSA, new species.

Plate 47, figs. 1, 2.

Description.—The zoarium is orbicular. The zooecia are distinct, large, elongated or transverse, large or small, generally rhomboidal; the opesium is regular, oval, variable in form in the wide zooecia. The interzooecial vibraculum is very large and auriculate. On the inner face the zooecia are rectangular and each one bears two large pores.

Affinities.—This interesting new species differs from Cupuladria canaricasis Busk, 1852, in its larger zooecia, which bear on the inner face only two large pores. It is also very well characterized by the presence of wider zooecia in the vicinity of the zoarial margins.

Occurrence.—Miocene (Bowden marl): Santo Domingo.

Cotypes.—Cat. No. 68425, U.S.N.M.

Genus ACANTHODESIA Canu and Bassler, 1919.

1919. Acanthodesia Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, No. 291, p. 79.

No ovicell. The opesium is garnished laterally by small spinous processes and inferiorly by a serrate denticle. Fifteen tentacles.

Genotype.—Acanthodesia (Flustra) savarti Savigny-Audouin, 1826. Range.—Lutetian-Recent.

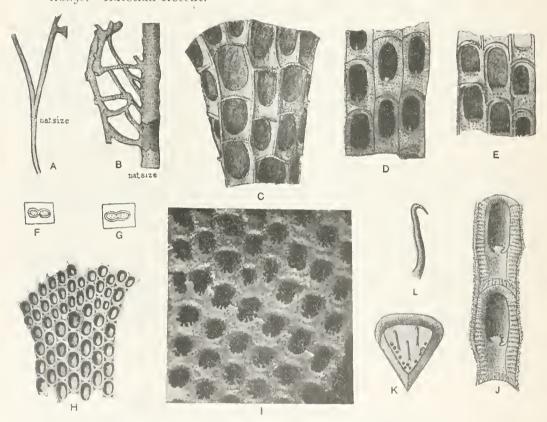


Fig. 2.—Genus Acanthodesia Canu and Bassler, 1919.

Figs. A-I. Acanthodesia savartii Savigny-Audouin, 1826. A and B. Zoaria of recent specimens natural size. C. Young zooccia, \times 25. D. Older zooccia, \times 25, showing small proximal denticles. E. Older zooccia, \times 25, with a serrate denticle directed inward toward the basal wall. F, G. Distal septular plates, \times 250 (A-G, after Waters, 1908). H. Zoarial fragment, \times 12 (after Audouin, 1826). I. A fossil specimen, \times 20, showing zooccia with spicules.

Figs. J-L. Acanthodesia (Mcmbranipora) limosa, Waters, 1908. J. Two zooecia, \times 55. K. Distal wall, \times 55, with the peculiar spine-like processes. L. A spine-like process more highly magnified, \times 200. (Figs. J-L, after Levinsen, 1909.)

ACANTHODESIA SAVARTI Savigny-Audouin, 1826.

1812. Flustra savarti Savieny, Description de l'Egypte, Polypes, pl. 10, fig. 10.

1826. Flustra savarti Audouin, Explication sommaire des planches de Polypes de l'Egypte et de la Syrie, p. 240.

The variations in form and size of this species are extraordinary. We group them around some principal forms, but it is evident that the same zoarium may exhibit all of them. These forms are as follows:

- 1. Forma typica. The opesium is crenulated and elongated; the large zooecia have a cryptocyst. The zoarium is incrusting, unilamellar or bilamellar.
- 2. Forma texturata. The cryptocyst is developed on all the zooceia. No areal spines. No tubercles. The zoarium is unilamellar and subcylindrical.
- 3. Forma *reyti*. The zooecia are irregular. There are areal spines. No cryptocyst. No tubercles. The opesium is denticulated and not crenulated. The zoarium is unilamellar and subcylindrical.
- 4. Forma monilifera. Like typica, but the mural rim is beaded. The zoarium is unilamellar and subcylindrical.
- 5. Forma delicatula. The cryptocyst is short. There are areal spines. The zoarium is bilamellar.
- 6. Forma bifoliata. There are areal spines. The opesium is surrounded by a sort of salient collar. The zoarium is bilamellar. Tubercles at the angles.

These various forms and occurrences are noted under the following headings:

ACANTHODESIA SAVARTI forma TYPICA.

Plate 11, figs. 1-3.

The opesium is crenulated and elongated; the large zooecia have a cryptocyst. The zoarium is encrusting, unilamellar or bilamellar.

Our few American specimens are encrusting. The zooecia are less elongated than in the forma *monilifera* and less spinous than the forma *reyti*. The micrometric variations are very great. The granulations are as beautiful as in the other two forms.

Occurrence.—Miocene (St. Mary's formation): Bowler's wharf, Rappahannock River, 18 miles above Urbana, Middlesex County, Virginia (rare). Miocene (Yorktown formation): Near Suffolk Virginia (rare). Pleistocene: Vero, Florida (common).

Plesiotypes.—Cat. No. 68426, U.S.N.M.

ACANTHODESIA SAVARTI forma REYTI Canu, 1909.

1909. Membranipora reyti Canu, Les Bryozoaires fossiles des Terraines du Sud-Ouest de la France; III Burdigalien, Bulletin de la Société Géologique de France, ser. 4, vol. 9, p. 443, pl. 15, fig. 2.

1909. Membranipora savarti Canu, Les Bryozoaires fossiles des Terraines du Sud-Ouest de la France; III Burdigalien, Bulletin de la Société Géologique de France, ser. 4, vol. 9, p. 444, pl. 15, fig. 3.

Measurements.—Opesia $\begin{cases} \dot{ho} = 0.34 - 0.50 \text{ mm.} \\ lo = 0.20 - 0.30 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$

There are some long and some wide zooecia. The opesium is very finely denticulated and bears areal spines. The mural rim is granulated on the better preserved specimens, which then resemble forma monilifera from Cercado de Mao, Santo Domingo. There are in the Canu collection specimens almost as beautiful as the Santo Domingo form.

Occurrence.—Miocene (Aquitanian): Léognan (LeThil), St. Médard-Gajac Gironde) and St. Avit (Landes), France. Miocene (Burdigalian): Saucats (Le-Peloua), Léognan and Pontiac (Gironde) France.

ACANTHODESIA SAVARTI forma MONILIFERA Canu and Bassler, 1919.

Plate 2, figs. 2, 3.

1919. Acanthodesia savarti forma monilifera Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publication of the Carnegie Institution of Washington, No. 291, p. 79, pl. 2, figs. 2, 3.

Like typica, but the mural rim is beaded. The zoarium is unilamellar and subcylindrical.

This form is intermediate between forma reyti and forma typica. It is evidently the first representative in the American Basin.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (rare.)

Holotype.—Cat. No. 68427, U.S.N.M.

ACANTHODESIA SAVARTI forma TEXTURATA Reuss, 1847.

Plate 5, figs. 1-5; plate 46, figs. 8, 9.

1847. Flustrellaria texturata Reuss, Die fossilen Polyparien des Wiener-Tertiärbeckens, Haidinger's naturwissenschaftliche Abhandlungen, vol. 2, p. 73, pl. 9, fig. 1.

1872. Biflustra savarti Smitt, Floridan Bryozoa, collected by Count L. F. de Pourtales, Part I, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 10, No. 11, p. 20, pl. 4, figs. 92-95.

1877. Flustrellaria texturata Manzoni, I, Briozoi fossili del Miocene d'Austria ed Ungheria, II Parte, Denkschriften der math. natur. Classe der k. Akademie der Wissenschaften, vol. 37, Abtheil. 2, p. 67, pl. 13, fig. 45.

1917. Acanthodesia savarti forma texturata Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publication of the Carnegie Institution of Washington, No. 291, p. 79, pl. 5, figs. 1-5.

The cryptocyst is developed on all the zooecia. No spinous processes. No tubercles. The zoarium is unilamellar and subcylindrical.

Measurements.—Opesia
$$\begin{cases} ho = 0.35 \text{ mm.} \\ lo = 0.20 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.28 \text{ mm.} \end{cases}$

Variations.—The zoarium incrusts fine algae at their bifurcation; it is therefore unilamellar and subcylindrical. The zooecia are elongated, ogival, distinct; the mural rim is striated, salient only in the distal portion; the cryptocyst is large and concave. The opesium is elliptical, very finely denticulated; anteriorly it often bears thin and short spinous processes.

Smitt figured the serrate denticle on the recent specimens; it never persists on the fossil examples. On the inner face the zooecia are rectangular.

Affinities.—The presence of the spinous processes differentiates our specimens from Flustrellaria texturata from the European Tortonian which, according to the figures, does not bear them.

Occurrence.—Lower Miocene (Bowden horizon): Bowden, Jamaica (very com-

mon). Pleistocene or Recent: Vero, Florida (rare).

Geologic distribution.—Tortonian of Austria Hungary (Manzoni).

Habitat.—Recent, off Florida (Smitt).

Plesiotypes.—Cat. Nos. 68428, 68429, U.S.N.M.

ACANTHODESIA SAVARTI forma DELICATULA Bosk, 1859.

Plate 11, figs. 5-9.

1859. Biflustra delicatula Busk, Monograph fossil Polyzoa of the Crag, Publications Paleontographical Society, London, vol. 14, p. 72, pl. 1, figs. 2, 4; pl. 2, fig. 7.

1869. Biflustra delicatula Manzoni, Bryozoi fossili italiani, Terza contribuzione, Sitzungsberichte der kaiserlichen Akademie der Wissenschaften, vol. 60, Abtheil. 1, p. 4, pl. 1, fig. 5.

Our specimens have very narrow fronds. The mural rim is thick and finely tuberous. The cryptocyst is sometimes short, sometimes long. The traces of spinous processes are quite frequent.

The bilamellar form commences in the Helvetian. The variation with large

cryptocyst is abundant in the Mediterranian Pliocene.

Occurrence.—Miocene (Choctawhatchee marl':) Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (rare). Miocene (St. Marys formation): Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia (rare).

Geologic distribution.—Helvetian of St. Avit and Salles (Gironde), of the faluns of Touraine (Canu collection). Pliocene (Astian) of England (Busk) and of Italy (Manzoni).

Plesiotypes.—Cat. Nos. 68430, 68431, U.S.N.M.

ACANTHODESIA SAVARTI forma BIFOLIATA Ulrich and Bassler, 1904.

Plate 11, fig. 4.

1904. Membranipora bifoliata Ulrich and Bassler, Bryozoa, Maryland Geological Survey, Miocene, p. 411, pl. 112, figs. 2, 3, 4.

There are areal spines. The opesium is surrounded by a sort of salient collar. The mural rim is thin. The zoarium is bilamellar. Theoretes at the angles.

This form is much like forma *delicatula* and differs only in the absence of the cryptocyst and in its thin mural rim. The collar-like structure which surrounds the opesium is not constant; it exists sometimes in the unilamellar forms.

Occurrence.-Miocene (Choptank formation): Jones wharf and Cordova, Mary-

land (common).

Cotypes.—Cat. No. 68432, U.S.N.M.

ACANTHODESIA OIILONGULA Ulrich and Bassler, 1904.

Plate 10, figs. 1-3.

1904. Membranipora oblongula Ulrich and Bassler, Bryozoa, Maryland Geological Survey, Miocene, p. 407, pl. 110, figs. 2-5.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a furrow of little depth, clongated, rectangular; the mural rim is rounded, regular, granular, ornamented by two large distal tubercles. The openium is entire, elliptical, finely denticulated and provided with spicules of greater or less length.

$$\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{cases} \textit{ho} = 0.40 \text{ mm.} \\ \textit{lo} = 0.15 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} \textit{Lz} = 0.45 - 0.50 \text{ mm.} \\ \textit{lz} = 0.25 - 0.30 \text{ mm.} \end{cases}$$

Variations.—The figures of Ulrich and Bassler are incomplete as the tubercles have not been restored fully enough. On Plate 10 we reproduce a better photograph of the type (fig. 1). Sometimes the tubercles are joined and form a single large distal tuberosity (fig. 2). The micrometric measurements vary according to the specimens and even on the same specimen they are irregular. The zooecia giving rise to a new series are larger according to the rule in the genus.

Affinities.—This species differs from Membranipora flabellata Canu, 1904, in the presence of spicules and in the larger micrometric measurements. It differs from Acanthodesia savarti Audonin, 1826, in the presence of large distal tubercles.

Occurrence.—Miocene (Calvert formation): Plum Point, Reeds, and Chesapeake Beach, Maryland (common). Miocene (Choptank formation): Jones wharf, Pawpaw Point, Dover Bridge, etc., Maryland (rare).

Plesiotype.—Cat. No. 68433, U.S.N.M.

ACANTHODESIA RECTANGULARIA, new species.

Plate 9, fig. 11.

Description.—The zoarium incrusts oyster shells. The zooecia are distinct, elongated, rectangular; the mural rim is thin, salient; the cryptocyst is deep, short, a little convex. The opesium is large, oval, terminal.

$$\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{cases} ho = 0.35 \text{ mm.} \\ lo = 0.20 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.50 - 0.55 \text{ mm.} \\ lz = 0.30 - 0.35 \text{ mm.} \end{cases}$$

Affinities.—On certain isolated zooecia we have been able to observe some spines and even traces of the serrate proximal denticle. In spite of appearances of our figure, which is incomplete because of the disappearance of the spines, this species belongs really to the genus Acanthodesia. It is well characterized by the rectangular form of its zooecia.

Occurrence.—Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia (rare).

Holotype.—Cat. No. 68434, U.S.N.M.

SECTION II. MEMBRANIPORAE WITH ENDOZOOECIAL OVICELL.

Genus VIBRACELLINA Canu and Bassler, 1917.

(For description see Bulletin 106, U. S. National Museum, p. 110.)

VIBRACELLINA PUSILLA, new species.

Plate 10, figs. 4, 5.

Description.—The zoarium incrusts shells. The zooecia are small, oval, a little elongated, distinct, separated by a deep furrow; the mural rim is thin, rounded, salient, much enlarged at the base into a concave cryptocyst. The opesium is anterior, oval, regular. The ovicell is very small and endozooecial. The vibracula are small, salient, elliptical, auricular.

$$\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{cases} \textit{ho} = 0.14 - 0.16 \text{ mm.} \\ \textit{lo} = 0.10 \text{ mm.} \end{cases}$$
 Zooecia
$$\begin{cases} \textit{Lz} = 0.30 \text{ mm.} \\ \textit{lz} = 0.14 \text{ mm.} \end{cases}$$

Affinities.—According to the rule in this genus, the ancestrular zooccia are smaller and frequently calcified. The ancestrula engenders five normal zooccia and three vibracular ones. In the proximal portion of the zooccia there is often a very short gymnocyst.

This species differs from *Membrendoecium rectum* Canu and Bassler, 1920, from the Vicksburgian in its slightly smaller dimensions, its concave and not flat cryptocyst and in its nonsalient mural rim.

Occurrence.—Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida (very rare). Miocene (Duplin marl): Natural well, 2 miles southwest of Magnolia, North Carolina (rare).

Cotypes.—Cat. Nos. 68435, 68436, U.S.N.M.

VIBRACELLINA SIMPLEX, new species.

Plate 10, figs. 6, 7.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a furrow, elongated, pyriform; the gymnocyst is smooth, convex, rather short; the termen is sharp. The opesium is oval, the point above. The ovicell is very small and endozooecial. The vibracula are rare, very small, auriculated. The ancestrula is calcified and presents a small semilunar aperture.

Measurements.—Opesia
$$\begin{cases} ho = 0.20 \text{ mm.} \\ lo = 0.13 - 0.15 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.35 - 0.40 \text{ mm.} \\ lz = 0.25 \text{ mm.} \end{cases}$

Structure.—The rarety of vibracula give to this species an aspect of great simplicity. The heterozooccia appear to be zooccia in which the development has been arrested by adjacent zooccia. The ancestrula and three ancestrular zooccia are calcified. The hexagonal symmetry in the arrangement of the ancestral zooccia is remarkable. Nevertheless, there are really only five zooccia issuing from the ancestrula according to the rule, and no vibracula. We have observed calcified zooccia and also regenerated zooccia in vicinity of the ancestrula.

This species differs from Vibracellina pusilla in its larger micrometric dimensions and in the very great rarity of the vibracula.

Occurrence.—Miocene (Choctowhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (very rare). Miocene (Duplin marl): Natural well, 2 miles southwest of Magnolia, North Carolina (rare).

Cotypes.—Cat. Nos. 68437, 68438, U.S.N.M.

Genus MEMBRENDOECIUM Canu and Bassler, 1917.

(For description see Bulletin 106, U.S. National Museum, p. 119.)

MEMBRENDOECIUM PARVICAPITATUM, new species.

Plate 12, figs. 1. 2.

Description.—The zoarium encrusts shells. The zooecia are distinct, separated by a deep furrow, clongated, elliptical; the mural rim is thin and rounded. The opesium is of the same form as the zooecium. The ovicell is very small, transverse, endozooecial, scarcely salient. A very small triangular avicularium appears sometimes between the zooecia.

Measurements.—Opesia
$$\begin{cases} ho = 0.36 \text{ mm.} \\ lo = 0.26 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.44 \text{ mm.} \\ lz = 0.32-0.34 \text{ mm.} \end{cases}$

Variations.—The measurements are very irregular. The regeneration of the polypide is very frequent and our figure shows an ovarian zooecium replaced by an ordinary one. The mural rim is very finely granulated. The small avicularium is quite rare. According to the rule in this genus it is deprived of pivot.

All of the resemblances of this species are with Callopora guernei, Jullien, 1904, and it is close to this latter species that we would have to refer the present new species in case additional specimens should show that the small distal elevation of the mural rim is not an endozooccial ovicell.

Occurrence.—Miocene (Choctowhatchee marl); Jackson Bluff, Ocklocknee River 25 miles southwest of Tallahassee, Florida (rare). Miocene (Bowden marl): Bowden, Jamaica (rare).

Holotype and paratype.—Cat. Nos. 68439, 68440, U.S.N.M.

MEMBRENDOECIUM GRANDE, new species.

Plate 11, figs. 10-12.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a furrow of little depth, clongated, large, irregular; the mural rim is thin, bevelled, with a sharp termen. The opesium is large, entire, of the same form as the zooccium. The ovicell is endozooccial, small, little salient, transverse.

Measurements.—Opesia
$$\begin{cases} ho = 0.56 \text{ mm.} \\ lo = 0.40 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.70 \text{ mm.} \\ lz = 0.50 \text{ mm.} \end{cases}$

Affinities.—This species has the aspect of Membranipora irregularis Manzoni, 1875, of the Italian Pliocene, but differs in its larger dimensions (Lz=0.70 and not 0.60 mm.) and in the absence of a distal thickening in the form of a turret. It has also the general aspect of the recent Membranipora irregularis Smitt, 1872; but we have noted (see p. 142 of Bulletin 106. U. S. National Museum) that the latter is a characteristic Alderina; there is therefore a fundamental difference in the ovicell.

Occurrence.—Miocene (Duplin marl): Wilmington. North Carolina (very rare). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare). Recent: Various localities in the Gulf of Mexico.

Cotypes.—Cat. Nos. 68441,68442, U.S.N.M.

Genus OGIVALINA Cann and Bassler, 1917.

(For description see Bulletin 106, U.S. National Museum, p. 117.)

OGIVALINA MUTABILIS Canu and Bassler, 1919.

Plate 1, fig. 3.

1919. Ogivalina mutabilis Canu and Bassler. Geology and Paleontology of the West Indies, Bryozoa, Publication Carnegie Institution of Washington, No. 291, p. 80, pl. 1, fig. 4.

Description.—The zoarium is incrusting. The zooecia are elongated, oval, distinct, separated by a deep furrow; the mural rim is thin, smooth, rounded; there is often a small gymnocyst. The opesium is very large, irregular, more often oval. The ovicell is endozooecial, small, little convex. Sometimes there is a small interzooecial fusiform avicularium (?)

Measurements.—Opesia $\begin{cases} ho = 0.60-0.70 \text{ mm.} \\ lo = 0.30-045 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.75-0.95 \text{ mm.} \\ lz = 0.50-0.70 \text{ mm.} \end{cases}$

The great irregularity of form and zooecial dimensions of this species occasions its name. There are some variations which recall those of *Membranipora irregularis* Manzoni, 1875,⁵ which possesses a mural rim enlarged at the base and also some large interzooecial avicularia.

The present species differs from the splendid Ogivalina eximipora Canu and Bassler, 1917, from the Middle Jacksonian in its smaller dimensions, in the absence of cryptocyst and in the presence of a gymnocyst. The avicularium (?) is identical in form and position.

Occurrence.—Upper Oligocene (Emperador limestone): Old quarry, one-third mile north of west of Empire, Panama Canal zone (rare). Upper Oligocene (Anguilla formation): Southwest side Crocus Bay Hill, Anguilla, Leward Islands (rare).

Holotype.—Cat. No. 68443, U.S.N.M.

OGIVALINA PARVULA Ulrich and Bassler, 1904.

Plate 12, fig. 6.

1904. Membranipora parvula Ulrich and Bassler, Bryozoa, Maryland Geological Survey, Miocene, p. 310, pl. 111, fig. 1, 2.

The original description of this species is as follows:

In its general zoarial and zooecial characters this species resembles Membranipora germana Ulrich and Bassler, 1904 and M. plebeia Gabb and Horn, but it is readily distinguished by the smaller size and less elongate form of its zooecia. The walls are also relatively thicker while the longitudinal arrangement of the zooecia is more pronounced. Measuring longitudinally, 8 zooecia occur in 3 mm, and transversely 12 may be counted in the same space.

To the above we would add that there is a small gymnocyst and that the ovicell is endozooecial and the avicularium is interzooecial.

Occurrence—Miocene (Calvert formation): Reed's Maryland (rare.) Holotype.—Cat. No. 68444, U.S.N.M.

⁵ Briozoi del pliocene antico di Castrocaro Bologna, 1875, p. 10, pl. 1, figs. 5, S.

Genus HINCKSINA Norman, 1903.

(For description, see Bulletin 106, U. S. National Museum, p. 111.)

HINCKSINA QUADRISPINOSA, new species.

Plate 33, fig. 10.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a very deep furrow, large, elongated, elliptical; the mural rim is convex, thin, granulated, ornamented with four distal, hollow spines. The opesium has the same form as the zooccium. The ovicell is endozooccial, very small, scarcely salient.

Measurements.—Opesia $\begin{cases} ho = 0.50 - 0.60 \text{ mm.} \\ lo = 0.24 - 0.26 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz = 0.64 - 0.70 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$

Affinities.—The total regeneration of the polypide is frequent and manifests itself, as is customary by a double mural rim. This species differs from Membranipora doecium grande in the presence of four distal spines. It differs from Membranipora cchinata D'Orbigny, 1839, in the presence of four (and not six) distal spines and in a much smaller ovicell. It differs from Membranipora irregularis D'Orbigny, 1839, of which it has the zooccial form, in its endozooccial and very small ovicell. Our specimen which showed the ovicell was not in a condition to be photographed.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (rare). Holotype.—Cat. No. 68445. U. S. N. M.

HINCKSINA MULTISPINATA, new species.

Plate 33, fig. 11.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a deep furrow, elongated, oval or pyriform, provided with a small convex and smooth gymnocyst; the mural rim is thick, round, salient and bears 16 to 18 large, hollow spines; the opesium is anterior, oval, entire.

Affinities.—The four distal spines are sometimes smaller and more crowded; the others are regularly distributed around the opesium. The figured specimen only having been found, we have not been able to verify whether the ovicell is really endozooecial; a single zooecium shown in our figure appears to have this character. We have observed a case of total regeneration of the polypide.

This species differs from Membranipora variegata Hincks, 1881, in having more than thirteen spines. It differs from Membranipora cchinus Hincks, 1884, in the possession of more than two spines in the distal third of the zooecium. It differs from Membranipora echinata D'Orbigny, 1839, in having more than six spines. It differs from Membranipora pyrula Hincks, 1881, in its smaller dimensions and in the presence of a greater number of spines.

Occurrence.—Pleistocene: Santa Barbara, California (very rare).

Holotype.—Cat. No. 68446, U.S.N.M.

SECTION III. OVICELL HYPERSTOMIAL ALWAYS CLOSED BY THE OPERCULUM.

Genus MEMBRANIPORIDRA Canu and Bassler, 1917.

(For description, see Bulletin 106, U. S. National Museum, p. 133.)

MEMBRANIPORIDRA PARCA, new species.

Plate 12, fig. 7.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a deep furrow, regularly elliptical; the frontal is formed of a convex gymnocyst principally developed in the inferior part of the zooecium. The openium is regularly elliptical and bears a distal indentation in which the operculum is placed. The ovicell is convex, smooth, transverse, placed on the gymnocyst of the distal zooecium; it is always closed by the operculum.

$$\label{eq:measurements} \begin{aligned} & \textit{Measurements.} - \text{Opesia} & \begin{cases} ho = 0.30 \text{ mm.} \\ lo = 0.22 \text{ mm.} \end{cases} \end{aligned} \quad & \text{Zooecia} & \begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases} \end{aligned}$$

Variations.—It is very difficult to interpret the operation of the operculum on the fossils. Here the ovicell is indeed separated from the opesium by the mural rim, but the superior indentation of the opesium which corresponds to the operculum is of exactly the same form as the orifice of the ovicell. It is therefore very probable that our generic assignment is exact.

This species rests directly upon the shell and does not secrete a calcareous dorsal. This eeqnomy of calcite is rather rare in the strata which are not exclusively arenaceous.

Occurrence.—Miocene (Choctowhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (very rare).

Holotype.—Cat. No. 68447, U.S.N.M.

DIVISION IV. OVICELL NEVER CLOSED BY THE OPERCULUM.

Genus ALDERINA Norman, 1903.

(For description, see Bulletin 106, U.S. National Museum, p. 140.)

ALDERINA CESTICELLA, new species.

Plate 12, figs. 4, 5.

Description.—The zoarium incrusts oysters. The zooecia are distinct, clongated, oval, separated by a deep furrow, ornamented by a short gymnocyst; the mural rim is thin; enlarged behind in the form of a cryptocyst, rounded, salient, garnished with six to eight large distal spines; the opesium is anterior, oval, surrounded by a salient and finely wrinkled collar. The ovicell is large, salient, globular, hemispherical, bearing a large, inferior collar, transverse and linear. The ancestrula bears an opesial sinus.

Measurements.—Opesia
$$\begin{cases} ho = 0.20 - 0.25 \text{ mm.} \\ lo = 0.15 - 0.22 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.40 - 0.50 \text{ mm.} \\ lz = 0.25 - 0.30 \text{ mm.} \end{cases}$

Affinities.—This species is irregular in its micrometric measurements, but the zooecial form remains always pyriform. The ancestrula is quite remarkable. It is elliptical and deprived of spines. It engenders two large and three small zooccia. Its opesium bears a deep sinus, the significance of which is unknown. The marginal

zooccia are larger and become almost triangular. Regenerated zooccia are quite frequent. Although the ovicell projects much on the opesium, we think, however, that it does not become closed by the operculum. Moreover, the zooccial form is that of Alderina imbellis Hincks, 1860.

This species differs from *Membranipora galeata* Busk, 1854, in the presence of six to eight distal spines (and not four) and in the absence of a triangular area on the ovicell.

Occurrence.—Miocene (Duplin marl): Natural well, 2 miles southwest of Magnolia, Duplin County, North Carolina (rare).

Holotype.—Cat. No. 68448, U.S.N.M.

Genus CALLOPORA Gray, 1848.

(For description see Bulletin 106, U. S. National Museum, p. 145.)

CALLOPORA DUMERILII Savigny-Audonin, 1826.

Plate 1, fig. 2; plate 2, fig. 23; plate 12, fig. 12.

- 1826. Flustra dumerilii Audouin, Explication Savigny's Egypte, Polypes, pl. 10, fig. 12.
- 1891. Membranipora dumerilii Waters, North Italian Bryozoa, Quarterly Journal Geological Society of London, vol. 47, p. 12, pl. 2, fig. 4.
- 1894. Membranipora dumerilii Levinsen, Mosdyr, Zoologica Danica, Hefte 9, p. 57, pl. 4, figs. 22-25.
- 1907. Membranipora dumerilii Calvet, Bryozoaires Expedition scientifique du Travailleur (1881-82) et du Talisman (1883), p. 385 (bibliography).
- 1909. Callopora dumerilii Norman, On the polyzoa of Madeira and neighboring islands, Linnean Society Journal, Zoology, vol. 30, p. 287.
- 1912. Membranipora dumerilii Canu, Etude des Bryozoaires Helvetiens de l'Egypt, Memoires de l'Institute Egyptien, vol. 6, p. 196, pl. 10, fig. 7 (see for complete bibliography).
- 1919. Callo pora dumerilii, var. lata Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa Publication Carnegie Institution of Washington, No. 291, p. 81, pl. 1, fig. 1.
- 1920. Callopora dumerilii Canu and Bassler, North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 148, pl. 3, fig. 15. (see for zoological and paleontological bibliography).

This species presents two different aspects. Very frequently the zooecium is almost elliptical and there are two symmetrical frontal avicularia (as figured by Waters, Levinsen, and Busk). More rarely the zooecium is oval and there is only a single small interopesial avicularium (as figured by Hincks and Jullien). One specimen from Anguilla has this second aspect which appears to be that of zooecia in the vicinity of the ancestrula (pl 1, fig. 2). In 1919 we separated this second form as a new variety, var. lata, but until more specimens are collected we now prefer to leave the synonymy as above.

We refer doubtfully to this species two specimens incrusting a shell from the lower Miocene of Florida which appear to agree with Busk's figure of 1850 (pl. 12, fig. 12).

American occurrence.—Oligocene (Anguilla formation): Southwest side of Crocus Bay bluffs, uppermost horizon, 125 feet above sea level, Anguilla, Leeward Islands (rare). Jacksonian (Castle Hayne limestone): Wilmington, North Carolina (rare). Miocene (Bowden marl); Bowden Jamaica (rare). Miocene (Chipola marl): Chipola River, Calhoun County, Florida (rare).

Habitat.—Dredged to a depth of 280 meters. Plesiotypes.—Cat. Nos. 68449, 68450, U.S.N.M.

CALLOPORA PARVIROSTRIS, new species.

Plate 12, fig. 3.

Description.—The zoarium incrusts the shell of a Pecten. The zooecia are distinct, separated by a deep furrow, somewhat elongated, ovoid but with irregular contours; the mural rim is salient, thin, granular, somewhat enlarged at the base. The opesium is of the same form as the zooecium. The ovicell is small, very globular, smooth, opening below the operculum by a very small orifice. The interzooecial avicularium is triangular, suberect, very small with two large condyles for pivots.

Measurements.—Opesia bo = 0.45 mm. bo = 0.30 - 0.35 mm. bo = 0.30 - 0.35 mm. bo = 0.45 mm. bo = 0.45 mm. bo = 0.45 mm.

Affinities.—This species belongs to the *C. tenuirostris* group in the presence of its interzooecial avicularium. It differs from the genotype in its much larger micrometric dimensions and in its very small avicularium.

Occurrence.—Miocene (Calvert formation): 1 mile south of Parkers Creek, Calvert County, Maryland (very rare).

Holotype.—Cat. No. 68451, U.S.N.M.

CALLOPORA LANCEOLATA, new species.

Plate 12 fig. 11.

Description.—The zoarium incrusts oysters. The zooecia are distiuct, elongated, fusiform, adjacent to each other by their mural rim; the mural rim is very thin, rounded; a little enlarged at the base; the opesium is elliptical, regular; traces of three pairs of spines are visible. The ovicell is large, salient, rectangular, garnished in front by a large convex area. The avicularium is placed at the base of each opesium; it is large, lanceolate; the beak is very salient and directed toward the base.

 $\label{eq:measurements} \begin{tabular}{l} \textit{Measurements}. \end{tabular} $-$Opesia $$ $| ho = 0.35 \text{ mm.} \\ $| lo = 0.20 - 0.25 \text{ mm.} $$ $Zooecia $$ $| Lz = 0.60 - 0.70 \text{ mm.} \\ $| lz = 0.35 \text{ mm.} $$ $$

Affinities.—Above the ovicell the avicularium is frequently divided into two smaller, symmetrical avicularia.

This species differs from Amphiblestrum constrictum Ulrich and Bassler, 1904, in its much larger avicularia, more elongated zooccia, and in the absence of two prominent condyles for the articulation of the operculum.

Occurrence.—Upper Miocene (Yorktown formation): 3 miles southwest of Petersburg (rare) and Beulahland, King and Queen County, Virginia (rare).

Holotype.—Cat. No. 68452, U.S.N.M.

CALLOPORA CRASSOSPINA, new species.

Plate 33, fig. 13.

Description.—The zoarium incrusts bryozoa. The zooecia are distinct, separated by a furrow, elongated, oval; the mural rim is thin, round, little salient and bears distally six large, hollow spines; the openium is large, oval, entire. The avicularium is interzooecial, long, fusiform, with pointed and salient beak. The ovicell is globular, salient, smooth.

12184-23-Bull. 125----4

 $\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{cases} ho = 0.30 - 0.32 \text{ mm.} \\ lo = 0.20 - 0.24 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Affinities.—Our ovicelled specimen was not in a condition for photography, but it is well preserved and our generic determination is exact. We have observed some calcified and perforated zooccia.

In the number of spines this species much resembles *Membranipora echinata* D'Orbigny, 1839. It differs from it in its oval opesia and in the presence of the interzooceial avicularium. It belongs to the group of *Callopora tenuirostris*.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Holotype.—Cat. No. 68453, U.S.N.M.

CALLOPORA GUERNEl Jullien, 1903.

Plate 45, figs. 3, 4.

1903. Membranipora guernei Jullien, Bryozoaires provenant des Campagnes de l'Hirondelle (1886-1888), Resultats des Campagnes scientifiques du Prince de Monaco, fasc. 23, p. 40, pl. 5, fig. 3.

Affinities.—The micrometric measurements noted on Jullien's figures are a little larger than ours; but as these are drawings and not photographs an error is always possible. Our specimens have the regularly oval zooeeia shown in the figures of the Freneh zoologist. Moreover, the interzooecial avicularia are identical with those described by Calvet in the same publication, "ils sont de forme ovoide a mandibule arrondic, et disposés soit obliquement, soit transversalement par rapport au grand axe de zooécies."

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (very rare).

Habitat.—Gulf of Gascony at 135 meters.

Plesiotypes.—Cat. No. 68454, U.S.N.M.

CALLOPORA FILUM Jullien, 1903,

Plate 45, fig. 5.

1907. Membranipora filum Calvet, Bryozoaires, Expédition Scientifique du Travailleur et du Talisman, p. 386 (bibliography).

The ovicell of this species has not yet been described exactly, but Calvet writes that there are many ovicells which have the peculiarity of being operculated; that they are rather salient, have a semicircular basal contour and are provided with a semicircular orifice placed below the zooceial orifice. We believe that there is, therefore, no doubt in assigning the species to the genus Callopora, although the figure of Smitt, 1872, suggested an endozooceial ovicell. Our specimens are encrusting a coral.

Occurrence.—Pleistocene, Mount Hope, Panama Canal Zone (rare).

Habitat.—Northeast of Moroeco (714 meters), Cape Verde Islands (110-180 meters), Azores (80-318 meters), Corse and Florida (21-97 meters).

Plesiotype.—Cat. No. 68455, U.S.N.M.

CALLOPORA HORRIDA Hincks, 1880.

Plate 33, figs. 18, 19.

1880. Membranipora horrida Hineks, Contributions towards a general history of the marine Polyzoa,
Annals and Magazine of Natural History, ser. 5, vol. 6, p. 82, pl. 10, fig. 6.

1908. Membranipora horrida Robertson, The incrusting Cheilostomatous Bryozoa of the West Coast of North America, University of California Publications, Zoology, vol. 4, No. 5, p. 260, pl. 4, figs. 3, 4.

1898. ? Membranipora californiensis Waters, Observations on Membraniponidae, Journal Linnean Society Zoology, vol. 26, p. 681, pl. 49, fig. 14.

 $\label{eq:measurements} \begin{tabular}{l} \textit{Measurements.} \end{tabular} - \begin{tabular}{l} \textit{Opesia} \\ \textit{lo} = 0.16 - 0.18 \ \mbox{mm}. \end{tabular} \end{tabular} \quad \begin{tabular}{l} \textit{Zooecia} \\ \textit{lz} = 0.30 - 0.40 \ \mbox{mm}. \end{tabular}$

Our specimens have two areal spines, sometimes four. They present micrometric measurements a little different from those noted on Miss Robertson's figure; it is difficult, however, to admit two distinct species, the other characters being very similar. In studying a recent specimen of this species we have been convinced that the orifice of the ovicell is very small and can not be closed by the operculum.

Membranipora californiensis Waters, 1898, presents six articulated areal spines; our form is therefore intermediate between Waters' species and that of Hincks.

This species differs from *Membranipora occultata* Robertson, 1908, in shorter opesium, and in its larger and differently placed avicularium.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Habitat.—Eastern Pacific: Pacific Grove (Robertson) and the Queen Charlotte Islands (Hincks).

Plesiotype.—Cat. No. 68456, U.S.N.M.

CALLOPORA CIRCUMCLATHRATA Hincks, 1881.

Plate 34, figs. 1-3.

1881. Membranipora circumclathrata Hincks, Contribution general history of Marine Polyzoa 7, Foreign Membranipora, Annals and Magazine Natural History, ser. 5, vol. 8, p. 131 (sep. 72), pl. 5, fig. 1.

1908. Membranipora circumclathrata Robertson, The incrusting Cheilostomatous Bryozoa of the West Coast of North America, University of California Publications, Zoology, vol. 4, p. 259, pl. 14, figs. 1, 2.

Structure.—In this species the ovicell is not closed by the operculum. Not only is this quite visible on the fossils but we have been able to verify it on a recent specimen dredged off Santa Monica. Sometimes a triangular pore is visible in the separating furrow of two zooecia; it is clearly attached to one of the mural rims and it is perhaps the base of a pedunculate avicularium as is the genus Cauloramphus. We have not observed these pores on our recent specimens. At the base of each opesium there is a small triangular avicularium with salient beak placed on the median zooecial axis. It sometimes becomes very large and is developed obliquely as in Callopora horrida Hincks, 1880. Nevertheless, the presence of wide-spaced areolar pores differentiates the two species clearly.

Occurrence.—Pleistocene: Santa Monica (Long Wharf Canyon), California (rare).

Habitat.—Pacific coast of California. Plesiotypes.—Cat. No. 68457, U.S.N.M.

CALLOPORA (?) SPECIOSA Gabb and Horn, 1862.

Plate 9, fig. 13.

1862. Membranipora speciosa Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences, Philadelphia, ser. 2, vol. 5, p. 159, pl. 20, fig. 45.

The original description is as follows:

Colony encrusting in irregular patches, composed of elongated, oval cells, often crowded out of their normal shape. Cells arranged in longitudinal lines and in somewhat irregular quincunx, often assuming a transverse arrangement; aperture occupying the whole of the surface, cell walls plain, angular, at their edge or slightly rounded. Interior of the cell, regularly concave, with the sides of the concavity reaching almost to the top of the walls; in new cellules the germinal plate only is seen. This germinal plate often extends for a considerable distance beyond the colony (half an inch) and is marked by irregular longitudinal lines, frequently bent suddenly in an oblique direction and then continued longitudinally as before. Between the cellules are frequently open angular spaces, caused by the inaccurate apposition to the cellules.

Occurrence.—Miocene: Chiriqui, Central America.

CALLOPORA (?) MULTIPORA Gabb and Horn, 1862.

Plate 33, fig. 12.

1862. Siphonella (Flustrellaria) multipora Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences, Philadelphia. ser 2, vol. 5, p. 154, pl. 21, fig. 38.

The original description follows:

Colony free (arranged in a tube in the only specimen we have seen), cellules outside, back, or inside of the tube showing the outline of the cellules. Cellules oval. juxtaposed, placed in irregular quincunx. Opening larger, occupying nearly the whole surface of the cellule; usually of the same shape as the cellule, sometimes having the proximal end wider than the distal end. Cell walls convex or flattened; marked by a variable number of pits often surrounded each by a distinct elevation or wall. The mouth is sometimes constricted by a small rim, parallel with the ordinary wall, placed inside of and below it. This rim is ornamented in the same manner as the larger one.

Occurrence.—Pleistocene: Santa Barbara, California.

Genus AMPHIBLESTRUM Gray, 1848.

(For description see Bulletin 106, U. S. National Museum, p. 156.)

AMPHIBLESTRUM CONSTRICTUM Ulrich and Bassler, 1904.

Plate 13, figs. 1-6.

1904. Amphiblestrum constrictum Ulrich and Bassler, Bryozoa, Maryland Geological Survey, Miocene, p. 413, pl. 115, figs. 6, 7; pl. 118, fig. 15.

The original description is as follows:

Zoarium forming crusts of small extent upon shells, the types growing on a *Pecten*. Zooecia arranged in rather irregular rows, 6 to 8 in 3 mm. Aperture subovate, more or less constricted in front of the midlength, enclosed a sloping and finely striated border, widest posteriorly. Frontal lamina very little developed forming the sloping and transversely striated border just mentioned. Ovicells abundant, large, moderately convex, the middle portion distinguished by being minutely pitted or porous. Avicularia of moderate size, usually one, rarely two to each zooecium, of the same type as in A. flemingi but with the apex more prominent.

This species is closely related to A. flemingi Busk, 1854, and A. trifolium Wood, both living in the seas of to-day and known also as fossils from late Tertiary beds of England and Italy. It is distinguished from both by the lesser development of the frontal lamina. The constriction of the aperture is usually more pronounced in A. constricta.

 $\label{eq:loss_decomposition} \begin{tabular}{l} \textit{Measurements.} \end{tabular} \begin{tabular}{l} \textit{Popesia} \\ \textit{lo} = 0.22 \text{ mm.} \end{tabular} \end{tabular} \begin{tabular}{l} \textit{Zooecia} \\ \textit{lz} = 0.52 - 0.54 \text{ mm.} \\ \textit{lz} = 0.36 \text{ mm.} \end{tabular}$

Variations.—The cryptocyst is very little developed: it is more like a proximal enlargement of the mural rim. By abrasion of the surface, the five dietellae are easily visible. The beak of the avicularium is salient and rounded. The two salient condyles arranged symmetrically in the opesium are intended to support the articulation of the operculum. The ancestrula is orbicular, without spines. It engenders one large and three small zooecia; the two groups are separated by two large zooecia derived from the small ones. We have observed some calcified zooecia perforated by a small median pore. Regenerated zooecia are rare.

Affinities.—In a recent letter, Mr. Waters remarks that the differences between this species and Amphiblestrum flemingi Busk, 1854, are not sufficient to separate the two species. It is evident that we have here a difficult problem in specific determination and that we are not yet able to distinguish the true characters of the species from those of a variety. The problem is further complicated by the great zooecial

variations.

The micrometric measurements taken on the recent specimens and on those from the Helvetian of Touraine and the Tortonian of Eisenstadt, Hungary, are always smaller than those of A. constrictum, a character which may be sufficient to maintain the American species. The American specimens also appear deprived of oral spines. However, the specific name is a small matter if the illustrations are good for careful comparisons.

Occurrence.—Miocene (St. Mary's formation): Cove Point, Maryland (rare). Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia (rare). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina

(rare).

Plesiotypes.—Cat. Nos. 68456, 68459, U.S.N.M.

AMPHIBLESTRUM TENUIPARIETIS, new species.

Plate 13, fig. 7.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a deep furrow, hexagonal, rounded above; the mural rim is very thin, rounded, very finely granulated; the cryptocyst is of little depth, concave, granular; the opesium is oval, narrowed laterally by two condyles serving for articulation of the opercular valve; it bears a concave or undulated proximal border. The avicularium is elliptical, salient, with pointed beak directed below.

engenders three small and two large zooecia.

This species is quite close to Amphiblestrum flemingi Busk, 1854; it differs from it in the absence of gymnocyst, and in the larger ancestrular zooecia. It

appears to be the equatorial equivalent. Imphiblestrum flemingi is a species from the cold and temperate regions; it occurs from the polar circle to the forty-fourth parallel in depths where the temperature is from 3.1° to 6.7° C. Our American species represent, therefore, the simplification which the warmth of the waters may provoke in a species.

Occurrence.-Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee

River, 25 miles southwest of Tallahassee, Florida (rare).

Holotype.—Cat. No. 68460, U.S.N.M.

Genus RAMPHONOTUS Norman, 1894.

(For description, see Bulletin 106, U. S. National Museum, p. 163.)

RAMPHONOTUS ASPERUS, new species.

Plate 13, figs. 11-13.

Description.—The zoarium incrusts shells and masses of Stylopoma spongites. The zooecia are elongated, distinct, oval, much enlarged at the base; the mural rim is thin, much enlarged in the lower part of the cryptocyst, and bears two large areal spines and generally four smaller ones; the opesium is anterior, oval, trifoliated, narrowed laterally by two little salient condyles on which articulates probably the opercular valve. The ovicell is hyperstomial, salient, globular, ornamented by a small transverse area. The avicularium is large, salient, transverse, triangular, with beak pointed and thin and placed regularly below each opesium.

Elongate zooccia:

Measurements.—Opesia
$$\begin{cases} ho = 0.24 \text{ mm.} \\ lo = 0.20 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$ Transverse (short) zooecia:

$$Opesia \begin{cases} ho = 0.18 - 0.20 \text{ mm.} \\ lo = 0.20 \text{ mm.} \end{cases}$$
 Zooecia
$$\begin{cases} Lz = 0.44 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$$

Variations.—The zooecia are often short and wide. The number of spines is variable and may be cight, but there are always two large hollow spines on the ovicelled zooecia. The calcified and perforated zooecia and regenerated zooecia are not rare. All of the avicularia are not large and transverse; they are sometimes much reduced and orbicular, especially in the vicinity of the ancestrula. The walls are fragile and fossilization gives the species quite variable and curious aspects (fig. 12). The dietellae are arranged as in Callopora (fig. 13).

The presence of species of this genus seems to indicate that in the localities

where they lived the waters were calm and little rich in diatoms.

Affinities.—This species differs from Ramphonotus agellus Ulrich and Bassler, 1904, in its triangular avicularium placed transversely. It differs from Ramphonotus rhynchota Busk in its very different dimensions and in the presence of more than two spines.

Occurrence.—Miocene (Yorktown formation): 3 miles southwest of Petersburg, Yorktown, Beulahland, King and Queen County, 1 mile northeast of Suffolk, and other localities in Virginia (rare).

Cotypes.—Cat. No. 68461, U.S.N.M.

RAMPHONOTUS MULTISPINATUS, new species.

Plate 34, fig. 4.

Description.—The zoarium incrusts shells. The zooccia are oval or elliptical, elongated, distinct; separated by a deep furrow; the mural rim is wide, salient, round, and bears four small distal spines, two large hollow spines at the level of the condyles, and a variable number of areal spines; the opesium is anterior, oval, trifoliated. On the gymnocyst there is a large avicularium.

narrow the opesium and which serve for the articulation of the opercular valve.

This species differs from Ramphonotus asperus in its spines, which are more numerous and of two kinds.

Occurrence.—Pleistocene: Santa Barbara, California (rare). Holotype.—Cat. No. 68462, U.S.N.M.

RAMPHONOTUS AGELLUS Ulrich and Bassler, 1904.

Plate 13, figs. 8-10.

1904. Amphiblestrum agellus Ulrich and Bassler, Bryozoa, Maryland, Geological Survey, Miocene, p. 414, pl. 112, figs. 7a, 7b; pl. 118, fig. 14.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a deep furrow, ovoid, much enlarged at their base; the mural rim is thin and sharp; the cryptocyst is short, oblique, concave. The opesium is transverse, trifoliate, much narrowed at the level of two large spines and of the hinge of the opercular valve. The ovicell is quite salient, globular, smooth, formed of two calcareous lamellae the superior one of which is incomplete and limits a small semilunar frontal area. The gymnocyst bears a large very salient oblique avicularium with its beak turned toward the base.

 $\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{cases} ho = 0.20 \text{ mm.} \\ lo = 0.22 - 0.24 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.46 \text{ mm.} \\ lz = 0.30 - 0.46 \text{ mm.} \end{cases}$

Variations.—The original type of the species is incomplete and formed of ancestrular zooecia, which in this genus are always much smaller than the normal zooecia. Other specimens found at the same horizon in Virginia appear to represent the true form of the species. The avicularium is triangular and very salient. On account of its oblique arrangement it is very fragile and is not well preserved in fossilization. The opesium is elongated in the ancestrular zooecia but transverse in the other.

Affinities.—This species differs from Ramphonotus minax Busk, 1864, in its trifoliate opesium and in its avicularium oriented in the zooccial axes. It differs from R, as perus in the presence of two large spines instead of six and in its avicularium not arranged transversely. The worn or broken forms of this species are very difficult to distinguish.

Occurrence.—Miocene (St. Mary's formation): Cove Point, Maryland (rare); Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia (rare).

Plesiotype—Cat. No. 68463, U.S.N.M.

Genus CAULORAMPHUS Norman, 1903.

For description, see Bulletin 106, U. S. National Museum, p. 174.)

CAULORAMPHUS POROSUS, new species.

Plate 33, fig. 17.

Description.—The zoarium incrusts shells and bryozoa. The zooccia are large, distinct, separated by a deep furrow, irregularly elliptical; the mural rim is thin flat, finely striated, very little enlarged at the base; the opesium is large of the same form as the zooccium. Between the mural rims in the separating furrow there are numerous pores (8 to 10) which are bases of pedunculate and articulate avicularia.

Measurements.—Opesia
$$\begin{cases} ho = 0.40-0.44 \text{ mm.} \\ lo = 0.28-0.32 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.44-0.50 \text{ mm.} \end{cases}$

Affinities.—It should be remarked that the pores are not arranged on the mural rim where the termen is sharp. There are therefore no hollow spines; but they form the base, the place of attachment of pedunculate avicularia which surmount them. It is therefore in the genus Cauloramphus that this remarkable species must be classified.

The avicularia being organs of oxygenation we suppose that this species must live in absolutely calm waters little rich in diatoms. The incessant agitation of the mandibles appears to be intended to renew the impure water and to thus carry oxygen and nourishment.

Occurrence.—Pleistocene: Santa Barbara, California (rare). Holotype.—Cat. No. 68464, U.S.N.M.

CAULORAMPHUS TRIANGULARIS, new species.

Plate 33, figs. 14-16.

Description.—The zoarium incrusts shells and bryozoa. The zooecia are distinct, separated by a wide and deep furrow, elongate, elliptical, with a very short proximal gymnocyst; the mural rim is wide, rounded, salient and bears two small distal and eight large areal hollow spines placed in the distal half of the zooecium; the opesium is elliptical or oval according to the form of the zooecium. The ovicell is hyperstomial, globular, ornamented by a semicircular area. In the separating furrow with each zooecium there is a small triangular pore which is the base of the articulation of a pedunculate and articulate avicularium.

Measurements.—Opesia
$$\begin{cases} ho = 0.30-0.34 \text{ mm.} \\ lo = 0.18-0.22 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.48-0.52 \text{ mm.} \\ lz = 0.32-0.40 \text{ mm.} \end{cases}$

Affinities.—The presence of the small exterior avicularium at the line of the spines seems to indicate Membranipora (Cauloramphus) corniculifera Hincks, 1882, from the Queen Charlotte Islands. It differs from it in a smaller number of spines (less than 18) and in the very different arrangement of distal and areal spines. It differs from Cauloramphus spinifcrum Johnston in the larger micrometric measurements (Lz > 0.43 mm.), in the presence of less than 12 spines and in the triangular form of the pore of the pedunculate avicularium. The two species are evidently very close.

The ancestrular zooecia are smaller than the others. The ancestrula is oval. garnished by 12 spines; it engenders three large and three small zooecia.

This is the sixth species with large spines found at Santa Barbara, California. We are ignorant of the true function of these spines, but there is reason to believe that their ensemble forms a trap for diatoms the usual food of bryozoa. The very calm waters off California, therefore, obliged these animals to multiply their means of capture.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Cotypes.—Cat. No. 68465, U.S.N.M.

Family AETEIDAE Smitt, 1867.

Genus AETEA Lamouroux, 1812.

(See Bulletin 106, U. S. National Museum, p. 179, for discussion.)

AETEA ANGUINA? Linnaeus, 1758.

Plate 24, fig. 15.

To this recent species we have referred doubtfully the creeping network of a bryozoan from the Pliocene of South Carolina. Without a more complete zoarium it is impossible to make a more definite identification, but we figure the specimen in order to call attention to fossil bryozoa of this type of structure. The geological distribution and other features of this species have been given in our Monograph on North American Early Tertiary Bryozoa.

Occurrence.-Pliocene (Waccamaw marl): Waccamaw River, Horry County,

South Carolina.

Plesiotype.—Cat. No. 68466, U.S.N.M.

Family CHAPERIIDAE Jullien, 1888.

1888. Chaperiidae Jullien, Mission scientifique du Cap Horn, VI, Zoologie, Bryozoaires, p. 61.

The orifice is semilunar, or subcircular, very large, entirely closed by the operculum, provided interiorly with one or more calcareous plates serving for the insertion of the retractor muscular fibers of the operculum; the frontal is deprived of pores. The ancestrula, of the same form as the zooecia, is oval and bears some articulated spines on the border. (Translated after Jullien.)

Genus CHAPERIA Jullien, 1881 (first group).

1881. Chaperia Jullien, Remarques sur quelques Espèces de Bryozoaires Cheilostomiens, Bulletin Société zoologique de France, vol. 6, p. 61.

Two internal calcareous plates, with extremities fixed and serving for the insertion of the retractor muscular fibers of the operculum. (Translated, after Jullien.)

Type.—Chaperia (Flustra) acanthina Quoy and Gaymard, 1824. Range:

Miocene-Recent.

The species of this genus corresponding exactly to Jullien's definition are: Chaperia (Flustra) acanthina Quoy and Gaymard, 1824; C. (Amphiblestrum) spinosa MacGillivray, 1881; and C. spinosissima Calvet, 1904.

The ovicell of these forms has never been figured. Jullien alone affirmed (but he has given no figure) that the operculum does not close the ovicell. Waters and Calvet classify in the same genus a certain number of other forms with concave

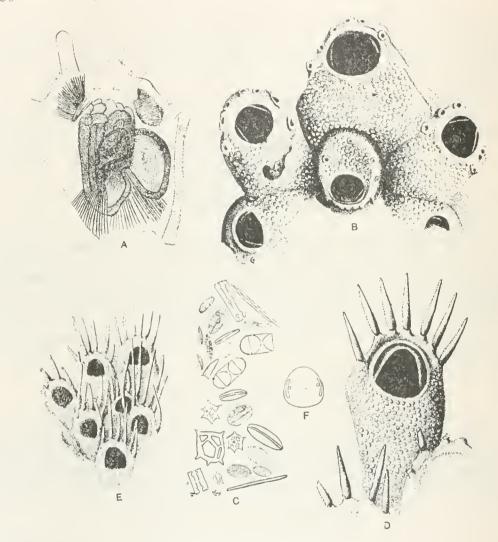


Fig. 3.—Genus Chaperia Jullien, 1881.

Figs. A-G. Chaperia acanthina Quoy and Gaymard, 1824.

A. Interior of a zooecium showing the polypide and the retractor muscles of the operculum. These are the two enormous muscular bundles which characterize the family Chaperiidae, determining in the zooecia the formation of the calcareous lateral plates situated below the orifice. Such plates are the most characteristic remains of this anatomical arrangement which existed as far back as the Cretaceous period.

B. A young colony, × 70, treated with "cau de javelle," showing the ancestrula with the base of the spine of the circumference. In this ancestrula the two lateral funnels which serve for the insertion of the retractor muscles of the operculum, can be seen.

C. Diatoms and radiolaria found in the digestive apparatus of this species, × 216. The radiolarian is Dyctioca speculum Ehrenberg, a species eucountered in many other bryozoa.

D. Very young zooecium hearing spines and in the orifice of which the funnels in process of formation, still unseparated, can be seen.

E. Zooecia covered over by the ectocyst and bearing marginal spines. (Figs. A-E, after Jullien, 1888.)

F. Operculum (after Kirkpatrick, 1890).

frontal and characterized by the same calcareous lamellae. With our present poor state of knowledge of the larva we can not be absolutely certain that this single anatomical peculiarity is sufficient to characterize a family. We think, therefore, that it is preferable to group in a distinct section these species with concave frontal under the same name but placed under the authority of Waters.

The Cretaceous forms figured by D'Orbigny and Hagenow have already been separated under the name of *Hagenowinella* Canu, 1900.

Jullien, who has studied and figured Membranipora galeata Busk, 1852, has not classified it in his genus.

Genus CHAPERIA (Waters, 1898) Levinsen, 1909 (second group).

1909. Chaperia Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa p. 115.

The original description of this genus by Levinsen follows:

The distal part of the zooecinm is furnished internally with two lateral spaces open towards the frontal surface (sometimes coalesced to a single horseshoe shaped one), formed by two plates which project from the side walls and converge towards the distal wall. Each distal wall has two multiporous

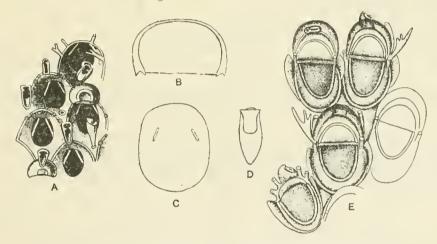


Fig. 4.—Genus Chaperia (Waters, 1898) Levinsen, 1909.

Figs. A-D. Chaperia bilamellata Waters, 1898. A. Specimen, ×25, from Port Elizabeth, South Africa, showing the principal variations of the avicularia, etc. B. Operculum of the orifice of the ovicell, ×85. C. Operculum of the aperture, ×85. D. Mandible of the avicularium, ×85. (Figs. Λ-D, after Waters, 1898.)

Fig. E. Chaperia cervicornis Busk, 1852. Zooecia showing the operculum of the aperture and that of the ovicell in position (after Hincks, 1881).

rosette plates [septular plates] and the distal half of each side wall a single one. Hyperstomial free ooecia with a completely calcified ectooccium [and a special operculum]. The zooecia, which may sometimes have a membraneous opercular valve, sometimes a chitinous compound operculum, are generally strongly provided with spines and have often a well-developed [concave] cryptocyst. Avicularia sometimes trumpet shaped, not always present. The colonies are generally incrusting.

Type.—Chaperia bilamellata Waters, 1898. To this genus belong the following species: Chaperia (Amphiblestrum) cristata Busk, 1884. Chaperia (Membranipora) annulus Manzoni, 1875 (Chaperia galeata Busk, 1852)

Chaperia (Membranipora) cervicornis Busk, 1852.

Chaperia (Electra) cylindracea Busk, 1884.

Chaperia (Membranipora) albispina MacGillivray, 1882.

Chaperia (Amphiblestrum) capensis Busk, 1884.

Chaperia palulosa Waters, 1904.

Chaperia tropica Waters, 1909.

Chaperia galeata Busk, 1852.

Chaperia bilamellata Waters, 1898.

In this list given by Levinsen there is not a single species recognized as identical with the genotype of Jullien. The best known species of this group is *Chaperia galeata* Busk, 1852. The best figures have been drawn from the fossils. The only good figures of the recent specimens have been given by Jullien, 1888, which have not the characteristic lamellae. On the contrary. *Chaperia bilamellata* Waters, 1898, has been perfectly drawn with its chitinous appendages. Levinson classified this genus in the Bicellariidae.

CHAPERIA GALEATA Busk, 1852.

Plate 34, figs. S-10.

- 1852. Membranipora galeata Busk, Catalogue of Marine Polyzoa in British Museum, Cheilostomata, pt. 2, p. 62, pl. 65, fig. 5.
- 1870. Membranipora annulus Manzoni. Briozoi fossili Italiani, Quarta contribuzione, Sitzungsberichte der kaiserlichen Akademie der Wissenschaften, Wien, vol. 61, Abth. 1, p. 7, pl. 1, fig. 6.
- 1875. Membranipora annulus Manzoni. I Briozoi del pliocene antico di Castrocaro, p. 12, pl. 1, fig. 9.
- 1877. Membranipora annulus Manzoni, Bryozoaires du pliocéne supérieur, de l'île de Rhodes, Memoirs de la Société Géologique de France, ser. 3, vol. 1, Mem. No. 2, p. 62.
- 1879. Membranipora galeata Busk, Polyzoa from Kerguelen Island, Philosophical Transactions Royal Society London, vol. 168, p. 195.
- 1879. Membranipora annulus Seguenza, Le formazioni terziarie nella Provincia di Reggio (Calabria), Reale Accademia dei Lincei, Memorie della Classe di Scienze Fisiche, Matematiche e Naturali, ser. 3, vol. 6, pp. 80, 198, 294, 328.
- 1882. Membranipora dentata Waters, Fossil Bryozoa from Mount Gambier, South Australia, Quarterly Journal Geological Society London, vol. 38, p. 263, pl. 8, fig. 14.
- 1884. Membranipora galeata Bush, Report on the Polyzoa collected by II. M. S. Challenger (1873-1876), Pt. I, vol. 10, pt. 30, p. 64.
- 1884. Amphiblestrum cristatum Busk, Report on the Polyzoa collected by H. M. S. Challenger (1873-1876), Pt. I, vol. 10, pt. 30, p. 63, pl. 15, fig. 1.
- 1887. Membranipora annulus Waters, Tertiary Chilostomatous Bryozoa from New Zealand, Quarterly Journal Geological Society, vol. 43, p. 47, pl. 6, figs. 2, 5, 9.
- 1887. Membranipora annulus Peroens, Pliocane Bryozoen von Rhodes, Annalen des k. k. naturhistorischen Hofmuseums, vol. 2, p. 17.
- 1888. Membranipora galcata Jullien, Mission scientifique du Cap Horn, VI, Zoologie, Bryozoaires, p. 75, pl. 5, figs. 6, 8.
- 1891. Membranipora annulus Neviani, Briozoi postpliocenici del sotto suolo di Livorno, Bollettino della Societia Geologica Italiana, vol. 10, p. 116.
- 1895. Amphiblestrum annulus MacGillivray, Monograph of Tertiary Polyzoa of Victoria, Transactions Royal Society Victoria, vel. 4, p. 43, pl. 6, fig. 3.
- 1895. Membranipora galeata Neviani, Briozoi fossili della Farnesina e Monte Mario presso Roma, Palaeontographica Italica, vol. 1, p. 96 sept. 201, no. 10.

1895. Membranipora galcata Neviani, Briozoi neozoici di alcune localita d'Italia, Bollettino della Società Romana per gli Studi Zoologici, vol. 4, pt. 2, p. 233 (sep. 9), 1896; vol. 5, pt. 3, p. 121 (sep. 20).

1896. Membranipora galeata Neviani, Briozoi Postpliocenici di Spilinga (Calabria), Atti Acca-

demia Gioenia di Scienze Naturali in Catania, ser. 4, vol. 9, p. 14.

1898. Membranipora galeata Neviani, Briozoi neozoici di alcune localita d'Italia, Bollettino de la Societa Romana per gli Studi Zoologici, vol. 7, pt. 4, p. 4; pt. 5, pp. 4, 6, 13 (sep.); 1900, pt. 6, p. 66 (sep. 9).

1898. Chaperia annulus Waters, Observations on Membraniporidae, Journal Linnean Society

Zoology, vol. 26, p. 673.

1898. Chaperia annulus, variety bilaminata Waters, Observations on Membraniporidae, Journal Linnean Society, Zoology, vol. 26, p. 673, pl. 47, figs. 5. 8, 9.

1901. Membranipora galcata Neviani, Bryozoi neogenici delle Calabrie, Paleontographia italica, vol. 6, p. 152.

1904. Membranipora galeata Calvet, Bryozoen, Ergebnisse der Hamberger Magalhaensiche Sammelreise, 1892, 1893, vol. 3, p. 10.

1908. Chaperia galeata Canu, Iconographie des Bryozoaires fossiles de l'Argentine, Pt. I, Anales del Museo Nacional de Buenos Aires, vol. 17, p. 262, pl. 3, figs. 13, 14.

1909. Chaperia galeata Calvet, Bryozoaires, Expédition Antarctique Française, Sciences Naturelles, p. 17.

Historical.—Waters cleared up the principal characters of this species; he made known its structure and showed its identity with the fossil Membranipora annulus Manzoni of the Miocene. The complete bibliography which we give above, resulted from his work.

The name galeata is the oldest, but from the description and figure in the British Museum Catalogue identification has not been made, and I should not have recognized it as a synonym without an examination of the Museum specimen. (Waters, 1908.)

However, in 1888 Jullien believed he had rediscovered Busk's species: he gave an excellent figure, but he did not classify it in his genus *Chaperia*. Jullien appeared to be mistaken, since Waters did not cite his work at all in the synonymy. On the other hand, Calvet, 1904, who had studied numerous specimens, rectified the omission of Waters. The fossil specimens of the Canu collection have never the aspect shown in Jullien's figures.

Affinities.—The number of spines varies from four to six. The zooecia in a circle are the ancestrular zooecia. It is remarkable that the genus Chaperia widespread in both hemispheres since the Miocene has emigrated into the Southern

Hemisphere.

We do not believe that the entire synonymy adopted above is exact; there are certainly many species confounded under this name. Even in Italy under the name of *Chaperia annulus* the authors appear to have confused at least two species. Notably our specimens from Farnesina are absolutely distinct from those (and they are quite numerous) which we have collected in the Pleistocene of Palermo. But all these species are so variable that it is very difficult to find constant characters.

Occurrence.—Pleistocene: Santa Barbara, California (very rare). Santa Monica

(Rustic Canyon), California (very rare).

Geological distribution.—Helvetian of Italy (Seguenza); Zanclean of Italy (Seguenza); Entrerrian of Patagonia (Canu); Miocene of Australia (Waters); Pliocene of New Zealand (Waters): Plaisancian of Italy (Manzoni, Neviani);

Astian of Italy (Seguenza, Neviani); Sicilian of Italy (Seguenza, Neviani); Post Pliocene of Italy (Neviani).

Habitat.—Kerguelen Islands (42 meters); Malouines Islands (6 to 16 meters); Bare Orange, Smiths Strait (12 to 27 meters); Straits of Magellan (16 meters); Southern Tierra delle Fuego (11 meters); Port Charcot (40 meters); Booth Wandel Island (30 meters); station 320 of the Challenger, near Montevideo (960 meters). The varieties bilaminata and multifida have been found in the Indian Ocean and at the Cape of Good Hope, the latter at a depth of 720 meters.

Plesiotype.—Cat. No. 68467, U.S.N.M.

CHAPERIA CAMINOSA Ulrich and Bassler, 1904.

Plate 14, figs. 2-4.

1904. Membranipora caminosa Ulrich and Bassler, Bryozoa, Maryland Geological Survey, Miocene, p. 409, pl. 111, figs. 3, 4.

Ulrich and Bassler's original description follows:

Zoarium adnate, forming a thin, single sheet of indefinite extent. Zooecia arranged quincuncially, 6½ measuring longitudinally and 7½ to 8½ diagonally, in 3 mm. Opesia subcircular, separated by one-half to two-thirds their diameter; when the ooecium is wanting, a rim-like border encloses the anterio-half of the opening. Ooecia very high, with a rib across the top, and just in front of the latter a prominent chimney-like tube or hollow spine projecting obliquely over the zooecium next above. When the ooecium is broken away a semiovate or quadrangular concave space is uncovered between the ends of following zooecia. The hollow tubule behind the zooecial aperture is always present, but it is usually worn down so as to appear as merely a thick-rimmed pore. Where the zooecial arrangement is irregular or changed, a second or even a third tubule, each directed forward, may occur between the three zooecia.

This species is a perfectly characterized *Chaperia*. We reproduce figures of the types corrected with more care. Above each zooecium there is a very constant, small, erect, triangular avicularium with its beak above. A large pedunculate avicularium appears in a corner of the cryptocyst; it is almost always broken and appears only as a very irregular pore. There are four large distal articulated spines. We have from the Pleistocene of Palermo (Sicily) a variety of *Chaperia annulus* Manzoni, 1875, very close and which presents also a very constant, small, distal, triangular avicularium. The present species differs from it in the presence of the large, pedunculate avicularium, and its ovicell not smooth and formed of two thick, separable calcareous layers and in the triangular avicularium placed on the ovicell.

Occurrence.—Miocene (Choptank formation): Jones Wharf, Maryland (rare). Cotypes.—Cat. No. 68469, U.S.N.M.

CHAPERIA PARVISPINA, new species.

Plate 14, fig. 1.

The figured specimen is the only one which has been found. It incrusts an oyster shell and is too incomplete to permit an exact description. However, the complete absence of large distal articulated spines is a specific character which amply justifies the establishment of a new species. They are replaced by a variable number of minute and very fragile spines.

Measurements.—Opesia |ho = 0.30 mm. Zooecia |Lz = 0.40 - 0.50 mm. |lz = 0.40 - 0.50 mm.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Oekloeknee River, 25 miles southwest of Tallahassee, Florida (very rare).

Holotype.—Cat. No. 68470, U.S.N.M.

Family FARCIMINARIIDAE Busk, 1852.

Genus NELLIA Busk, 1852.

(For description, see Bulletin 106, U. S. National Museum, p. 195.)

NELLIA OCULATA Busk, 1852.

Plate 2, figs. 5-7.

- 1816. Farcimia tenella Lamarck, Histoire naturelle des animaux sans vertèbres, vol. 2, pl. 2, figs. 26, 27.
- 1816. Cellaria tenella Lamarck, Histoire naturelle des animaux sans vertèbres, ed. 1, p. 135.
- 1834. Cellaria tenella Blainville, Manuel d'Actinologie ou de Zoophytologie, p. 455.
- 1850-1852. Cellaria tenella D'Orbigny, Paleontologie Française: Terrain Crétace, vol. 5, p. 28.
- 1852. Nellia oculata Busk, Catalogue Marine Polyzoa, British Museum, Pt. I, Cheilostomata, p. 18, pl. 64, fig. 6; pl. 65, fig. 4.
- 1873. Nellia oculata Smrtt, Floridau Bryozoa, Pt. II: Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 10, no. 11, p. 3, pl. 1, figs. 53, 54.
- 1880. Nellia oculata MacGillivray in McCoy's Prodomus of the Zoology of Victoria, vol. 1, decade 5, p. 51, pl. 49, fig. 5.
- 1881. Nellia oculata Haswell, On some Polyzoa from the Queensland coast, Proceedings Linnean Society New South Wales, vol. 5, p. 36.
- 1884. Nellia oculata Busk, Report on Polyzoa collected by the Challenger, vol. 10, pt. 30, p. 27.
- 1887. Nellia oculata Hincks, On the Polyzoa and Hydroida of the Mergui Archipelago, Journal Linnean Society, Zoology, vol. 21, p. 121.
- 1887. Farcimia oculata WATERS, Bryozoa from New South Wales, etc., Annals and Magazine Natural History, ser. 5, vol. 20, p. 92.
- 1909. Nellia tenella Levinsen, Morphologic and Systematic Studies on the Cheilostomatous Bryozoa, p. 120, pl. 1, fig. 13 (ovicell).
- 1919. Nellia oculata Canu and Bassler, Goology and Paleontology of the West Indies, Bryozoa, Publication Carnegie Institution of Washington, No. 291, p. 82, pl. 2, figs. 5-7.
- 1920. Nellia oculata Canu and Bassler, Monograph North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 196, pl. 82, figs. 6-10.

Small fragments of this delicate bryozoan occurring in the Bowden horizon of Santo Domingo are quite sufficient for the determination of the species in the American Miocene. The dimensions of the opesium are not equal on the four sides, one of which is often larger than the others. The authors are not in accord as to the name which should be given to this species. We have adopted that of Busk. The species is discussed on page 196 of our monograph on North American Early Tertiary Bryozoa, where a more detailed bibliography is given the above references being additional ones.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo

Domingo (rare).

Geological distribution.—Vicksburgian of Mississippi; Lutetian of the environs of Paris (Canu); Helvetian of Egypt (Canu); Miocene of Australia (Waters).

Plesiotypes.—Cat. No. 68471, U.S.N.M.

Family OPESIULIDAE Jullien, 1888.

Genus FLORIDINA Jullien, 1881.

(For description, see Bulletin 106, U.S. National Museum, p. 219.)

FLORIDINA PYRIPORA Canu and Bassler, 1919.

Plate 1, fig. 1.

1919. Floridina pyripora Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa. Publications Carnegie Institution of Washington, No. 291, p. 82, pl. 2, fig. 2.

The few specimens of this incrusting species which have been collected in the Leeward Islands seem hardly sufficient for the description of a new Floridina, because polymorphism is the rule in this genus; but we believe more specimens will prove this to be a good species. The polypidian convexity is little salient; the opesiules are large and poorly defined; the lateral apophyses are very little salient; the general aspect of the opesium is pyriform. The ovicell is endozooecial, smooth, and salient.

Measurements.— Opesia (including opesiules) $\begin{cases} ho = 0.20 \text{ mm.} \\ lo = 0.22 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.48 - 0.50 \text{ mm.} \\ lz = 0.40 - 0.44 \text{ mm.} \end{cases}$

Floridina antiqua Smitt, 1872, a recent species in the Gulf of Mexico, is marked by large lateral denticles and by small opesiules, giving it a quite different aspect.

Occurrence.—Oligocene (Antigua formation): Rifle Butts, Antigua, Leeward Islands (rare). Oligocene (Anguilla formation): Southwest side of Crocus Bay, Anguilla, Leeward Islands (rare).

Holotype.—Cat. No. 68472, 68473, U.S.N.M.

FLORIDINA FUSIFERA Canu and Bassler, 1919.

Plate 1, fig. 5.

1919. Floridina fusifera CANU and BASSLER, Geology and Paleontology of the West Indies, Bryozoa. Publications Carnegie Institution of Washington, No. 291, p. 83, pl. 1, fig. 3.

Description.—The zoarium is incrusting. The zooecia are hexagonal; the cryptocyst is larger than the opesium; it is of little depth and is finely granulated: the opesium is trifoliate; the lateral apophyses are very salient; the opesiules are large and rounded; the polypidian convexity is very salient and convex. The onychocellarium is large, somewhat constricted laterally, and is provided with a very large orifice. Here and there are small fusiform avicularia.

Measurements.—Opesia $box{loc} = 0.20 \text{ mm}$. Zooecia $box{lz} = 0.50 \text{ mm}$. Onychocellaria Opesium = 0.30 by 0.20 mm. Zooecium = 0.60 by 0.36 mm.

There are only four zooecia intact on the single zoarium obtained. presence of small fusiform avicularia has appeared to us sufficient to justify the creation of this species, this feature never having been observed in the genus.

Occurrence.-Oligocene (Antigua formation): Rifle Butts, Antigua, Leeward Islands (very rare).

Holotype.—Cat. No. 68474, U.S.N.M.

FLORIDINA REGULARIS, new species.

Plate 14, fig. 7, 8.

Description.—The zoarium incrusts shells and consists of one or two superposed lamellae. The zooecia are distinct, separated by a furrow, little elongated, hexagonal, regular; the cryptocyst is smooth, somewhat concave, limited laterally by two convergent grooves which end at the opesiules. The apertura is orbicular and forms the distal part of the trifoliate opesium; the polypidian convexity is limited laterally by two linear opesiules. The ovicell is little salient and endozooecial. The onychocellarium is fusiform; its beak is very salient on the zooecial plane; its opesium is anterior.

Measurements (maximum).—Opesia
$$\begin{cases} ho = 0.15 \text{ mm.} \\ lo = 0.12 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.35 \text{ mm.} \end{cases}$

Affinities.—The marginal zooecia are elongated; the central zooecia are transverse.

This species differs from *Floridina antiqua* Smitt, 1872, in its onychocellarium with salient beak of a different form. It differs from *Floridina parvicella* in its larger micrometric measurements.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (common). Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina (common). Miocene (Yorktown formation): Yorktown, Virginia (rare).

Holotype and paratype.—Cat. Nos. 68475, 68476, U.S.N.M.

FLORIDINA PARVICELLA, new species.

Plate 31, fig. 12.

Description.—The zoarium incrusts oysters. The zooecia are small, hexagonal, little elongated, distinct but with adjacent mural rims; the opesium is trifoliate; the anterior part of aperture is elongate or elliptical; the polypidian convexity is wide and limits two linear and transverse opesiules.

Measurements.—Opesia
$$\begin{cases} ho = 0.10 \text{ mm.} \\ lo = 0.07 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.38 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Affinities.—This species is sufficiently characterized by its small dimensions. It differs again from Floridina regularis in the absence of a furrow separating the zooecia.

The genus *Floridina* has been observed in the Cretaceous formations of northern Europe. It has, therefore, in the course of the geologic ages migrated slowly toward the Equator. The Waccamaw marl is its last known stage in America.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (very rare).

Holotype.—Cat. No. 68477, U.S.N.M.

FLORIDINA MINIMA, new species.

Plate 14, fig. 6.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a furrow, very small, hexagonal or ogival; the mural rim is wide, convex, little salient, very finely granulated; the cryptocyst is of little depth, somewhat convex

and finely granulated. The opesium is elliptical, elongated; the opesiular indentations are large, rectilinear, oblique, limited by a concave proximal border. The ancestrula is very small.

Measurements.—Opesia
$$|ho=0.12 \text{ mm.}|$$
 $|ho=0.12 \text{ mm.}|$ $|lo=0.07 \text{ mm.}|$ $|lo=0.30 \text{ mm.}|$

Affinities.—On our type specimen we have observed neither onychocellaria nor ovicells. This is the smallest species of *Floridina* known. It differs from *Floridina* parvicella in its zooecia separated by a deep furrow and in its smaller opesial dimensions.

Occurrence.—Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina (rare).

Holotype.—Cat. No. 68478, U.S.N.M.

Genus VELUMELLA Canu and Bassler, 1917.

(For description, see Bulletin 106, U.S. National Museum, p. 213.)

VELUMELLA ELONGATA, new species.

Plate 14, fig. 5.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a furrow, large, elongated, ogival; the mural rim is thin, rounded, granulated, salient; the cryptocyst is shallow, flat, granulated. The opesium is large, semilunar; the opesiular indentations are of little depth, rounded, almost symmetrical. The onychocellarium is large, elongated, straight, acuminate or elliptical; its opesium is median, elongated, elliptical.

Affinities.—In its general aspect, the relations of its dimensions and its frontal granulations, this species much resembles Vincularia abyssicola Hincks, 1881 (not Smitt, 1872), from Singapore or the Philippines. It differs from it simply in the absence of lateral facets to the cryptocyst; that is to say, in the peculiarity characterizing Smittipora. The onychocellaria are not fusiform as in Rectonychocella and Diplopholeos.

Occurrence.—Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, North Carolina (rare).

Holotype.—Cat. No. 68479, U.S.N.M.

Genus MICROPORA Gray, 1848.

(For description, see Bulletin 106, U. S. National Museum, p. 234.)

MICROPORA CORIACEA Esper, 1794.

1862. Reptescharellina disparilis Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences Philadelphia, ser. 2, vol. 5, p. 147, pl. 20, fig. 29.

1912. Micropora coriacca Barroso, Briozoos de la Estacion de Biologia maritima de Santander, Instituto nacional de Ciencias Físico naturales, p. 21. 1917. Micropora coriacea Canu, Bryozoaires fossiles des Terrains du Sud-Ouest de la France, Bulletin de la Société Géologique de France, ser. 4, vol. 16, p. 135 (cites hibliography).

1920. Micropora coriacea CANU and BASSLER, Monograph Early Tertiary Bryozoa of North America, Bulletin 106, U. S. National Museum, p. 235, pl. 4, figs. 20-22.

The only known occurrence in American post Oligocene strata of this wellknown recent and fossil bryozoan is in the Pleistocene rocks of California, where it received the name of Reptescharellina disparilis Gabb and Horn, 1862. The species is discussed on page 235 of our volume on the North American Early Tertiary Bryozoa, where a more complete bibliography is given.

Occurrence.—Midwayan, Jacksonian, and Vicksburgian of the United States

Pleistocene: Santa Barbara, California (rare).

Plesiotype.—Cat. No. 68480, U.S.N.M.

Genus SELENARIA Busk, 1854.

1854. Selenaria Busk, Catalogue marine Polyzoa, pt. 2, p. 101.

The ovicell is endozooecial; it appears on the surface of the colony as low, rounded, pent-roof shaped swellings. The cryptocyst is perforated by the opesiules or limited by the opesiular indentations. The opesium is irregular. The opercular valve is limited by the distal part of the zooecial mural rim. Porous vibracula are disseminated among the zooccia. The zoarium is discoid and cupuliform; its inner surface is perforated by numerous pores. No spines.

Genotype.—Selenaria maculata Busk, 1854.

Range.—Claibornian to Recent.

According to Levinsen, who wrote in 1909:

The vibracula are an arched frontal surface perforated by numerous pores or by slits. A high ribl onshaped lamina issuing from the one lateral margin in the distal part of the vibracularian chamber stretches over toward the opposite margin and not far from this bends inward toward the hasal surface. It serves no doubt for the attachment of the flagellum. Distal wall with two multiporous septules, and the distal half of each lateral wall with a single one. Lateral walls are common to the contiguous neighboring zooecia.

SELENARIA AURICULARIA, new species.

Description.—The zoarium is a Lunulites form of 6 mm. in diameter. The zooecia are distinct, separated by a salient and wide mural rim, hexagonal, regular or transverse; the cryptocyst is deep, flat, smooth, and perforated by two large equal opesiules symmetrically arranged. The apertura is elliptical and transverse. The vibracula are very large, auriform, terminated by a short, hooked, small canal. The inner side is perforated by numerous pores regularly arranged on the inner layer and very irregularly disposed on the outer central layer.

Variations.—The zooecia are very irregular in size and we have not been able to discover any constant micrometric measurements. The apertura is likewise irregular in its measurements. The vibracula are larger at the periphery than at the

center. The opesiules alone are of the same diameter on all the zooccia.

Affinities.—The form of the opesiules is quite variable in the genus Selenaria. They are perforated (as in our American species) in S. parvipunctata Maplestone, 1904; S. bimorphocella Maplestone, 1904; and S. magnipunctata Maplestone, 1904. They are formed by deep lateral indentations in S. maculata Busk, 1862. They

are transformed into simple, lateral, irregular sinuosities in S. cupola Tenison Woods, 1879; S. concinna Tenison Woods, 1879; S. marginata Tenison Woods, 1879; and S. otwayensis Maplestone, 1904. They are very little visible in S. nitida Maplestone, 1909, and S. flagellifera Maplestone, 1910. The inner face of all the species is porous except in S. marginata Tenison Woods, 1879.

Our American species differs from the known forms only in the auriculate form of vibracula. This difference can not form a generic character; first, because the function is the same; second, because porous and cribriform vibracula have been observed in the genus Coscinopleura Marsson, 1885, which belongs to another quite distinct family; third, because auriform vibracula analogous to those of our species may be observed in the genus Otionella Canu and Bassler, 1917, which does not appear to be opesiulated; fourth, because the large auriform vibracula are possibly porous vibracula which have lost their frontal. There are specimens of Sele-

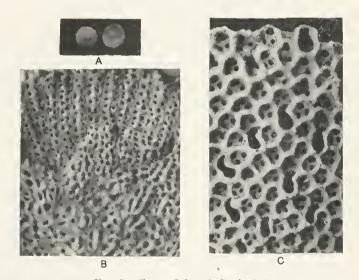


Fig. 5.—Genns Selenaria Busk, 1854.

A-C. Selenaria auricularia, new species.

A. Zoaria, natural size.

B, C. Noncelluliferous and celluliferous sides, ×20.

Eocene (Claibornian); Claiborne, Alabama.

naria maculata in the National Museum Collections provided with analogous auriculate vibracula.

Occurrence.—Eocene (Claibornian) Gosport sand; Claiborne, Alabama (rare). Cotypes.—Cat. No. 68481, U.S.N.M.

Family THALAMOPORELLIDAE Levinsen, 1909.

Genus MANZONELLA Jullien, 1888.

1888. Manzonella Jullien, Mission scientifique du Cap Horn, VI, Zoologie, Bryozoaires, p. 79. Opesiules well formed, sometimes multiple. Opesium terminal in the form of

an orifice. (After Jullien.)

Genotype. — Manzonella (Membranipora) exilis Manzoni, 1869, Pliocene.

Genus WOODIPORA Jullien, 1888.

1888. Woo lipora Jullien, Mission scientifique du Cap Horn, VI, Zoologie, Bryozoaires, p. 79.

Cryptocyst entirely developed; two regular opesiules; opesium with rounded contour. (After Jullien.)

Genotype.— Woodipora (Membranipora) holostoma (Wood), Pliocene.

Genus THALAMOPORELLA Hincks, 1887.

(For description, see Bulletin 106, U. S. National Museum, p. 268.)

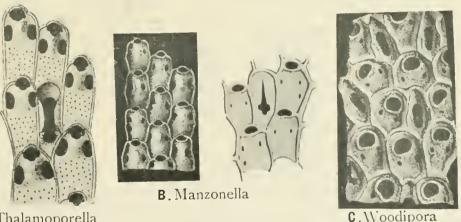
THALAMOPORELLA GRANULATA Levinsen, 1909.

Plate 2, fig. 14.

Thalanto porella granulata Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, p. 188, pl. 6a, figs. 1, 2; pl. 6, fig. 5.

Thalamoporella granulata Osburn, The bryozoa of the Tortugas Islands, Florida, Publica-1914. tion Carnegie Institution of Washington, No. 182, p. 197, fig. 8

Thalamoporella granulata CANU and BASSLER, Geology and Paleontology of the West 1919. Indies, Bryozoa, Publications Carnegie Institution of Washington, No. 291, p. 88, pl. 2, fig. 14.



A. Thalamoporella

Fig. 6.—Genera of the family Thalamoporellidae Levinsen, 1909.

- A. Thalamoporella rozieri Audouin, 1826, ×25 (after Waters, 1908).
- B. Manzonella exilis Manzoni, 1870 (after Manzoni, 1870, and Neviani, 1895).
- C. Woodipora holostoma Busk, 1859, ×25.

$$\label{eq:Measurements} \textit{Measurements.} - \text{Opesia} \left. \begin{array}{l} ho = 0.16 - 0.18 \text{ mm.} \\ lo = 0.16 - 0.18 \text{ mm.} \end{array} \right. \quad \text{Zooecia} \left. \begin{array}{l} |Lz = 0.80 \text{ mm.} \\ |z = 0.50 \text{ mm.} \end{array} \right.$$

Affinities.—We possess only a small fragment consisting of two zooecia; we are not certain therefore of our determination, as this has been made almost entirely on the micrometric measurements. The opesium is oval, the point above; the cryptoeyst is granulated; the mural rim has a width of 0.08 mm. and is granulated transversally.

Occurrence.—Lower Miocene (Bowden horizon): Rio Gurabo, Santo Domingo (rare).

Habitat.—Andaman or Nicobar Islands! (Levinsen); Tortugas Islands, Florida (Osburn).

Plesiotype.—Cat. No. 68482, U.S.N.M.

THALAMOPORELLA HIPERFORATA Cann and Bassler, 1919.

Plate 6, figs. 10-15.

1919. Thalamoporella biperforata CANU and BASSLER, Geology and Paleontology of the West Indies, Bryozoa, Publication Carnegie Institution of Washington, No. 291, p. 88, pl. 6, figs. 10-15,

Description.—The zoarium is bilamellar. The zooecia are elongate, distinct rectangular; the mural rim is thin, salient, bevelled, and bears two hollow tubercles on each side of the aperture. The cryptocyst is deep, flat, ornamented with large widely spaced pores and with numerous small pores closely placed together; the opesiules are very large, far distant from the apertura, placed symmetrically, but very unequal in size. The apertura is transverse, oval; the proximal border is always concave and the polypidian tube is limited by two small lateral indentations. The reticulocellarium is large, quite elongate; its cryptocyst is perforated by two opesiules; the opesium bears two lateral denticles serving as pivot to the corneous mandible.

Affinities .- The oral tuberosities are quite variable in size; they are often replaced by two fossettes.

This species is characterized by its onychocellarium, which resembles that of Thalamoporella granulata Levinsen, 1909, figured by Osburn, 1914. It differs, however, in the very different oval form of its apertura and in the presence of oral tuberosities. In its tuberosities this species is quite close to Thalamoporella rozieri Savigny-Audouin, 1826. It differs from it in the form of its onychocellarium provided with two opesiules, which are not figured by Levinsen in the numerous drawings which he has given.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Rio Cana and Rio Gurabo, Santo Domingo; and Bowden, Jamaica (common).

Cotypes.—Cat. No. 68483, 68484, U.S.N.M.

Family STEGANOPORELLIDAE Levinsen, 1909.

Genus STEGANOPORELLA Smitt, 1873.

(For description see Bulletin 106, U. S. National Musenm, p. 259.) STEGANOPORELLA PARVICELLA Canu and Bassler, 1919.

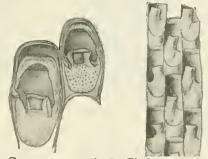
Plate 6, figs. 6-9.

1919. Steganoporella parvicella CANU and BASSLER, Geology and Paleontology of the West Indies. Bryozoa, Publication of the Carnegie Institution No. 291, p. 89, pl. 6, figs. 6-9.

Description.—The zoarium is unilamellar incrusting algae or bryozoa. The zooccia are distinct, elongate, separated by a shallow furrow; the mural rim is thin,

salient, finely granular. The cryptocyst is finely porous; the opesium is irregular or semilunar, a little elongate; the polypidian tube forms a rectangular surface, salient and excentric between the two dissimilar opesiular indentations. The large zooecia (B) are provided with a wide distal floor, and the polypidian tube is almost median.

Affinities.—The dimensions are quite variable, but they are always smaller than those of Steganoporella magnilabris Busk, 1854, which this species resembles in all its other characters.



A. Steganoporella B. Siphonoporella

Fig. 7.—Genera of the family Steganoporellidae Levinsen, 1909.

A. Steganoporella Smitt, 1873. S. magnilabris Busk, 1852. Recent. (After Harmer, 1900.)

B. Siphonoporella Hincks, 1880. S. delicatissima Busk, 1840, ×40. Recent. (After Levinsen, 1909.)

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (rare) and Bowden, Jamaica (common).

Cotypes.—Cat. Nos. 68485, 68486, U.S.N.M.

STEGANOPORELLA MAGNILABRIS Busk, 1854.

Plate 14, figs. 12, 13.

1854. Membranipora magnilabris Busk, Catalogue Marine Polyzoa, British Museum, pt. 2, Cheilostomata, p. 62, pl. 65, fig. 4.

1872. Steganoporella elegans Smitt, Floridan Bryozoa, collected by Count Pourtales, Pt. I. Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 10, No. 11, p. 15, pl. 4, figs. 96-101.

1890. Steganoporella magnilabris HARMER, A revision of the Genus Steganoporella, Quarterly Journal Microscopical Science, vol. 43, p. 279, pl. 12, fig. 10; pl. 31, figs. 44-46 (cites bibli ography).

1909. Steganoporella magnilabris Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, pp. 167, 168.

1913. Steganoporella magnilabris WATERS, Bryozoa from Zanzibar, Proceedings Zoological Society, London, p. 498, pl. 72, figs. 12-20 (cites bibliography).

1914. Steganoporella magnilabris Osburn, The bryozoa of the Tortugas Islands, Florida, Publication Carnegie Institute, Washington, No. 182, p. 196.

The zoaria are bilamellar; the two lamellae are easily separated. The micrometric measurements given by Harmer are quite variable; ours are more constant and always greater than those in *Steganoporella parvicella*.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (very rare). Pliocene (Caloosahatchee marl); Monroe County, Florida (very rare).

Habitat.—Western Atlantie: Off Florida (24-59 meters), Jamaica (16-19 meters), St. Vincent, Brazil (32 meters). Western Pacific: Off Japan (324 meters), China Sea (43 meters), Philippines (29 meters), Singapore to Honolulu (32-64 meters), Queensland (19-32 meters). Indian Ocean: Trincomalie, Admirality Islands (32-40 meters), East Africa (3-16 meters).

Plesiotypes.—Cat. No. 68487, U.S.N.M.

Family ASPIDOSTOMIDAE Canu, 1908.

Genus ODONTIONELLA Canu and Bassler, 1917.

1917. Odontionella Canu and Bassler, A Synopsis of American Early Tertiary Cheilostome Bryozoa, U. S. National Museum, Bull. 96, p. 12.

1920. Odontionella Canu and Bassler, Monograph North American Early Tertiary Bryozoa, Bull. 106, U. S. National Museum, p. 256.

Ovicell hyperstomial. No condyle for the opercular valve. The opesium is not symmetrical. The proximal border of the opesium bears irregular spinous processes, short, flat, more or less wide. The opesiular indentations are dissymmetrical. The operculum projects beyond the ridge. Sixteen tentacles. The avicularium is placed on the gymnocyst.

Genotypes.—Odontionella (Membranipora) hians Hincks, 1885, and Odontionella (Membranipora) occultata Waters, 1887. Range: Pliocene-Recent.

In 1917 we gave the generic definition of Waters's *M. hians* group. Later studies have permitted us to gain a better knowledge of the structure of this remarkable genus, which must be definitely placed in the Aspidostomidae on account of the nature of the ovicell. The genus differs from *Hemiseptella* Levinsen, 1912, also deprived of condyles in the nature of the opercular valve, in the presence of a gymnocyst and of a sort of vestibular arch, in the rarity of the lateral processes, and in

the presence of an ovicell. The retractor muscles of the polypide are attached in one of the zooecial angles. This results in a dissymmetry of the opesium and of

the opesiules as in Steganoporella, Onychocella, etc.

Like Waters, 1899, we have thought that *Membranipora savartii* Audouin, 1826, belonged to this group of species. It is in fact also provided with spinous processes and rather frequently with a proximal denticular plate. But this analogy is not sufficient; for, first, the opesium is symmetrical; second, we do not know whether there are opesiules in the chitinous cryptocyst; third, the operculum is totally different; and, fourth, there are only 15 tentacles.

The characteristics of this genus are much more visible on *Membranipora* occultata Waters, 1887, wherefore we believe it wise to add it as a second genotype.

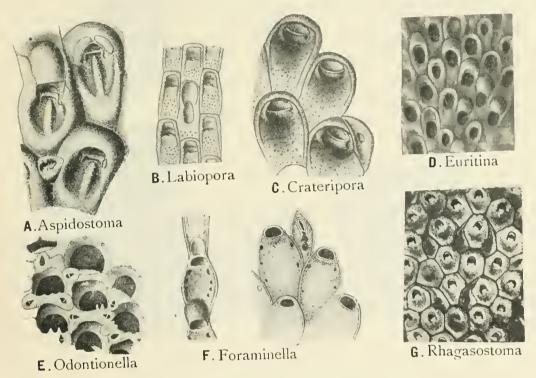


Fig. 8.—Genera of the family Aspidostomidae Canu, 1908.

- A. Aspidostoma, Hincks, 1881. A. giganteum Busk, 1884, × 25. Recent.
- B. Labiopora Levinsen, 1909. L. crenulata Levinsen, 1909, imes 23. Recent.
- C. Crateropora Levinsen, 1909. C. falcata. Levinsen, 1909, \times 23. Recent. (A-C after Levinsen, 1909.)
 - D. Euritina Canu, 1900. E. curita D'Orbigny, 1852, imes 20. Cretaceous.
 - E. Odontionella Canu and Bassler, 1917. O. occultata Waters, 1887. (After Waters, 1887.)
 - F. Foraminella Levinsen, 1909. F. lepida Hincks, 1881, \times 25. Recent. (After Hincks, 1881.)
- G. Rhagasostoma Koschinsky, 1885. R. hexagonum Koschinsky, 1885, ×17. Eocene. (After Koschinsky, 1885.)

Genus FORAMINELLA Levinsen, 1909.

1909. Foraminella Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, p. 165.

The somewhat arched frontal wall has no pores, but on each side a longitudinal series of (1-5) foramina (opesiules). A membranous opercular valve. Independent avicularia without crossbar, with an elongate mandible, on the one side furnished with a wing-like expansion. Hyperstomial ovicell with a membraneous ectooecium. Dietellae (Levinsen).

Genotype.—Foraminella (Monoporella) lepida Hincks, 1881. Recent.

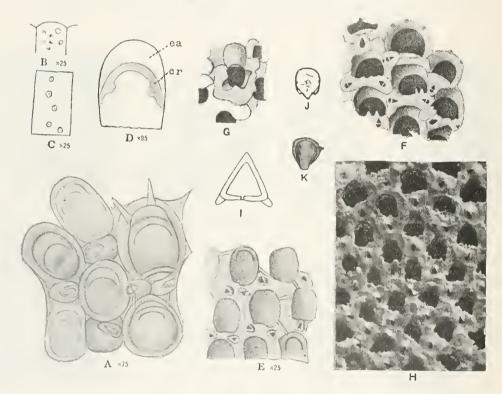


Fig. 9-Genus Odontionella Canu and Bassler, 1917.

A-E. Odontionella (Membranipora) hians Hineks, 1885.

A. Zooecia, \times 75. (After Hincks, 1885.) B. Distal wall with septulae, \times 25. C. Lateral wall with septulae, \times 25. D. Operculum, \times 85, showing chitinous ridge (cr) and expansion (ca). E. Zooecia \times 25. (B-E after Waters, 1898.)

F, G, I. Odontionella occultata Waters, 1887.

F. Specimen from recent seas with its avicularia. G. Example from the Tertiary rocks of New Zealand showing a thick calcareous deposit between the zooccia. I. Avicularian mandible, × 250. J. K. Opercula of another species (introduced by error.).

H. Photograph of a specimen of Odontionella hians Hincks from New Zealand showing the disymmetry of the opesiular indentations, \times 20.

Genus LABIOPORA Levinsen, 1909.

(For description, see Bulletin 106, U. S. National Museum, p. 258.)

LABIOPORA MIOCENICA Canu and Bassler, 1919.

Plate 6, fig. 1.

1919. Labiopora miocenica Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publication of the Carnegie Institution of Washington, No. 291, p. 89, pl. 6, fig. 1.

Description.—The zoarium is unilamellar. The zooecia are elongate, distinct, rectangular; the mural rim is quite thin, round. The cryptocyst is little deep, flat, and formed of a tremocyst superposed on an olocyst perforated with corresponding pores. The opesium is tranverse, terminal or placed at the base of a funnel, formed by the much developed vestibular arch. The polypidian tube is visible, wide, median.

$$\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{cases} ho = 0.10 \text{ mm.} \\ lo = 0.24 - 0.28 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.50 - 0.64 \text{ mm.} \\ lz = 0.32 - 0.34 \text{ mm.} \end{cases}$$

Affinities.—This new species differs from Labiopora crenulata Levinsen, 1909, in the wide form of the zooecia and in its transverse opesium.

Occurrence.—Lower Miocene (Bowden horizon): Rio Gurabo, Santo Domingo (rare).

Holotype.—Cat. No. 68488, U.S.N.M.

CALPENSIIDAE, new family.

No ovicell. The cryptocyst is perforated by one or two opesiules.

We suppose that the ovicell is formed by an internal sac placed in the vicinity of the opercular region as in the genus *Diplodidymia* Reuss, 1869. The different genera which we classify here belong perhaps to distinct families; but as we are ignorant of the larva, it is preferable to group them provisionally under the same name. They may be classified in three groups.

1. A single opesiule. Diplodidymia Reuss, 1869.

2. Cryptocyst with spinous processes. Hemiseptella Levinsen, 1909; Cupularia Lamouroux, 1821.

3. Complete cryptocyst. Calpensia Jullien, 1888; Verminaria Jullien, 1888; Microporina Levinsen, 1909; Corynostylus Canu and Bassler, 1919.

Genus DIPLODIDYMIA Reuss, 1869.

1869. Diplodidymia Reuss, Zur fossilen Fauna der Oligocänschichten von Gaas, Sitzungsberichte der k. Akademie der Wissenschaften, Wien, vol. 59, Abth. 1, p. 469.

The zoarium is articulated and radicelled. Each segment is formed of four rows of zooecia placed diagonally. No ovicell. Two very small pores are placed on each side before the apertura. The cryptocyst is perforated laterally by a long linear opesiule; it gives passage to the wide muscles attached to the ectocyst in a scelerity (=chitinous thickening). The avicularium is by the side of the apertura

and the mandible is triangular. The avicularian chamber bears a radicular perforation. Eleven to twelve tentaeles.

Genotype.—Diplodidymia complicata Reuss, 1869. Range: Montian-Recent. A detailed study of this genus was made by Waters, 1913. The ovicell is a sac hanging from the opercular region; the egg evolves there until the formation of the larva. Reuss classified this genus in a special family.

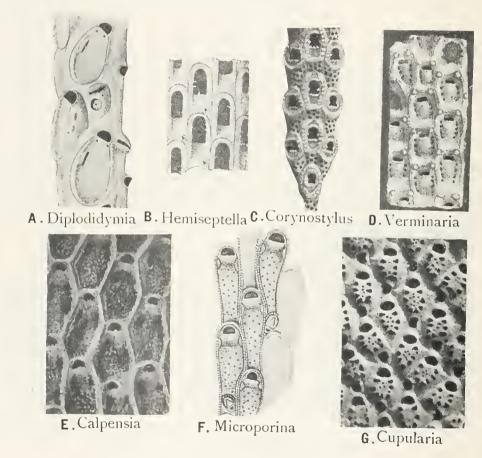


Fig. 10.—Genera of the family Calpensiidae, new family.

- A. Diplodidymia Reuss, 1869, D. complicata Reuss, 1869, \times 85. Recent, Africa. (After Waters, 1913.)
- B. Hemiseptella Levinsen, 1909, H. labiata Busk, 1884, \times 12. Recent, Cape Horn. (After Waters, 1895.)
 - C. Corynostylus Canu and Bassler, 1919, C. labiatus Canu and Bassler, 1919, imes 20. Miocene.
 - D. Verminaria Jullien, 1888, V. oblonga Busk, 1859. Crag of England. (After Busk, 1859.)
- E. Calpensia Jullien, 1888, C. impressa Moll, 1803, \times 25. Recent, Atlantic. (After Guerin-Ganivet.)
- F. Microporina Levinsen, 1909, M. elongata Hincks, 1880 \times 25. Recent, Australia. (After Hincks, 1801.)
 - G. Cupularia Lamouroux, 1821. C. umbellata Defrance 1823, \times 20.

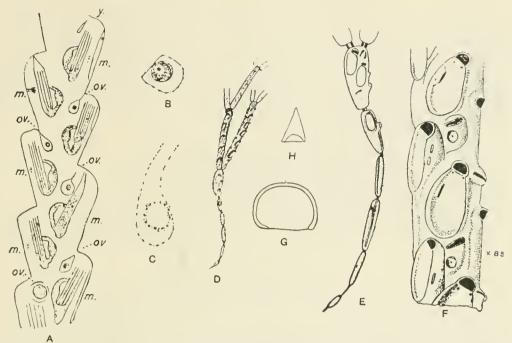


Fig. 11.—Genus Diplodidymia Reuss, 1869.

Figs. A-H. Diplodidymia complicata Reuss, 1869.

A. Sketch, × 50 showing small sacs hanging from the opercular region, and in each an ovum grows (ov). At y. there seems as yet no ovum in the small sac; m., the muscles of the opesium. B. Section showing an ovum in the small sac, × 750. C. Section, × 750, in which the ovum has segmented and a blastula has formed. D. Colony from Chuaka, Africa, × 6. E. Lower part of colony, × 25. F. Portion of zoarium, × 85, showing the structure of the zooecia and avicularia. G. Operculum, × 250.
H. Mandible of the avicularium, × 250. (A-H, after Waters, 1913.)

Genus HEMISEPTELLA Levinsen, 1909.

1909. Hemiseptella Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, p. 164.

No ovicell. The distal extremity of the cryptocyst plunges into the zooecium. The dissymmetric opesium bordered laterally and inferiorly by spines. The two opesiules, distinct or not are limited sometimes by the cryptocyst, sometimes by the spinous processes. The opercular valve is garnished superiorly by an opercular arch strongly chitinized. Small avicularium. Twenty-six tentacles.

Genotype.—Hemiseptella (Vincularia) labiata Busk, 1884. Range: Miocene-Recent.

Structure.—We have chosen Vincularia labiata as the genotype because it presents two sorts of spinous processes, the larger of which can unite in order to form a bridge intended to support the hinge of the opercular valve. But this bridge is only a secondary character. The cryptocyst covers the spinous processes. Levinsen attributed to this genus a certain number of species from the Cretaceous figured by D'Orbigny. This is an error; these species belong to the genus Floridina, for their ovicell is endozoocial and they bear onychocellaria.

One must not confuse the spinous processes or spinules with the spines. Their structure is not similar and their physiologic rôle is absolutely distinct. The spinous processes are here only internal apparatus for support, probably intended to limit the movements of the hypostege.

The large retractor muscle of the polypide is inserted in a corner of the zooecium as in Onychocella, Odontionella, and Steganoporella. It results in a great dissymme-

try of the opesium especially on its proximal border.

The known species of this genus are:

Hemiseptella (Vineularia) labiata Busk, 1884.

Hemiseptella (Vincularia) steganoporoides Goldstein, 1882.

Hemiseptella (Thalamoporella) michaelseni Calvet, 1904.

Hemiseptella (Biflustra) denticulata Smitt, 1872.

Hemiseptella (Membranipora) tenuis Desor, 1848.

Hemiseptella (Membranipora) laeinia Tuomey and Holmes, 1857.

Hemiseptella (Membranipora) minor Canu, 1908 (M. suleata, var. minor Canu).

HEMISEPTELLA LATA Canu and Bassler, 1919.

Plate 2, fig. 4.

1919. Hemiseptella lata Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa Publication of the Carnegie Institution of Washington, No. 291, p. 85, pl. 2, fig. 4.

Description.—The zoarium incrusts fronds of the bryozoan Metrarabdotos. The zooecia are distinct, little elongated, wide, subrectangular; the mural rim is thin. The opesium is elliptical or orbicular, never symmetrical; the cryptocyst is short, little deep, irregular; the opesiular indentations are represented by two lateral dissymmetric concavities and are often separated by a wide and serrate denticle.

Measurements.—Opcsia
$$\begin{cases} ho = 0.30 \text{ mm.} \\ lo = 0.22 - 0.30 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.42 \text{ mm.} \\ lz = 0.30 - 0.32 \text{ mm.} \end{cases}$ It is always difficult to characterize a species from the study of a

Affinities.—It is always difficult to characterize a species from the study of a single specimen. Exteriorly the zooecia have the form of certain zooecia observed in Aeanthodesia savartii Savigny-Audouin, 1826; it differs from it in its nonsymmetric opesium and in two opesiular sinuosities.

This species differs from *Biflustra savarti* Smitt, 1872, which is not perhaps the species of Audouin, in the more constant development of its cryptocyst and in the nature of its zoarium, which does not incrust algae. The proximal denticle has been clearly observed on three zooecia.

Oecurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (very rare).

Holotype.—Cat. No. 68498, U.S.N.M.

HEMISEPTELLA? LACINIA Tuomey and Holmes, 1857.

Plate 14, fig. 10.

1857. Membranipora lacinia Tuomey and Holmes, Pleiocene Fossils of South Carolina, p. 14, pl. 4, fig. 10.

We have been unable to find any specimens corresponding to the figure of Membranipora lacinia, which, moreover, is somewhat confusing, although probably indicating the genus Hemiseptella. We reproduce the original figure in the hope that some future student will rediscover the species.

Occurrence.—Miocene?: Smith's, Goose Creek, South Carolina.

HEMISEPTELLA GRANDICELLA Cann and Bassler, 1919.

Plate 5, fig. 12.

1919. Hemiseptella grandicella Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, No. 291, p. 84, pl. 5. fig. 11.

Description.—The zoarium incrusts shells. The zooeeia are large, elongate, ogival, distinct, separated by a furrow of little depth; the mural rim is narrow, little salient, round, finely granulated; the cryptocyst is large, flat, granulated; the opesium is elongated, elliptical, irregular and nonsymmetrical in the proximal portion.

Measurements.—Opesia
$$\begin{cases} ho = 0.40 \text{ mm.} \\ lo = 0.25 \text{ mm.} \end{cases}$$
 Zooceia $\begin{cases} Lz = 0.65 - 0.70 \text{ mm.} \\ lz = 0.35 \text{ mm.} \end{cases}$ Affinities.—The group of inferior opesiular spines was certainly dissymmetrical;

trace of the lateral spines is searcely visible on our specimens.

This species differs from Hemiscotella denticulata Smitt, 1872, in the absence of nodosities in the interzooecial angles and in the nontrifoliate opesium. It differs from Hemiseptella tenuis Desor, 1848, in the larger micrometric measurements and in the more fragile and less numerous spines.

Occurrence.—Lower Miocene (Bowden marl): Bowden, Jamaica (rare). Holotype.—Cat. No. 68490, U.S.N.M.

HEMISEPTELLA FILIMARGO, new species.

Plate 10, fig. 9.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a very thin salient thread, elongated, subhexagonal; the mural rim is very thin, filiform, salient; the cryptocyst is large, flat, shallow. The opesium is semiclliptical, transverse, with an irregular proximal border, two large lateral spicules and many spicules placed inferiorily.

Measurements.—Opesia
$$box{lo = 0.12-0.18 mm.} \\ box{lo = 0.15-0.20 mm.}$$
 Zooecia $box{lz = 0.40-0.50 mm.} \\ box{lz = 0.25-0.30 mm.}$

Variations.—The micrometric measurements are very irregular. Interzooccial tubereles are sometimes present. On our single specimen the spicules are badly preserved. The extreme thinness of the mural rim well characterizes this species. Cases of regenerated zooecia have been observed.

Occurrence.-Miocene (Yorktown formation): York River, Virginia (very rare).

Holotype.—Cat. No. 68491, U.S.N.M.

HEMISEPTELLA TUBEROSA, new species.

Plate 46, fig. 10.

Description.—The zoarium incrusts shells. The zooecia are distinct, elongated, rectangular, ornamented with large interzooccial tubercles; the mural rim is thick, rounded, granulated; the cryptocyst is much reduced, deep, granulated. opesium is elongated, elliptical, with an irregular proximal border.

$$\label{eq:measurements} \textit{Measurements.} - \text{Opesia} \begin{cases} ho = 0.25 \text{ mm.} \\ lo = 0.15 \text{ mm.} \end{cases}$$
 Zooecia $\begin{array}{c} Lz = 0.35 \text{ mm.} \\ lz = 0.25 \text{ mm.} \end{array}$

Variations.—On our type specimen there are very few spicules visible. micrometrie measurements are very irregular.

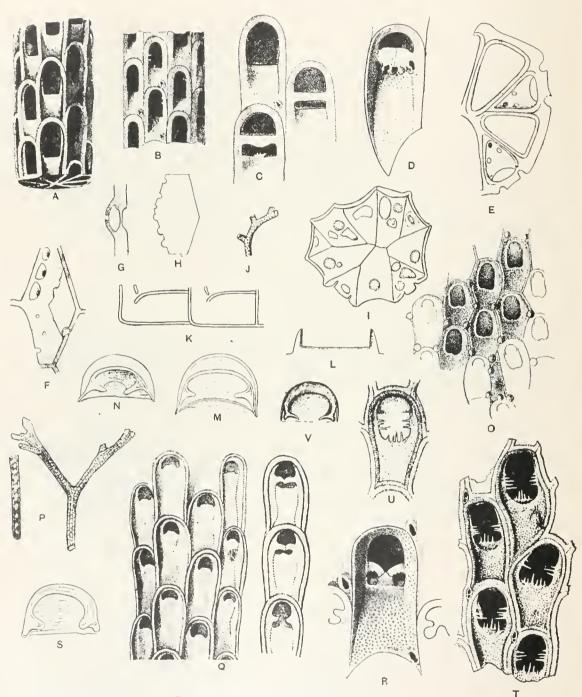


Fig. 12.—Genus Hemiseptella Levinsen, 1909.

Fig. 12.—Genus Hemiseptella Levinsen, 1919.

Figs. A-N. Hemiseptella labiata Busk, 1884.

A. A branch of the form Vincularia elegans (after D'Orbigny, 1839). B. A branch from Cape Horn, \times 12. A few minute denticles or hooks occur on the border of the opesium. C. Several zooecia of the type of Vincularia elegans preserved at the Museum of Paris. A growth from each side forms a calcareous bridge over the opesium. It is placed under the membraneous ectocyst. D. A calcined zooecium of a specimen from the Atlantic (Challenger collection), \times 25. E. Transverse section, \times 25. F. Calcareous section diagonally through a zooecium, thus showing the projecting rosette plates (septulae), \times 25. G. Section through the dietellae, \times 250. H. Diagrammatic section showing the position of the septulae, \times 25. I. Transverse, decalcified section, \times 25. J. A branch, natural size (after D'Orbigny, 1839). K. Section in the zooecia, \times 25. Besides the lip which projects upward, the front wall of the zooecium is directed downward thus contracting the zooecium near the aperture. (Figs. D-H, K, (after Waters, 1889). L. Transverse section in operculum, \times 85 (B, C, I, L, after Waters, 1895). M., N. Operculum (after Busk 1884).

Fig. O. Hemiseptella denticulata Smitt 1872. A zoarium from Florida. The unshaded zooecia

are covered by the ectocyst. They show the chitinous arch of the opercular valve.

Fig. P-8.—Hemiseptella steganoporoides MacGillivray, 1881. P. A zoarium, natural size, from Prince Edward Island. Q. Zooecia with their opesiular in dentations and occasionally the bridge, \times 25 (forma Vincularia gothica). R. A zooecium showing the sutures between the three processes and the avicularium (after Waters, 1889). S. Operculum, \times 85 (figs. P, Q and S, after Busk, 1884).

Figs. T-V.—Hemiseptella michaelseni Calvet, 1904. T. Zooecia, × 25, showing the three groups of spinous processes bordering the opesium. U. A zooecium covered by its membranous transparent ectocyst under which the spinous processes are seen. V. Operculum, × 85 (T-V, after Calvet, 1907).

This species is much smaller than *Hemiseptella rectangulata*, in which the form of the zooccia is identical but which is deprived of tubereles.

Occurrence.—Pleistocene: Simmons Bluff, Yonges Island, Charleston County, South Carolina (rare).

Holotype.—Cat. No. 68492, U.S.N.M.

HEMISEPTELLA FISTULA Ulrich and Bassler, 1904.

Plate 14, fig. 9.

1904. Membranipora fistulu Ulrich and Bassler, Bryozoa: Maryland Geological Survey, Miocene. p. 413, pl. 112, fig. 5.

The original description is as follows:

Zoarium so far as observed forming small, hollow, subcylindrical stems, about 1.5 mm. in diameter, composed of twelve to fifteen longitudinal rows of zooecia. Walls about as wide as the zooecial openings, obtusely carinate, the carinac between the ends of the cells high and bent forward so as to impart a slightly imbricating appearance to successive cells. Opesial opening elongate elliptical; immediately behind it a minute pore is occasionally noticeable. Measuring longitudinally about 8 zooecia in 5.0 mm. No avicularian nor vibracular cells observed.

The original description of this slender rod-like bryozoan gives an adequate idea of the species, of which unfortunately no more specimens have been discovered. We reproduce a new illustration of the type specimen which shows spines irregularly distributed on the proximal border of the opesium, thereby causing the species to be referred to *Hemiseptella*.

Occurrence.—Miocene (St. Mary's formation): St. Mary's River, Maryland (very rare).

Holotype.—Cat. No. 68493, U.S.N.M. 12184—23—Bull, 125—6

HEMISEPTELLA RECTANGULATA, new species.

Plate 14, fig. 11.

Description.—The zoarium incrusts shells. The zooccia are distinct, adjacent, elongated, rectangular, with rounded distal border. The cryptocyst is deep, granular, flat; the opesium is elliptical, elongated with a proximal border irregular and without symmetry; the spinous processes are short, fragile, thin, numerous. No dietellae.

Measurements.—Opesia $box{lo} = 0.30 \text{ mm}$. Zooecia $box{lz} = 0.47 \text{ mm}$. $box{lz} = 0.22 \text{ mm}$.

The spinous processes are very fragile; they little resist fossilization, but their

vestiges are easily observed.

Affinities.—This species differs from Hemiseptella granulosa in its smooth mural rim and its smaller micrometric measurements. It differs from Hemiseptella grandicella in its deep cryptocyst and its smaller micrometric measurements.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (rare).

Holotype.—Cat. No. 68494, U.S.N.M.

HEMISEPTELLA GRANULOSA, new species.

Plate 29, fig. 8.

Description.—The zoarium is unilamellar. The zooccia are distinct, separated by a furrow of little depth, elongated, rectangular; the mural rim is thin convex, granular; the cryptocyst is of little depth, flat, slightly granular. The opesium is elliptical, irregular on its proxial border which is never symmetrical; the spinous processes are distributed in the lower half and the proxial bundle is wider and more salient.

Measurements.—Opesia
$$\begin{cases} ho = 0.40 \text{ mm.} \\ lo = 0.20 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.70 \text{ mm.} \\ lz = 0.35 \text{ mm.} \end{cases}$

Affinities.—The micrometric measurements are quite variable, even on a simple zoarium; but the relationship of one-half between the two dimensions remain rather constant.

This species differs from *Hemiseptella planulata*, in which the cryptocyst is also very superficial in its granulated mural rim and in its crenulated opesium. It differs from *Hemiseptella grandicella* in its micrometric measurements, generally smaller, and in its granular mural rim.

The spinous processes are rather fragile; only their base resists fossilization, and they appear to be grouped into three bundles. The lowest one is wide and vigorous. The asymmetry of the proximal border of the opesium seems to indicate that the large retractor muscle of the polypide was inserted, not in the median axis of the zooecium, but in a lower corner, as in *Onychocella* and *Steganoporella*.

Occurrence.-Miocene: Near Charleston, South Carolina (very rare).

Holotype.—Cat. No. 68495, U.S.N.M.

HEMISEPTELLA PLANULATA, new species.

Plate 29, figs. 5, 6.

Description.—The zoarium incrusts shells (Pectunculus). The zooecia are distinct, elongated, with adjacent mural rims, hexagonal, ogival; the mural rim is thin, little salient; the cryptocyst is little deep, large, flat. The opesium is ogival or

subtrifoliate; the mural rim is thin, little, salient; the cryptocyst is little deep, large, flat. The spinous processes are very fragile and are distributed into five principal bundles; the lower bundle is salient and placed without symmetry on the proximal border of the opesium.

Measurements.—Opesia
$$\begin{cases} ho = 0.24 \text{ mm.} \\ lo = 0.16 - 0.20 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.30 \text{ mm.} \\ lz = 0.24 - 0.30 \text{ mm.} \end{cases}$

Affinities.—The walls of this species are very fragile and are easily worn; the zooecia are then scarcely distinct, and certain zooecia bear tubercles at the angles.

This species is quite close to *Hemiseptella denticulata* Smitt, 1872, which also presents tubercles. It differs from it in its less deep cryptocyst and in the inconstancy of the tubercles. However, we possess no specimen of this species and our comparisons are made only from the figures. It differs from *Hemiseptella granulata* in the absence of granules on the mural rim and in its subtrifoliate opesium.

· Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare).

Cotypes.—Cat. No. 68496, U.S.N.M.

Genus CUPULARIA Lamouroux, 1821.

1821. Cupularia Lamouroux, Exposition methodique des genres de l'ordre des Polypiers, p. 44.

The zoarium is a more or less expanded cup; it is provided with a special hydrostatic system accompanied by vibracula. The opesium is fringed with spinous processes which are flat, free, or joined together; the two distal processes are symmetrically placed and serve as support to the opercular valve. The two distal opesiules are always rounded. No ovicell. Vestibular arch present.

Genotype.—Cupularia umbellata Defrance, 1823. Range: Miocene-Recent.

Affinities.—The genus Cupularia is identical in its frontal with the genus Hemiseptella Levinsen, 1909. It differs from it in its cup-shaped zoarial form, in the presence of vibracula, the flat form of the spinous processes, and in the union of the latter.

Cupularia lives like Lunularia, of which we have explained the hydrostatic zoarial mechanism. (See North American Early Tertiary Bryozoa, p. 238). The cellular or external face is the inferior face; the noncellular or internal face is the superior face. The ancestrular zooccia are often calcified, indeed only the opercular valve may be visible there; these are the hydrostatic zooccia (aborted of D'Orbigny); they are never radicular as in the genus Lunularia. The larvae probably develop in an oral sac, as in the genus Diplodidymia Ruess, 1869.

The union of the spines is not a generic character. In fact it may be accidental (as in Cupularia denticulata), partial (C. reussiana), almost complete (C. umbellata).

This union of the spinous processes forms a cryptocyst.

Smitt, 1872, recognized that this genus ought to be classed in the Microporidae. This was correct, as the opesiules or opesiular slits are always clearly visible. In order to show the character of the genus in more detail we have introduced descriptions of a few European species.

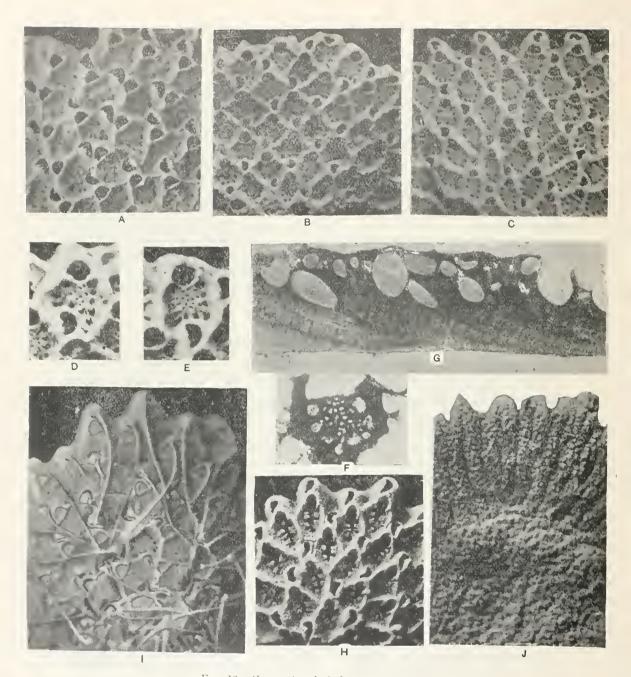


Fig. 13. Genus Cupularia Lamouroux, 1921.

A. F. Cupularia umbellata l'efrance, 1923.

A. Zooccia. × 25 of specimen dredged in the Mediterranean at Oran (Algeria). B. An example, × 25, with small, wide zooccia. C. Specimen with small, long zooccia, × 25. D. E. Zooccia enlarged × 50 showing the formation of the cryptocyst by the partial union of spinous processes. F. Tangential thin section of the frontal, × 75, showing the successive ramifications of the lateral spinons processes.

G. Longitudinal thin section, \times 75, showing the successive zones of calcification.

J. Internal face (superior) of a zoarium, × 25. (B-G, J-Burdigalian of Merignac, Gironde France. H, I. Cupularia multispinata, new species.

II. Portion of a zoarium deprived of its ectocyst, > 25.—I. Zoarium > 25., provided with its ectocyst, and its vibracula, Mediterranean at Oran (Algeria).

CUPULARIA HAIDINGERI Reuss, 1847.

Plate 1, figs. 13-17.

1847. Lunulites haidingeri Reuss, Die fossilen Polyparien des Wiener Tertiarbeckens, Haidinger's naturwissenschaftliche Abhandlungen, vol. 2, p. 58, pl. 7, fig. 26, 27.

1859. Cupularia denticulata Busk, Monograph Fossil Polyzoa of the Crag, Publications Paleontolo-

graphical Society, p. 85, pl. 13, fig. 1. 1877. Cupularia haidingeri Manzoni, I Briozoi fossili del Miocene d'Austria ed Ungheria, parte 2, Denkschriften der k. Akademie der Wissenschaften, Wien, vol. 37, p. 73, pl. 16, fig. 54.

1880. Cupularia haidingeri Seguenza. La formazioni tergiarie nella Provincia di Reggio (Calabria), Reale Accademia dei Lincei, Memorie della Classe di Scienze Fisiche, Matematiche e Naturali, ser. 3, vol. 6, pp. 84, 131.

 $\label{eq:large_loss} \textit{Measurements.} - \text{Large zooeeia} \begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.36 - 0.40 \text{ mm.} \end{cases}$

Variations.—The place of the opercular valve is clearly indicated by a distal arch limited by two symmetrical apophyses or condyles. The spinous processes are flat, often very wide, claviform or finely denticulated; their number is five (which includes the two oral apophyses). Their form is inconstant, and we figure some interesting variations. Our determination of the species has been verified by specimens from Porzteich, Austria-Hungary. The internal face bears radial, bifureated, and tuberous ribs.

Affinities.—This species differs from Cupuladria canariensis, Busk, 1859, in the presence of spinous processes and in the absence of the large pores of the internal face. It differs from Cupularia denticulata Conrad, 1841, in its larger and flatter zoarial form, its larger zooccia, and in the larger tuberosities of the internal face.

Geological distribution.—Helvetian of Italy (Seguenza); Tortonian of Italy (Seguenza), of Austria-Hungary (Reuss); Redonnian of France (Canu); Plaisancian of England (Busk).

CUPULARIA DOMA D'Orbigny, 1851.

Plate 1, fig. 18; plate 15, figs. 1-5.

1851. Discoflustrellaria doma D'Orbigny, Paléontologic française, Terrain Crétacé, vol. 5, Bryozoaires, p. 561.

1859. Cupularia johnsoni Busk, Zoophytology: On some Madeiran Polyzoa, Quarterly Journal

Microscopical Science, vol. 7, p. 67, pl. 23, figs. 1-6.
1872. Cupularia doma Smrt, Floridan Bryozoa, collected by Count Pourtales, Kongl. Svenska, Vetenskaps-Akademiens Handlingar, pt. 2, vol. 11, No. 4, p. 15, pl. 3, figs. 81-84 (not Busk,

1877. Cupularia reussiana Manzoni, Bryozoaires du pliocene superieur de l'Île de Rhodes, Memoires de la Société Géologique de France, ser. 3, vol. 1, p. 67.

1909. Cupularia johnsoni Norman, Polyzoa of Madeira and neighboring islands, Journal Linnean Society London, Zoology, vol. 30, p. 290, pl. 38, figs. 1-6.

Variations.—The spinous processes are flat or acuminate; there are even some which are spatulate and some fimbriated. The two superior ones form two symmetrical condyles serving as pivot to the opercular valve. The others are three in number; they are dissymmetrical, irregularly placed, without definite and constant form, never joined. The cryptocyst is more or less developed and its presence considerably modifies the aspect of the opesium. The internal face bears hydrostatic tuberosities (as usual), very inconstant in their size; they are generally very large, but may disappear almost totally. The zoarium is conical, higher than wide. The two opesiules are formed by two rounded indentations symmetrically placed below the condyles.

Affinities.—Smitt has confused this species with Cupularia reussiana Manzoni, 1869. He saw D'Orbigny's type in Paris. Manzoni and Norman have adopted the same synonymy. For all these authors the essential character was the absence of a complete calcareous pivot for the opercular valve, as in Cupularia umbellata Defrance, 1823. In reality all the denticulata group possess this character and the species are differentiated from each other not only in their zoarial form but also by the number and nature of their spinous processes and in the form of their hydrostatic tubercles.

This species differs from Cupularia multispinata, new species, in the presence of three (and not five) spines, and in the conical form of the zoarium. It differs from Cupularia denticulata Conrad, 1841, which has the same number of spinous processes, in the more conical form of the zoarium, and in the presence of large hydrostatic tuberosities on the internal face. It differs from Cupularia reussiana Manzoni, 1869, in the more conical form of the zoarium and in its spinous processes never joined together.

Occurrence.—Sicilian of Rhodes (Manzoni). Miocene (Duplin marl): Wilmington, North Carolina.

Habitat.—Mediterranean: Shores of Algeria (72 meters), Oran (81–103 meters). Atlantic: Madeira (64–113 meters). Gulf of Mexico: Florida (Smitt).

CUPULARIA MULTISPINATA, new species.

Description.—The zoarium is discoidal, thin, very convex, much less high than wide. The zooccia are elongated, distinct, hexagonal; the mural rim is thin, little salient; the cryptocyst is more or less developed laterally. The opesium is ornamented by at least five flat spinous processes nonsymmetrical, rather wide, finely denticulated. Two symmetric salient condyles limit the apertura. A vestibular arch is present. The vibracula are large and auriform.

Measurements.—Large zooecia
$$\begin{cases} Lz = 0.48 \text{ mm.} \\ lz = 0.32 \text{ mm.} \end{cases}$$

Affinities.—This species bears the most numerous spinous processes and this character is amply sufficient to differentiate it from Cupularia reussiana Manzoni, 1869; C. doma D'Orbigny, 1851; and from C. denticulata Conrad, 1841. The hydrostatic zooecia have their cryptocyst entirely calcified.

Habitat.—Mediterranean: Gulf of Oran (105 meters).

CUPULARIA REUSSIANA Manzoni, 1869.

Plate 1, figs. 19-22.

1869. Cupularia reussiana Manzoni, Bryozoi Pliocenici Italiani, Sitzungsberichte der kaiserlichen Akademie der Wissenschaften, Wien, vol. 59, p. 27, pl. 2, fig. 19.

1895. Cupularia reussiana Neviani, Briozoi fossili della Farnesina e Monte Mario presso Roma, Palaeontographia Italica, vol. 1, p. 102 (sep. 26).

1895. Cupularia reussiana Neviani, Briozoi neozoici di alcune localita d'Italia, Bollettino della Societe Romana per gli Studi Zoologici, parte 1, vol. 4, p. 45 (sep. 7); parte 2, p. 243 (sep. 19); parte 5, vol. 7, p. 101 (sep. 4), p. 103 (sep. 6).

1900. Cupularia reussiana Neviani, Briozoi neogenici delle Calabrie, Paleontologia italica, vol. 6, p. 169 (55) (regional bibliography). 1900. Cupularia reussiana Neviani, Briozoi Terziari della Toscana, Bollettino Sociètá geologica italiana, vol. 19, p. 19 (sep.).

1913. Cupularia reussiana Canu, Contributions a l'Etude des Bryozoaires fossiles: IV. Pliocene d'Alger, Bulletin Société Géologique de France, ser. 4, vol. 13, p. 126.

1913. Cupularia haidingeri Canu, Bulletin Société Géologique de France, ser. 4, vol. 13, p. 128.

Historical.—Manzoni's figures of 1869 are exact, although the zoarinm is much less conical and almost cupuliform. As the figure of the cellular face has served for the determination by Seguenza, Canu, and Neviani we can hold as exact the

synonymy given, moreover partially verified by Canu.

Variations.—The apertural arch is constant; it forms with the two wide, lateral opesiular indentations a trifoliate opesium. The inferior spinous processes are partially united and separated at their base by three large pores which are perhaps secondary opesiules. This character is specific. The vestibular arch is very large; the internal face is covered with large tuberosities.

Affinities.—All the authors, on the authority of Smitt, have confused this species with Cupularia doma D'Orbigny, 1851, the examination of the zoarial form having been judged sufficient. It differs much from it in its cupuliform zoarium (and not very conical) and in the union of the spinous processes occasioning the formation of three large frontal pores.

This species differs from Cupularia umbellata Defrance, 1823, in the presence of three frontal pores only, and in its opesiules not separated from the apertura and

reduced to the state of indentations.

Geological distribution.—Tortonian of Italy (Seguenza, Neviani); Zanclean of Italy (Seguenza); Plaisancian of Italy (Neviani); Astian of France and Italy (Canu); Sicilian of Italy (Waters, Seguenza).

This species has not yet been found in the Mediterranean dredgings carried out

by Canu. Waters did mention it at Naples.

CUPULARIA DENTICULATA Conrad, 1841.

Plate 15, figs. 6-10.

1841. Lunulites denticulata Conrad (in Hodge), Observations on the Secondary and Tertiary formations of the southern Atlantic States, American Journal Science and Arts, ser. 1, vol. 41, p. 348.

1845. Lunulites denticulata Lonsdale, Report on the Corals from the Tertiary formations of North America, Quarterly Journal Geological Society, London, vol. 1, p. 503.

1857. Lunulites denticulata Tuomey and Holmes, Pleiocene Fossils of South Carolina, p. 11, pl. 4, figs. 1-5.

1904. Cupularia denticulata Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 414, pl. 112, fig. 6.

Measurements.—Large zooecia $\begin{cases} Lz = 0.30\text{--}0.40 \text{ mm.} \\ lz = 0.20\text{--}0.24 \text{ mm.} \end{cases}$

Variations.—The hydrostatic zooccia are thin, calcified, probably deprived of polypide, garnished with a complete cryptocyst; their apertura is small and serves probably as passage for a more or less long bristle; they surmount the grain of sand on which the larva is fixed. The internal face is flat or curved; the radiating ribs are generally smooth, but small tuberosities are not rare. The two condyles are not always exactly symmetrical. The spinous processes are acuminate or flat,

short, finely denticulated, numbering three or four; they are irregular and their position never symmetrical. The zoarium is rather variable. The small specimens are always wider than high. The large specimens are cupuliform but always very convex.

This species differs from Cupularia haidingeri Reuss, 1847 (= C. denticulata Busk, 1859), in the smaller micrometric measurements and in the absence of the large hydrostatic tuberosities of the internal face. It differs from Cupularia reussiana Manzoni, 1869, in its spinous processes never united. It differs from Cupularia doma D'Orbigny, 1851, which it most resembles in its large, thin, cupuliform zoarium and in the absence or attenuation of the large tuberosities of the internal face.

Occurrence.—Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina (type locality); Wilmington, and many other localities in North and South Carolina. Miocene (Choctowhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (common). Miocene (St. Mary's formation): St. Marys River, Maryland (rare). Miocene (Yorktown formation): Williamsburg, York River, and other localities in Virginia (rare). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (very common). Pliocene (Caloosahatchee marl): Monroe County, Florida (very common).

Plesiotypes.—Cat. Nos. 68497-68505, U.S.N.M.

CUPULARIA UMBELLATA Defrance, 1823.

Plate 2, figs. 15-19.

- 1823. Lunulites umbellata Defrance, Dictionnaire du Sciences Naturelles, vol. 27, p. 361.
- 1862. Discoporella denticulata Gabb and Horn, Monograph of the fossil Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy of Natural Sciences of Philadelphia, ser. 2, vol. 5, p. 142, pl. 20, fig. 25.
- 1872. Cupularia umbellata Smitt, Floridan Bryozoa, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 11, p. 14, pl. 3, figs. 75-80.
- 1889. Cupularia umbellata Jelly, A synonymic Catalogue of marine Bryozoa, p. 79 (general bibliography).
- 1895, 1900. Cupularia umbellata Neviani, Briozoi, neozoici di alcune localita d'Italia, Bollettino della Società Romana per gli Studi Zoologici, pt. 2, vol. 4, 1895, pp. 237, 243; pt. 4, vol. 7, 1898, p. 38; pt. 5, vol. 8, 1898, pp. 98, 100, 106; pt. 6, (2), vol. 1, p. 60.
- 1896. Cupularia umbellata Neviani, Briozoi fossili della Farnesina, Palaeontographia Italica, vol. 1, p. 101 (sep. 25).
- 1900. Cupularia umbellata Neviani, Bryozoi, terziari et posterziari della Toscana, Bolletino della Società geologica Italiana, vol. 19, p. 362 (sep. 18).
- 1900. Cupularia umbellata Neviani, Brioizoi neogenici della Calabrie. Palaeontographia Italica vol. 6, p. 168 (regional bibliography).
- 1908. Cupularia umbellata Canu, Iconographie des Bryozoaires fossiles de l'Argentine, Anales del Museo Nacional de Buenos Aires, vol. 17, p. 274, pl. 5, figs. 4, 5 (bibliography).
- 1909. Cupularia umbellata Canu, Bryozoaires fossiles du Sud-Ouest de la France, Bulletin de la Société Geologique de France, ser. 4, vol. 9, pp. 448, 454, pl. 16, figs. 16, 17 (regional bibliography)
- 1909. Cupularia lowei Norman, On the Polyzoa of Madeira, Journal Linnean Society, vol. 30, p. 290, pl. 37, figs. 7-12.
- 1913. Cupularia umbellata Canu, Etude comparée des Bryozoaires Helvetiens de l'Egypte avec les Bryozoaires vivants de la Mediterranée et de la Mer Rouge: Mémoire de l'Institut Egyptien, vol. 6, fasc. 3, p. 205.

1913. Cupularia umbellata Canu, Contributions à l'étude des Bryozoaires fossiles, Bulletin Société Géologique de France, ser. 4, vol. 13, pp. 125, 127.

1914. Cupularia lowei Osburn, The Bryozoa of the Tortugas Islands, Florida, Publication No. 182 of the Carnegie Institution of Washington, p. 194.

1916. Cupularia umbellata Canu, Bryozoaries fossiles du Sud-Ouest de la France, Bulletin Société Géologique de France, ser. 4, vol. 15, p. 322.

1919. Cupularia umbellata Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications Carnegie Institution of Washington, No. 291, p. 85, pl. 1, figs. 5-7; pl. 2, figs. 17-21.

We are ignorant as to why Norman, who is a great lover of archaic names, has not preserved the name of Defrance. The figures published by this author and by D'Orbigny are excellent and leave no doubt as to the identity of the species. The fossils which are identified as above are rather well preserved and their determination is easy. The pores of the hydrostatic zooecia are not radicular.

Variations.—In this species the two oral condyles are developed and united in a manner to produce a pivot serving as a support to the hinge of the opercular valve. The spinous processes number from seven to eight; they are united on the greater part of their length and form a false cryptocyst perforated with very irregular holes; at their base there is no union and they thus limit a certain number of opesiules arranged all over the zooecium.

As Smitt thought in 1872, this species differs only from the species of the denticulata group in a greater calcification and in a more complete development of the spinous processes. In spite of its superficial appearance it therefore belongs to the same genus.

The specimens from Santo Domingo, where the species had been noted by Busk in 1859, are quite vigorous. They represent a variation that is remarkable in the size of the zooccia and in the aspect of the inner side. The latter does not show the usual tuberose ribs and the tuberosities are equally distributed on the zoarial surface. The ancestrula is not always visible; it is often covered over by a normal zooccium or replaced by a special region in which the zooccia are arranged in contrary order.

Measurement.—Opesia
$$\begin{cases} ho = 0.12 \text{ mm.} \\ lo = 0.16 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.44 - 0.50 \text{ mm.} \\ lz = 0.34 \text{ mm.} \end{cases}$

Occurrence.—Lower Miocene (Gatun formation): Banana River, Costa Rica (common). Lower Miocene: Chipola River, Calhoun County, and Shoal River, Walton County, Florida (common). Lower Miocene: (Oak Grove); Oak Grove, Yellow River, Florida (common). Lower Miocene (Bowden horizon): Bowden, Jamaica: Cercado de Mao, Rio Cana, and Rio Gurabo, Santo Domingo (common). Miocene (Duplin marl): Wilmington and other localities in North Carolina and South Carolina (common). Pliocene of South Carolina and Florida.

This species is almost always associated with *Cupuladria canariensis* Busk, 1859. Like the latter it commences in the Lower Miocene and continues through the latter Miocene and Pliocene of the United States.

Geological distribution.—Aquitanian of Italy (Seguenza, Neviani), of Bordeaux (Canu); Burdigalian of Italy (Seguenza, Canu), of Bordeaux (Canu); Helvetian of Egypt, Bordeaux, and Touraine; Tortonian of Provence (Canu), of Italy (Seguenza); Plaisancian of England (Busk), of Italy (Manzoni); Astian of Italy (Neviani,

Canu), of Provence (Canu); Sicilian of Italy (Neviani); Quaternary of Italy (Seguenza), of Argentina (Canu).

Habitat.—Mediterranean; Atlantic, Canary Islands, and Florida. It is common in the Gulf of Gascony in the Miocene, but it has now disappeared from this region. The species has been dredged at a depth of 11 to 48 meters in America and from 81 to 113 meters at Madeira.

Plesiotypes.—Cat. Nos. 68506-68511, U.S.N.M.

CUPULARIA ROBERTSONIAE, new species.

Plate 34, figs. 5-7.

1908. Cupularia canariensis Robertson, The incrusting Cheilostomatous Bryozoa of the West Coast of North America, University of California Publications 11, Zoology, vol. 4, no. 5, p. 314, pl. 24, figs. 90, 91.

Description.—The zoarium is discoid, little convex. The zooccia are distinct, much elongated, lozenge shaped; their mural rim is thin, sharp. The cryptocyst is deep, flat, perforated in the middle by some irregular pores and laterally by seven large opesiules; the opesium is ogival, transverse, with a proximal concave border. The vibraculum is large, salient, auriform and provided with a very large opesium.

$$Measurements. — Apertura \begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.15 \text{ mm.} \end{cases}$$
 Zooecia
$$\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.30 - 0.35 \text{ mm.} \end{cases}$$

Affinities.—The few fossil fragments in our collections appear to agree with Cupularia canariensis Robertson, 1908, dredged in the Pacific off the shores of California. This species is very close to Cupularia umbellata Defrance, 1828; but differs in its larger and less numerous opesiules and in its smaller zoarium. It differs from Cupularia reussiana Manzoni, 1869, in its more numerous and smaller opesiules and in its flatter zoarium.

Occurrence.—Pleistocene: Santa Monica (Long Wharf Canyon), California (rare).

Habitat.—Pacific, off San Pedro (6 meters) and island of Santa Catalina, California.

Cotypes.—Cat. No. 68512, U.S.N.M.

Genus CALPENSIA Jullien, 1888.

1888. Calpensia Jullien, Mission scientifique du Cap Horn, VI, Zoologie, Bryozoaires, p. 78.

The eryptocyst is entirely developed; it bears two distinct secondary opesia [opesiules]. The operculum is semilunar. (After Jullien.)

The opesiular outgrowths join the lateral walls, forming a closed hollow on each side. A simple, feebly chitinized operculum. Numerous pores, but no spines. No ovicell. No avicularia. The distal wall consists of a basal horizontal and a frontal ascending part, the former being furnished with a narrow transverse group of small uniporous rosette plates [=septulae]. The distal half of each distal wall with a single multiporous plate. (Levinsen.)

Genotype.—Colpensia (Flustra) impressa Moll, 1803. Range: Auversian, Recent.

CALPENSIA IMPRESSA Moll, 1803.

Plate 1, fig. 4.

1803. Eschara impressa Moll, Die Seerinde aus der Ordnung der Planzenthiere, Wien, p. 57, pl. 11, fig. 9.

1847. Eschara nobilis Michelin, Iconographie zoophytologique, p. 329, pl. 79, fig. 1.

1854. Membranipora calpensis Busk, Catalogue of the Marine Polyzoa, British Museum, Cheilostomata, pt. 2, p. 60, pl. 104, fig. 5, 6.

1879. Micropora impressa Waters, On the Bryozoa of the Bay of Naples, Annals Magazine Natural History, ser. 5, vol. 3, p. 1230.

1902. Micropora impressa Calvet, Bryozoaires marins des cotes de Corse, Travaux de l'Institut de Zoologie de l'Universite de Montpellier, ser. 2, mem. 12, p. 17.

1910. Micropora impressa Canu, Bryozoairs fossiles de terrains du Sud. Ouest de la France, Bulletin Société Géologique de France, ser. 4, vol. 10, p. 844, pl. 16, fig. 6 (variety).

1911. Micropora impressa Mme. Guerin-Ganivet, Contribution a l'étude des Bryozoaires des cotes Armoricaines, I, Travaux scientifique du Laboratoire de Concarneau, vol. 3, p. 5, pl. 1

1919. Calpensia impressa Canu and Bassler, Geology and Palentology of the West Indies, Bryozoa, Publication of the Carnegie Institution of Washington, No. 291, p. 84, pl. 1, fig. 11.

The discovery of this species in America was a great surprise, since it has heretofore been noted only in the Mediterranean area, where its zoaria occur in great abundance, especially along the African coast. Our determination is nevertheless an exact one, as we possess so many specimens for comparison that we should not be mistaken.

The bibliography of this species given by Pergens, Jelly, and Waters is incorrect, for they have confounded many distinct species. Canu noted five of them in 1911. In spite of its antiquity there are no unquestioned references to the species other than those we have given above.

The zoarium may be incrusting or Eschara-like. We have never observed an ovicell—indeed, the species may have none.

Occurrence.—Oligocene (Antigua formation): Carlisle marl pit, Antigua, Leeward Islands (rare).

Geological distribution.—Helvetian of Brittany, France (Michelin); Sahelian of Oran (Canu collection); Sicilian of Italy (Neviani).

Habitat.—Mediterrançan: Gibraltar, Corsica, Tuscany, Naples, Adriatic, Aegean Sea, Tunis, Algeria, Morocco; Atlantic off Brittany. It abounds at a depth of 25–30 meters and is very common at the depths of 75–100 meters.

Plesiotype.—Cat. No. 68513, U.S.N.M.

Genus VERMINARIA Jullien, 1888.

1888. Verminaria Jullien, Mission scientifique du Cap Horn. VI, Zoologie, Bryozoaires, p. 78.

The cryptocyst bears many opesiules on each side. The opesium is subterminal. (After Jullien.)

Genotype.—Verminaria (Membranipora) oblonga Busk, 1859. Pliocene.

Genus MICROPORINA Levinsen, 1909.

1909. Microporina Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, p. 162.

The two opesiulae, which, however, are sometimes filled up, appear as simple perforations. Numerous pores. No ovicell, but avicularia with crossbar occur.

Each distal wall and the distal half of each lateral wall with a row (6-8) of one or two pored rosette plates [septulae]. A longitudinal series of parietal muscles is placed on each side between the cryptocyst and the covering membrane.

Genotype. — Microporina (Cellaria) borealis Busk and Microporina (Micropora)

elongata Hincks, 1880. Range: Campanian, Recent.

Homalostega amphora Marsson, 1887, from the Campanian of Rügen, also belongs to this genus.

Genus CORYNOSTYLUS Canu and Bassler, 1919.

1919. Corynostylus Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publication of the Carnegic Institution of Washington, No. 291, p. 86.

No ovicell. The opercular valve articulates on two condyles. The zooecia are club shaped and provided with a gymnocyst. The zoarium is articulated.

Genotype.—Corynostylus labiatus Canu and Bassler, 1919. Miocene.

This genus has no recent equivalent but its structure is easy to interpret. The two condyles serve as a hinge for the strongly chitinized opercular valve. The lower part of the opesium placed below served evidently as passage for the opesiular fibers attached to the ectocyst. The deep cavity of the cryptocyst served as a hydrostatic apparatus for the entrance and exit of the polypide. Like most of the articulated genera, the zoarium in this one probably was fastened on large mobile algae.

CORYNOSTYLUS LABIATUS Cann and Bassler, 1919.

Plate 2, figs. 11-13.

1919. Corynostylus labiatus Canu and Bassler, Geology and Paleontology of the West Indies.
Bryozoa, Publications Carnegie Institution of Washington, No. 291, p. 87, pl. 2, figs. 11-13.

Description.—The zoarium is articulated and formed of long regular segments. The segments are compressed, bilamellar, formed of three longitudinal rows of zooecia on each side. The zooecia are elongate, oval, distinct, rounded in front, narrowed behind and are provided with a convex gynmocyst with large pores; the mural rim is thick, regular, granular. The opesium is elongate, oval, provided with a proximal, salient lip placed between two rounded opesiular indentations. The two condyles are quite salient. The cryptocyst is deep and smooth.

Measurements.— Opesia
$$\begin{vmatrix} ho = 0.12 \text{ mm.} \\ lo = 0.10 \text{ mm.} \end{vmatrix}$$
 Zooceia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.26 - 0.28 \text{ mm.} \end{cases}$

Affinities.—The first zooecium of each segment is radicular; it gives rise to three polypidian zooecia.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (common).

Holotype.—Cat. No. 68514, U.S.N.M.

CORYNOSTYLUS ELLIPTICUS Cann and Bassler, 1919.

Plate 2, figs. 8-10.

1919. Corynostylus ellipticus Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, No. 291, p. 87, pl. 2, figs. 8-10.

Description.—The zoarium is articulated. The segments are formed of two longitudinal rows of zooecia placed only on one side of the zoarium. The zooecia

are distinct, club shaped, with long, convex, and porous gymnocyst. The mural rim is thin and granular. The opesium is elliptical, elongate, surrounded by a salient rim; the two condyles are large and median. The cryptocyst is deep, smooth, and small.

Affiinities.—This species differs from Corynostylus labiatus in its unilamellar segments with only two zooecial rows and in the absence of a salient lip proximal to the opesium. Only the fragments figured have been found; they are extremely fragile.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (rare).

Cotypes.—Cat. No. 68515, U.S.N.M.

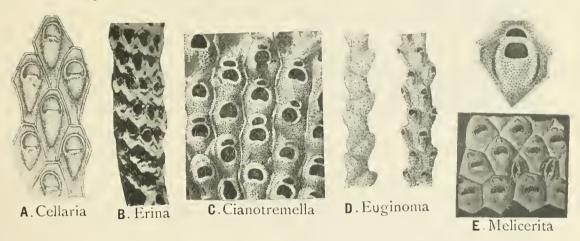


Fig. 14.—Genera of the family Cellariidae Hincks, 1880.

- A. Cellaria Authors. C. sinuosa Hassall, 1841. X 50. Recent. (After Hincks, 1880.
- B. Erina Canu 1908. E. patagonica Canu, 1908. Cretaceous. (After Canu, 1908.)
- C. Cianotremella Canu. 1911. C. gigantea Canu, 1911. Cretaceous. (After Canu, 1911
- D. Euginoma Jullien, 1882. E. rermiformis Jullien, 1882. \times 21. Recent. (After Jullien, 1882. Upper right-hand figure, zooecium enlarged, showing ovicell.
- E. Melicerita Milne-Edwards, 1838. M. charlesworthi Busk, 1859. X 25. Pliocene. (After Busk, 1859.)

Family CELLARIIDAE Hincks, 1880.

Genus CELLARIA Authors.

(For description see Bulletiu 106, U.S. National Museum, p. 272.)

CELLARIA FISSURIFERA, new species.

Plate 34, figs. 15-18.

Description.—The segments are cylindrical, slender at the base. The zooccia are rhomboidal, distinct, adjacent or partially separated by a very deep furrow: the mural rim is thin, salient, sharp, and ornamented with small tuberosities; the cryptocyst is smooth, deep, scarcely convex. The apertura is semilunar, sur-

rounded by a thin and salient peristome; the proximal lip is very convex and bears two very small lateral denticles. The orifice of the ovicell, which is endotoichal, is small. The zooecia bearing branches are narrow, with an elliptical elongate orifice. The interzooecial avicularium is elliptical or subrectangular; its opesium is irregular.

Measurements.—Apertura
$$\begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.17 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.85 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Affinities.—In the determination of Cellaria, one must take into consideration the zooecia bearing branches which are somewhat equivalent to the "basis ramae" of Crisia. Their orifice serves as a place for a horny cylindrical joint.

This species differs from *Cellaria mandibulata* Hincks, figured by Miss Robertson, 1905, in the small round orifice of the ovicell and in the form and size of its zooecia bearing branches, which are narrow and provided with an elongate orifice and not transverse, and in the absence of large avicularian zooecia.

Occurrence.—Pleistocene: Santa Monica, California (rare). Cotypes.—Cat. No. 68516, U.S.N.M.

CELLARIA MANDIBULATA Hincks, 1882.

Plate 34, figs. 11-14.

1882. Cellaria mandibulata Hineks, Polyzoa of the Queen Charlotte Islands, Annals and Magazine Natural History, ser. 5, vol. 10, p. 462; 1884, ser. 5, vol. 13, p. 203, p. 9, fig. 7.

1905. Cellaria mandibulata Robertson, Nonincrusting Cheilostomatous Bryozoa of the West Coast of North America, University of California Publications, Zoology, vol. 4, no. 5, p. 288, pl. 15, figs. 87, 89; pl. 16, fig. 103.

$$\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.07 \text{ mm} \\ la = 0.14 \text{ mm}. \end{cases}$$
 Zooecia
$$\begin{cases} Lz = 0.60 \text{ mm} \\ lz = 0.39 \text{ mm}. \end{cases}$$

Variations.—The orifice of the ovicell is quite variable; sometimes it is an ellipse, elongate and narrow; more often it is an orbicular orifice. The two forms may exist on the same segment. The avicularian zooccia are enormous and comparable to those which are observed on the recent specimens. The reader will comprehend their function by studying Miss Robertson's figure 88. The orifice of the zooccia bearing branches is round, surrounded by a salient peristome.

Occurrence.—Pleistocene: Los Angeles, California (very common). Habitat.—Pacific Ocean off San Pedro and San Diego, California. Plesiotypes.—Cat. No. 68517, U.S.N.M.

CELLARIA DIFFUSA Robertson, 1905.

Plate 34, figs. 19, 20.

1905. Cellaria diffusa Robertson, Nonincrusting Cheilostomatous Bryozoa of the West Coast of North America, University of California Publications, Zoology, vol. 4, no. 5, p. 289, pl. 15, fig. 90; pl. 16, fig. 104.

We have found very few fragments of this large and superb species. We have observed the same proximal tongue in the orifice of the ovicell, the same proximal convexity of the apertura, and the same rectangular interzooccial avicularium. Our micrometric measurements would be slightly larger if the scale of Miss Robertson be exact.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon) and Los Angeles, California (rare).

Habitat.—Pacific Ocean off California. Plesiotype.—Cat. No. 68518, U.S.N.M.

Genus ERINA Canu, 1908.

1908. Erina Canu, Iconographie des Bryozoaires fossiles de l'Argentine, pt. 1, Anales del Museo Nacional de Buenos Aires, vol. 17, p. 273.

The opesium is round, without oral denticles.

Genotype.—Erina patagonica Canu, 1908. Patagonian.

Genus MELICERITA Milne-Edwards, 1836.

1836. Meliccrita Milne-Edwards, Recherches anatomiques sur les Eschares, Annales des Sciences Naturelles, Zoologie, ser. 2, vol. 6, p. 26.

The zoarium is bilamellar and not articulated.

Genotype. — Melicerita charlesworthi Milne-Edwards, 1836. Astian.

Genus EUGINOMA Jullien, 1882.

1882. Euginoma Jullien, Dragages du Travailleur: Bryozoaires, Bulletin Société Zoologique de France, vol. 7, p. 520.

The zooecia are all turned forward and arranged in longitudinal rows; they are hexagonal, limited by a suture in relief; the orifice is semicircular, deprived of spines; the ovicell is formed by the raising of the wall of two zooecia superior to that which bears it; it is divided into two halves by the sutural line of these zooecia; its opening, like a policeman's helmet, is entirely above and in front of the orifice. Dorsal face of the zoarium is divided into trapezoidal areas which are the dorsal faces of two zooecia. (After Jullien.)

Genotype.—Euginoma vermiformis Jullien, 1882. Recent.

THE COSTULAE.

Family CRIBRILINIDAE Hincks, 1880.

Genus CRIBRILINA Gray, 1848.

(For description see Bulletin 106, U.S. National Museum, p. 290.)

CRIBRILINA PUNCTATA Hassall, 1841.

Plate 15, fig. 11.

- 1841. Lepralia punctata Hassall. Supplement to Catalogue of Irish Zoophytes, Annals and Magazine Natural History, vol. 7, p. 368, pl. 9, fig. 7.
- 1889. Cribrilina punctata Jelly, A Synonymic Catalogue of the Recent Marine Bryozoa, p. 67 (synonymy).
- 1901. Cribrilina (Arachnopusia) punctata Neviani, Briozoi neogenici della Calabrie, Paleontographia Italica, vol. 6, p. 174 (local bibliography).
- 1904. Lepralia marylandica Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 425, pl. 117, fig. 2.
- 1905. Cribrilina (Arachnopusia) punctata Neviani, Briozoi fossili di Carrubare (Calabria), Bollettino Societa Geologica Italiana, vol. 23, p. 523.
- 1907. Cribrilina punctata Calvet, Bryozoaires, Expedition scientifique du Trapailleur et du Talisman, p. 398 (bibliography).
- 1912. Cribrilina punctata Osburn, The Bryozoa of the Woods Hole Region, Bulletin Bureau Fisheries, vol. 30, 1910, p. 232, pl. 24, fig. 41 (bibliography).

This species lives mainly in the temperate and frigid regions and does not approach the Tropics. Its presence in the Miocene at Cove Point, Maryland, where it was described as *Lepralia marylandica* by Ulrich and Bassler, is very remarkable and would indicate the existence of a very cold boreal current.

Occurrence.—Miocene (St. Mary's formation): Cove Point, Maryland (rare). Geological distribution.—Pliocene and Quaternary of Austria-Hungary (Busk,

Bell) and of Italy (Seguenza).

Habitat.—Arctic Ocean: Jean Mayen (140–180 meters), Franz Joseph Land (210 meters); Sea of Kara; North Sea; Norway; Germany; Denmark (9–25 meters). Eastern Atlantic off England in the English Channel, in the Gulf of Gascony. Mediterranean (57–77 meters) and Adriatic. Madeira Islands. Western Atlantic from the St. Laurence to the Woods Hole Region.

Plesiotype.—Cat. No. 68519, U.S.N.M. (Holotype of Lepralia marylandica Ulrich and Bassler.)

CRIBRILINA LIGULATA, new species.

Plate 15, fig. 14.

Description.—The zoarium is encrusting a Pecten. The zooecia are distinct, separated by a deep furrow, clongated, regularly elliptical; the frontal is convex and bears 15 narrow, little salient costules; the lacunae are rectangular and larger on the margin than on the median zooecial axis. The apertura is large, tranverse, semilunar but with a concave proximal border; the peristome is wide, smooth, little salient; it bears four spines which can be transformed into two or three tongues by coalescence. The ancestrula is membraniporoid.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.30 - 0.32 \text{ mm.} \end{cases}$

Affinities.—In its exterior aspect this species much resembles Lepralia manzonii Reuss, 1874, from the Tortonian of Austria-Hungary, but it differs in the presence of its liguliform spines. Normally there are only four large spines, but they become thickened easily and joined together to form two or three very salient tonguelike plates.

Occurrence.—Miocene (Calvert formation): 1 mile south of Parkers Creek. Calvert County, Maryland (very rare).

Holotype.—Cat. No. 68520, U.S.N.M.

CRIBRILINA CUSPIDATA, new species.

Plate 15, fig. 15.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a deep furrow, large, elongate; the frontal is very convex; it is surrounded by a line of small lacunae and ornamented with 5 or 6 large lacunae in the form of a crescent. The lumen is not visible. The apertura is semilunar; the anter is very large and the poster is concave. The ovicell is hyperstomial. Between the zooecia are large triangular avicularia in the form of the head of a lance; the beak is rounded and turned toward the top.

Measurements.—Apertura |ha=0.17 mm|. Zooecia |Lz=0.85 mm|. |lz=0.35-0.50 mm|.

Affinities.—This species is very little distinct from Escharipora mucronata Smitt, 1872, formerly dredged in the waters off Florida. It is distinguished from it in its larger and less salient mucro and in having more than three lacunae on the frontal.

The species of Smitt has not yet been rediscovered; we are ignorant, therefore, of its variations. It is hardly probable that the frontal pores would always be so constant as they are drawn on Smitt's figures, but as we are not able to judge the nature of the variations we are obliged to create a new species which will become perhaps in the future simply a variety. Only the figured specimen has been found.

Occurrence.—Miocene: Santiago, Cuba (very rare).

·Holotype.—Cat. No. 68521, U.S.N.M.

Genus PUELLINA Jullien, 1886.

(For description see Bulletin 106, U. S. National Museum, p. 293.)

PUELLINA HERRMANNI Gabb and Horn, 1862.

Plate 35, figs. 2, 3.

1862. Reptescharella herrmanni Gabb and Horn, Monograph Fossil Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy of Natural Sciences, Philadelphia, ser. 2, vol. 5, p. 137, pl. 19, fig. 20.

Description.—The zoarium incrusts shells. The zooecia are large, elongate, elliptical, distinct, separated by a furrow and little convex. The costules are wide, flat, numbering from eight to ten; the lacunae are small and increase in size from the talon toward the zooecial axis. The apertura is semilunar, transverse, with a rectilinear proximal border.

$$\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.20 \text{ mm.} \end{cases}$$
 Zooecia
$$\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$$

Affinities.—The trace of the five oral spines as figured by Gabb and Horn is sometimes visible. In its exterior aspect and the width of the costules, this species is similar to *Cribrilina reniformis* Ortmann, 1890, and it differs from it only in the micrometric measurements if the magnification indicated by this author is exact.

Occurrence.—Pleistocene: Santa Barbara, California (very rare).

Plesiotype.—Cat. No. 68522, U.S.N.M.

PUELLINA RADIATA forma SCRIPTA Reuss, 1847.

Plate 15, fig. 12; plate 35, fig. 1.

1847. Cellepora scripta Reuss, Die fossilen Polyparien des Wiener Tertiärbeckens, Haidinger's Naturwissenschaftliche Abhandlungen, vol. 2, p. 82, pl. 9, fig. 28.

$$\label{eq:measurements} \begin{aligned} \textit{Measurements.} - \text{Apertura} & \begin{cases} ha = 0.06 \text{ mm.} \\ la = 0.08 \text{ mm.} \end{cases} & \text{Zooecia} & \begin{cases} Lz = 0.46 - 0.50 \text{ mm.} \\ lz = 0.24 - 0.30 \text{ mm.} \end{cases} \\ & \text{Apertura of ovicelled zooecia} & \begin{cases} ha = 0.08 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{cases} \end{aligned}$$

Variations.—The variations figured by the authors are very great; they depend on the number of the costules and on their width and also on the size of the avicularia. The photographs of our specimens are quite similar to the figures of Reuss

and Manzoni; but the dimensions are generally larger than in Eocene examples, all of them corresponding to specimens dredged in the Mediterranean.

The operculum closes the ovicell. The latter is often keeled. The apertura of the ovicelled zooccia is always larger. The number of costules is never greater than 12 to 14. There are always five spines.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (rare). Miocene (Yorktown formation): 1 mile west of Fort Nonsense, Gloucester County, Virginia (rare). Pleistocene: Santa Barbara, California (rare).

Plesiotype.—Cat. Nos. 68523-68525, U.S.N.M.

PUELLINA RADIATA forma RARECOSTA Reuss, 1847.

1847. Cellepora rarecosta Reuss, Die fossilen Polyparien des Wiener Tertiärbeckens, Haidinger's Naturwissenschaftliche Abhandlungen, vol. 11, p. 82, pl. 10, fig. 4.

This form is recognizable by the small number and prominence of the costules. It accompanies the typical form of the species at many recent and fossil localities.

Occurrence.—Pleistocene: Santa Monica, California (rare).

Plesiotypes.—Cat. No. 68526, U.S.N.M.

PUELLINA RADIATA CAROLINENSIS Gabb and Horn, 1862.

Plate 1, fig. 10.

1862. Reptescharella carolinensis Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences Philadelphia, vol. 5, p. 136, pl. 19, fig. 18.

1919. Puellina radiata carolinensis Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, No. 291, p. 90, pl. 1, fig. 12.

1920. Puellina radiata carolinensis Canu and Bassler, Mongraph North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 297, pl. 41, fig. 19.

This variety is characterized by its smooth peristome, without tuberosities, and by the small canal of the avicularium, which is larger than in the type.

In reality the spines have not disappeared; on the perfect specimens they are still visible, but they never have the size and importance of those which decorate the type. The zooecia are a little larger and have 16 costules.

Occurrence.—Jacksonian of South Carolina and Alabama; Vicksburgian of Alabama. Oligocene (Anguilla formation): Southwest side of Crocus Bay Bluff, 125 feet above sea level, Anguilla, Leeward Islands (rare). Oligocene (Antigua formation): Rifle Butts, Antigua, Leeward Islands (rare).

Plesiotype.—Cat. No. 68527, U.S.N.M.

PUELLINA INNOMINATA Couch, 1844.

Plate 15, fig. 13.

1844. Lepralia innominata Couch, Cornish Fauna, pt. 3, p. 114.

1900. Cribrilina radiata, var. innominata Neviani, Briozoi neogenici della Calabrie, Palaeonto-graphia Italica, vol. 6, p. 171 (regional bibliography).

1905. Cribrilina radiata, var. innominata Neviani, Briozoi fossili di Carrubare, Bollettino della Societa Geologica Italiana, vol. 23, p. 523 (sep. 21).

1909. Puellina innominata Norman, The Polyzoa of Madeira, Journal Linnean Society London, Zoology, vol. 30, p. 291 (zoological bibliography).

Variations.—The genus Puellina as limited by Jullien, 1886, and Levinsen, 1909, does not appear to us as well characterized as it is based on the presence of papillae whose physiologic function is not very evident. The relationship of the operculum to the ovicell constitutes for us generic characters of a greater importance in the Costulae as in the other Cheilostomata. It is in this feature that the present species is quite different from Puellina (Cribrilina) radiata, for the ovicell is never closed by the operculum. According to our views of classification it will be necessary to form a new genus for this type.

On our specimen the suboral pore is often replaced by a tubercle. The costules

are fine and the lacunae are arranged in concentric series.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee

River, 25 miles southwest of Tallahassee, Florida (very rare).

Geological distribution.—Helvetian of Italy (Seguenza); Astian of Italy (Seguenza); Sicilian of Italy (Seguenza, Neviani); Quaternary of Italy (Waters, Seguenza, DeStefani).

Habitat.—England, Mediterranean, Florida.

Plesiotype.—Cat. No. 6852\$ U.S.N.M.

PUELLINA CRASSILABIATA, new species.

Plate 29, fig. 7.

Description.—The zoarium incrusts oysters. The zooeeia are elliptical, regular, very little elongated, distinct, convex, separated by a furrow; the radial costules number 12 and taper toward the center, they are garnished with three large lumen pores and separated by four lacunae of diminishing size. The aperture is semilunar; the peristome is salient, very thin, and bears three hollow spines; a thick transverse lip borders the proximal edge of the apertura, between the two small oral avicularia. The ovicell is large, globular, salient, punctured by small, scattered pores; it opens by a special orifice in front of the oral lip.

Measurements.—Apertura
$$\begin{bmatrix} ha = 0.12 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{bmatrix}$$
 Zooecia $\begin{bmatrix} Lz = 0.50 \text{ mm.} \\ lz = 0.35 \text{ mm.} \end{bmatrix}$

Affinities.—The apertura of the ovicelled zooecia is wider and measures 0.12 mm. in width. The ovicell is not closed by the operculum and the passage of the eggs is protected by the wide oral mucro. This species does not belong, therefore, to the genus Puellina, since the two functions of the passage of the eggs and the escape of the larva operate in an absolutely different fashion than in Puellina radiata Moll, 1803. There are six dietellae.

In the presence of two small oral avicularia, it resembles the following species: It differs from Puellina puncturata Busk, 1859, in its elongate and nontransverse ovicell. It differs from Puellina calomorpha Reuss, 1866, in its apertura nontransverse and garnished with less than five spines. It differs from Puellina parisiensis Canu, 1917, in its thin nontransverse peristome, garnished with three spines and not with two tuberosities.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County. South Carolina (rare).

Holotype.—Cat. No. 68529, U.S.N.M.

Genus METRACOLPOSA Canu and Bassler, 1917.

(For description, see Bulletin 106, U. S. National Museum, p. 304.)

METRACOLPOSA MUCRONATA, new species.

Plate 35, fig. 4.

Description:—The zoarium incrusts shells. The zooecia are large, elongate, elliptical, distinct, convex, separated by a deep furrow. The costules, numbering 10 to 12, are wide, robust, convex, and bear some lumen pores; the lacunae are 4 in number and are small and orbicular. The apertura is oblique, elliptical, transverse; its proximal border bears a bifid, salient nucro. The ovicell is large, salient, convex; keeled, sunken in the distal zooecium; it opens in front of the mucro and can not be closed by the operculum.

$$\label{eq:measurements} \begin{split} \textit{Measurements.} - \text{Apertura} & \begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.18 \text{ mm.} \end{cases} & \text{Zooecia} & \begin{cases} Lz = 0.40 - 0.46 \text{ mm.} \\ lz = 0.34 - 0.36 \text{ mm.} \end{cases} \\ \text{Apertura of ovicelled zooecium} & \begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.22 \text{ mm.} \end{cases} \end{split}$$

Affinities.—The genus Metracolposa was discovered in the American Eccene formations, where it is widely represented by vigorous, bilamellar species. Its presence in the zone of the Pacific seems to indicate a great geographic extension in geologic time. Its presence even in the recent seas is quite probable. We should recall that the mucro of the Cheilostomatous zooecia is an apparatus of protection for the passage of the eggs.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Holotype.—Cat. No. 68530, U.S.N.M.

Family HIPPOTHOIDAE Levinsen, 1909.

Genus HIPPOTHOA (Lamouroux, 1821) Hincks, 1880.

(For description, see Bulletin 106, U. S. National Museum, p. 326.)

HIPPOTHOA HYALINA Linnaeus, 1768.

Plate 35, figs. 5-8.

- 1768. Cellepora hyalina Linnaeus, Systema Naturae, ed. 12, p. 1286.
- 1859. Lepralia hyalina Busk, Monograph Fossil Polyzoa of the Crag, Publications Paleontographical Society, London, vol. 14, p. 52, pl. 5, fig. 1.
- 1880. Schizoporella hyalina Hincks, British Marine Polyzoa, p. 271, pl. 18, figs. 8-10.
- 1889. Schizoporella hyalina Jelly, A synonymic Catalogue Recent Marine Bryozoa, p. 227 (bibliography).
- 1894. Schizoporella hyalina Levinsen, Mosdyr. Zoologica Danica (Danske Dyr), Hefte 9, p. 66, pl. 5, figs. 45-47.
- 1896. Celleporella hyalina Nordgaard, Systemetisk fortegnelse over de i Norge hidtil observer de, arter af marine polyzoa, I, Cheilostomata, Bergen's Musuem Aarbog for 1894-95, No. 2, p. 23.
- 1900. Hippothoa hyalina Waters, Bryozoa from Franz-Josef Land, Journal Linnean Society London, vol. 18, p. 70, pl. 8, figs. 16-18.
- 1900. Celleporella hyalina Nordgaard, Den Norske Nordnays-Expedition, Zoology no., vol. 17, p. 10.
- 1902. Schizoporella hyalina Calver, Bryozoaires marins de la region de Cette, Travaux de l'Institut Zoologie, Université Montpellier, ser. 2, Memoire XI, p. 44.

1903. Hippothoa hyalina Norman, Notes on the Natural History of East Finnmark, Annals Magazine Natural History, ser. 7, vol. 12, Polyzoa, p. 108.

1904. Schizoporella hyalina Calvet, Bryozoen, Ergebnisse der Hamberger Magalhaensiche Sammelreise, p. 25.

1905. Hippothoa hyalina Nordgaard, Hydrographical and biological investigations in Norwegian fiords, Bergen Museum, p. 165.

1906. Hippothoa hyalina Waters, Bryozoa from Chatham Island and d'Urville Island, New Zealand. Annals Magazine Natural History, ser. 7, vol. 17, p. 19.

1906. Celleporclla hyalina Nordgaard, Die Bryozoen des westlichen Norwegens, Bergen Musuem Meeres-fauna von Bergen, p. 87.

1907. Schizoporella hyalina Calvet, Bryozonires, Expédition scientifiques Travailleur et Talisman, p. 415.

1907. Hippothoa hyalina Nordoaard, Campagne arctique de 1907 de Duc D'Orleans. Bryozoaires, p. 9.

1908. Schizoporella hyalina Robertson, Incrusting Cheilostomatous Bryozoa of the West Coast of North America, University of California Publications, Zoology, vol. 4, No. 5, p. 289, pl. 19, fig. 43-45 (not synonomy).

1911. Schizoporella hyalina Guerin-Ganivet, Contributions a l'étude des Bryozoaires des cotes Armoricaines, II, Bryozoaires provenant de la rade de Brest, Travaux du Laboratoire de Zoologie de Concarneau, vol. 3, p. 2.

1912. Hippothoa hyalina Osburn, The Bryozoa of the Woods Hole Region, Bulletin Bureau Fisheries, vol. 30, 1910, p. 235, pl. 24, fig. 47 (American bibliography).

1912. Hippothoa hyalina Nordgaard, Revision av norske Bryczoer, Kgl. norske Videnskabers Selskabs Skriften, 1911, no. 3, p. 20.

1912. Schizoporella hyalina Guerin-Ganivet, Contributions a l'étude des Bryozoaires des cotes Armoricaines, III, Bryozoaires de la region de Concarneau, Travaux scientifiques du Laboratoire de Zoologie de Concarneau, vol. 4, p. 14.

1912. Schizoporella hyalina Barroso, Briozoos de la estacion maritima de Santander, Trabajos del Museo de ciencias naturales, no. 5, p. 16.

1913. Schizoporclla hyalina Guérin-Ganivet, Bryozoaires de la Mission arctique, Société d'oceanographique de Golfe de Gascogne, fasc. 7, p. 26.

A number of varieties of this widespread recent and fossil species have been described. Their present day arrangement is as follows:

Variety bouquinvillei D'Orbigny, 1839, is a distinct species.

Variety discreta Busk, 1852, is Diazeuxia reticulans Jullien, 1888.

Variety muricata Busk, 1852, is Diazeuxia kerquelenensis Jullien, 1888.

Variety cornuta Hincks, 1880, from Australia belongs to another genus.

Variety cornuta Hincks, 1880, from Natal is another species.

Variety from Santa Cruz (Hincks, 1880) is a distinct species.

Variety from Australia (MacGillivray) is a distinct species.

Variety from Australia (MacGillivray) is a distinct species.

Measurements.—Ordinary zooecia: Apertura
$$\begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{cases}$$
Zooecia
$$\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.14 \text{ mm.} \end{cases}$$
Ovicelled zooecia: Apertura
$$\begin{cases} ha = 0.06 \text{ mm.} \\ la = 0.08 \text{ mm.} \end{cases}$$
Zooecia
$$\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.20 \text{ mm.} \end{cases}$$
Male zooecia: Apertura
$$\begin{cases} ha = 0.04 \text{ mm.} \\ la = 0.04 \text{ mm.} \end{cases}$$
Zooecia
$$\begin{cases} Lz = 0.24 \text{ mm.} \\ lz = 0.12 \text{ mm.} \end{cases}$$

Variations.—The ordinary zooccia and the ovicelled zooccia develop on the same plane. The male zooccia have no fixed place and lap over very often on the others; when they are numerous and creet they give to the zoarium an aspect of Cellepora, which explains its generic name of Celleporella given by certain authors. The presence of three sorts of zooccia renders this species easy to determine; however, it is not rare to find zoaria uniquely formed of ordinary zooccia, in which case the determination is more difficult.

This species generally lives on algae and the depths indicated by the dredgings gives only information of secondary importance when the substratum is unknown. This is a species of the cold boreal zone and it never goes farther south than the forty-second parallel. We have found in the Pliocene of Monroe County, Florida, a mediocre specimen of a very closely related species. We will only give it a name if we have the chance to find better specimens. Generally its micrometric measurements are more reduced.

$$\label{eq:measurements} \begin{split} \textit{Measurements}. &-\text{Ordinary zooecia: Apertura} \left\{ \begin{array}{l} ha = 0.08 \text{ mm.} \\ la = 0.08 \text{ mm.} \end{array} \right. \\ & \text{Zooecia} \left\{ \begin{array}{l} Lz = 0.30 \text{ mm.} \\ lz = 0.20 \text{ mm.} \end{array} \right. \\ & \text{Male zooecia: } \quad \text{Apertura} \left\{ \begin{array}{l} ha = 0.04 \text{ mm.} \\ la = 0.04 \text{ mm.} \end{array} \right. \\ & \text{Zooecia} \left\{ \begin{array}{l} Lz = 0.16 - 0.20 \text{ mm.} \\ lz = 0.12 \text{ mm.} \end{array} \right. \end{split}$$

Occurrence.—Pleistocene: Santa Barbara (rare); Dead Man's Island off San Pedro (rare); Los Angeles and Santa Monica, California (rare).

Geological distribution.—Plaisancian of England (Busk).

Habitat.—Arctic Ocean: Iceland (162 meters); Franz Joseph Land (12-150 meters); Barents Sea (106 meters); Jean Mayen; Spitzberg; Greenland (4-48 meters); Sea of Kara; Nova Zembla (3-32 meters). North Sea: Norway; Germany; Denmark (8-27 meters). Eastern Atlantic: Off England, English Channel, Gulf of Gascony. Western Atlantic: Labrador, Woods Hole region. Eastern Pacific: Alaska, California.

Plesiotypes.—Cat. Nos. 68531, 68532, U.S.N.M.

HIPPOTHOA HYALINA, var. RUGOSA, new variety.

Plate 35, fig. 9.

Only the figured specimen has been found; it perhaps forms a distinct species. The large transverse wrinkles of the frontal are very characteristic. The species differ from *Hippothoa bougainvillei* D'Orbigny, 1839, in the absence of two large frontal beaks.

Occurrence.—Pleistocene: Santa Barbara, California (very rare). Holotype.—Cat. No. 68533,U.S.N.M.

Genus TRYPOSTEGA Levinsen, 1909.

(For description, see Bulletin 106, U. S. National Museum, p. 327.)

TRYPOSTEGA VENUSTA Norman, 1864.

Plate 16, fig. 1.

1920. Trypostega venusta Canu and Bassler, Monograph North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 330 p. 85, figs. 15, 16 (bibliography and description.).

The earliest appearance of this recent species in the Tertiary rocks of America is in the Vicksburgian of Mississippi and Alabama, where it is quite rare. The Miocene deposits of North Carolina have furnished a very few specimens which differ in no appreciable respect from the typical form.

Occurrence.—Miocene: (Duplin marl) Wilmington, North Carolina (very rare). Miocene (Yorktown formation): 1 mile west of Fort Nonsense, Gloucester County, Virginia (rare).

Geological distribution.—Miocene of Australia (Waters); Vicksburgian of the United States (Canu and Bassler).

Habitat.—Eastern Atlantic: English Channel, Madeira, Azores, Cape Verde Islands. Pacific: Lifu, Loyalty Island, Torres Straits. China Sea, Tozar Bank (43 meters). Indian Ocean: Saya de Malha (46–202 meters); Mauritius; Wasin, British East Africa (162 meters).

Plesiotype.—Cat. No. 68534, U.S.N.M.

Family ESCHARELLIDAE, Levinsen, 1909.

Group , SCHIZOPORELLAE Canu and Bassler, 1917.

Genus SCHIZOPORELLA Hincks, 1880.

As employed in our Monograph on the North American Early Tertiary Bryozoa, Schizoporella is retained for species showing no ovicell and which therefore can not be grouped in their proper place in the Schizoporellae.

SCHIZOPORELLA MAGNIPOROSA, new species.

Plate 45, figs. 1, 2.

Description.—The zoarium incrusts sponges. The zooecia are distinct, separated by a furrow, irregularly hexagonal; the frontal is little convex, porous, perforated by large, scattered tremopores. The apertura is formed of a semicircular anter and of a wide rounded rimule. Near the apertura there are one or two small setiform avicularia.

$$\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.18 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.65 \text{ mm.} \\ lz = 0.50 - 0.55 \text{ mm.} \end{cases}$$

Affinities.—Only the figured specimens have been found, and as they are incomplete and bear no ovicell it is impossible as yet to classify the species generically. The presence of small, oral, vibraculoid avicularia seems to indicate that this species belongs in reality to the genus Mastigophora.

The ancestrular zooecia are much smaller. The ancestrula bears a large elliptical aperture occupying almost all its frontal.

This species differs from Mastigophora porosa Smitt, 1872, in its larger tremopores and in the smaller apertura.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare).

Cotypes. - Cat. No. 68535, U.S.N.M.

SCHIZOPORELLA LATISINUATA Ulrich and Bassler, 1904.

Plate 18, fig. 4.

1904. Schizoporella latisinuata Ulrich and Bassler, Bryozoa: Maryland Geological Survey, Miocene, p. 421, pl. 119, fig. 2-4.

Description.—Zoarium forming thin parasitic expansions over foreign bodies. Zooecia sharply distinguished from each other, rather irregularly arranged, often subrhomboidal, four or five in 2 mm.; surface coarsely punctate. Orifice terminal, directed somewhat obliquely forward, subcircular, broadly sinuate proximally, enclosed by a slightly elevated peristome. Avicularia rather small, prominent, one on either or both sides of the orifice, rarely wanting, situated close to the peristome; apparently not divided by a septum. Ovicells not observed. (Ulrich and Bassler.)

$$\label{eq:measurements} \begin{split} & \textit{Measurements.} - \text{Apertura} \\ & \begin{array}{l} | ha = 0.10 \text{ mm.} \\ | la = 0.10 \text{ mm.} \\ | la = 0.40 \text{ mm.} \\ | la = 0.40 \text{ mm.} \\ | la = 0.40 \text{ mm.} \\ | la = 0.30 \text{ mm.} \\ \end{split}$$

Long zooecia
$$\begin{bmatrix} Lz = 0.60 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{bmatrix}$$

Affinities.—The aperture is as high as wide and buried at the base of a small peristome. The avicularium is infraoral; its beak is directed toward the median axis of the zooccia. The species has the exterior aspect of Stylopoma isabelleana Smitt, 1872, but differs in its larger avicularium and in the wider rimule of the aperture. We have not discovered the ovicell and are therefore not able to classify the species generically.

Occurrence.—Miocene (Choptank formation): Jones Wharf, Maryland (rare).

Holotype.—Cat. No. 68536, U.S.N.M.

SCHIZOPORELLA CUMULATA Ulrich and Bassler, 1904.

Plate 16, fig. 10.

1904. Schizoporella cumulata Ulrich and Bassler, Maryland Geological Survey Miocene, p. 422, pl. 17, fig. 7.

The original description is as follows:

Zoarium probably massive, composed of layers of zooccia arranged very irregularly and piled upon each other much as in Cellepora. Zooecia of irregular shapes, often broad ovate, convex, large, 0.5 mm. or 0.6 mm. in length and nearly 0.5 mm, in width. Orifice rounded, slightly transverse, the proximal side broadly notched; peristome somewhat elevated, thick. Surface distinctly and abundantly punctate. Avicularia rather variable in size, situated on only one or on both sides of the orifice, the acuminate end of the aperture elevated and turned outwardly. Ovicells not observed.

The general aspect of this bryozoan is decidedly like that of a Cellepora, and we can scarcely donbt that it is related to some of the species still referred to that genus. We have placed it under Schizoporella, not because we are satisfied that it really belongs there, but for the reason that its zooecial orifices and the avicularia are almost exactly as in other species (e.g., S. subquadrata and S. latisinuata) that we have referred to this genus. From these it is distinguished principally by the extremely irregular

arrangement and piling up of the zooccia.

In the absence of ovicells on the few specimens found we are unable to classify this species in the correct division of the Schizoporellae.

Occurrence.—Miocene (Choptank formation): Jones Wharf, Maryland (very rare). Miocene (St. Mary's formation): Cove Point, Maryland (rare).

Holotype.—Cat. No. 68537, U.S.N.M.

Genus ARTHROPOMA Levinsen, 1909.

(For description see Bulletin 106, U. S. National Museum, p. 351.)

ARTHROPOMA CORNUTA, new species.

Plate 16, fig. 3.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a deep furrow, little elongate, swollen; the frontal is convex covered by large tremopores and ornamented by a small salient median protuberance. The apertura is semilunar, transverse and bears on its rectillinear, proximal border a very small sinus.

$$\label{eq:measurements} \textit{Measurements.} - \textit{Apertura} \begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.14 \text{ mm.} \end{cases}$$
 Zooccia
$$\begin{cases} Lz = 0.70 \text{ mm.} \\ lz = 0.50 \text{ mm.} \end{cases}$$

Affinities.—The form of the rimule leaves no doubt regarding the arrangement of this species in the genus Arthropoma. It differs from the genotype Arthropoma cecili Audouin, 1826, only in the presence of the small frontal protuberance.

Occurrence.—Miocene (Choctawhatchee marl): Jaekson Bluff, Oeklocknee River, 25 miles southwest of Tallahassee, Florida (rare).

Holotype.—Cat. No. 68538, U.S.N.M.

Genus DAKARIA Jullien, 1903.

(For description see Bulletin 106, U.S. National Museum, p. 359.)

DAKARIA CHEVREUXI Jullien, 1903.

Plate 45, fig. 7.

1903. Dakaria chevreuxi Jullien, Bryozoaires provenant des campagnes de l'Hirondelle (1886-1888), Résultats des Campagnes scientifiques du Prince de Monaco, fasc. 23, p. 90, pl. 9, fig. 6.

We have found only a single specimen which is incrusting a coral, but in spite of its mediocre preservation we believe our determination to be exact. Each tremopore is surrounded by a small salient peristome. The oral avicularia are not constant. The proximal lip of the aperture ends in two condyles.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare).

Habitat.—Shores of Graciosa Isle, Azores.

Plesiotype.—Cat. No. 68539, U.S.N.M.

DAKARIA GRANDIS, new species.

Plate 20, fig. 16.

Description.—The zoarium incrusts shells. The zooecia are large, distinct, elongated, elliptical, separated by a deep furrow; the frontal is convex, ornamented with numerous small tremopores and minute granulations. The apertura is orbicular or somewhat transverse; two strong coundyles placed in the lower third limit the broad rounded sinus; the peristome is entire, thin, salient, finally denticulated.

Affinities.—This species is very close to the genotype Dakaria chevreuxi Jullien, 1904, but it differs in the absence of salient threads between the zooecia, which are also more elongated and especially in the larger micrometric dimensions (Lz=0.75 mm.).

Occurrence.—Miocene (Yorktown formation): Yorktown, Virginia (rare). Holotype.—Cat. No. 68540, U.S.N.M.

DAKARIA TORQUATA D'Orbigny, 1839.

Plate 12, figs. 8, 9.

1839. Escharina torquata D'Orbigny, Voyage dans l'Amerique-Meridionale, vol. 5, pt. 4, Zoophytes, p. 11, pl. 4, figs. 1-4.

1845. Escharina tumidula Lonsdale, Report on the Corals from the Tertiary formations of North America, Quarterly Journal Geological Society, London, vol. 1, p. 502. (Not Flustra torquata Lamouroux, 1827, Encyclopedie Méthodique, vol. 14, p. 407.)

1851. Cellepora subtorquata D'Orbigny, Paleontologie francaise, Terrain Cretace, vol. 5, Bryozoaires, p. 399.

1905. Cellepora subtorquata Waters, Notes on some Recent Bryozoa in D'Orbigny's Collection,
Annals Magazine Natural History, ser. 7, vol. 15, p. 6.

 $Measurements. - \text{Apertura} \begin{cases} ha = 0.15 \,\text{mm.} \\ la = 0.15 \,\text{mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.50 \,\text{mm.} \\ lz = 0.40 \,\text{mm.} \end{cases}$

Affinities.—In 1851 D'Orbigny having noted that his Escharina torquata was not Flustra torquata Lamouroux, 1827, changed its name. Examining the type of the species Waters, 1905, wrote "D'Orbigny's figure is good, but the lateral wings by the side of the sinus project more than is figured." In 1884 Hincks believed he had found D'Orbigny's species in the Pacific off California, but his good figure would not really permit this identification; the primitive name of Schizoporella dawsoni Hincks, 1883, applied by him to this species must therefore be preserved. The description and figure of Lonsdale leaves no doubt; we have indeed rediscovered his Escharina tumidula and in the same locality, namely, Petersburg, Virginia. D'Orbigny's figure of the species is much better than that of Lonsdale. This species differs from Schizobrachiella sanguinea Norman, 1868, and Dakaria chevreuxi Jullien, 1903, in a larger rimule and in the smaller micrometric dimensions. It incrusts shells and has no dietellae.

Occurrence.—Miocene (Yorktown formation): 3 miles southwest of Petersburg (rare) and 2 miles southwest of Cash, Gloucester County, Virginia (rare).

Habitat.—Atlantic: Bay of Rio de Janeiro, Brazil.

Plesiotypes.—Cat. No. 68541, U.S.N.M.

DAKARIA PARVIPOROSA, new species.

Plate 30, fig. 1.

Description.—The zoarium incrusts oysters. The zooecia are distinct, long, separated by a furrow; the frontal is convex and formed by a tremocyst with very small pores. The apertura is large, orbicular with a very wide rimule of little depth.

Measurements.—Apertura la=0.16 mm. Zooecia lz=0.50-0.60 mm. lz=0.36-0.40 mm.

Affinities.—This species differs from Schizoporella insculpta Hincks, 1883, only in the absence of the oral armature of the rimule and in the absence of a furrow near the apertura. The figured specimen is the only one that has been found; it lacks the ovicell and we can not make any close comparisons.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (very rare).

Holotype.—Cat. No. 68542, U.S.N.M.

Genus LACERNA Jullien, 1888.

(For description see Bulletin 106, U.S. National Museum, p. 345.)

LACERNA MUCRONATA Smitt, 1872.

Plate 16, fig. 2.

1872. Hippothoa mucroneta Smirr, Floridan Bryozoa, pt. 2, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 11, no. 4, p. 45, pl. 8, fig. 169.

Our determination of the American specimens is exact. We can observe on our specimens the same costules, the six spines, the wide rimule and the complete peristome indicated on Smitt's figure.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (very rare). Habitat.—Florida (47 meters).

Plesiotype.--Cat. No. 68543, U.S.N.M.

Genus STEPHANOSELLA Canu and Bassler, 1917.

(For description see Bulletin 106, U. S. National Museum, p. 343.)

STEPHANOSELLA BIAPERTA Michelin, 1842.

Plate 16, figs. 4-9.

Zoological bibliography.

1889. Schizoporella biapcrta Jelly, Synonymic Catalogue Recent Marine Bryozoa, p. 223 (bibliography).

1901. Schizoporella biaperta Whiteaves, Catalogue of the Marine Invertebrata of Eastern Canada, Geological Survey of Canada, Bull. no. 722, p. 100.

1907. Schizoporella biapcrta Calvet, Expedition scientifique Travailleur et Talisman, p. 419 (bibliography).

1908. Schizoporella biaperta Robertson, The Incrusting Chilostomatous Bryozoa of the West Coast of North America, University of California Publications, Zoology, vol. 4, p. 287, pl. 19, fig. 41.

1909. Schizoporella biaperta Norman, The Polyzoa of Maderia and neighboring Islands, Journal Linnean Society London, Zoology, vol. 30, p. 303, pl. 40, figs. 3, 4 (var. divergens).

1912. Schizoporella biaperta Osburn, The Bryozoa of the Woods Hole Region, Bulletin Bureau Fisheries, vol. 30, p. 237, pl. 29, fig. 49.

1912. Schizoporella biaperta Guerin-Ganivet, Contributions a l'étude des Bryozoaires des cotes Armoricaines, III, Bryozoaires de la region de Concarneau, Travaux scientifiques du Laboratoire de Zoologie de Concarneau, vol. 4, p. 13.

1912. Schizoporella biaperta Nordgaard, Campague arctique de 1907, de Duc d'Orleans, Bryozoaires p. 40.

1912. Schizoporella biaperta Barroso, Briozoos de la Estacion de Biologia maritima de Santander, Trabajos del Museo de Ciencias naturales, no. 5, p. 40.

1914. Schizoporella biapcrta Osburn, The Bryozoa of the Tortugas Islands, Publicatiou Carnegie Institution of Washington, no. 182, p. 207.

Paleontological bibliography.

1842. Eschara biaperta Michelin, Iconographie zoophytologique, p. 330, pl. 79, fig. 3.

1859. Lepralia biaperta Busk, Monograph Fossil Polyzoa of the Crag, Publications Paleontographical

Society, London, vol. 14, p. 47, pl. 7, fig. 5.

1862. Reptescharellina cornuta Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences of Philadelphia, ser. 2, vol. 5, p. 147, pl. 20, fig. 31.

1875. Lepralia biaperta Manzoni, I Briozoi del pliocene antico di Castrocaro, p. 21, pl. 2, fig. 28.

1878. Lepralia biaperta Waters, Bryozoa (Polyzoa) from the Pliocene of Bruccoli (Sicily), Transac-

tions Manchester Geological Society, vol. 14, p. 13, fig. 8.

1879. Lepralia biaperta Seguenza, Le formazioni terziarie nella Provincia di Reggio (Calabria), Reale Accademia dei Lincei, Memorie della Classe di Scienze Fisiche, Matematiche e Naturali, ser. 3, vol. 6, pp. 81, 202.

1887. Lepralia biaperta Waters, On Tertiary Bryozoa from New Zealand, Quarterly Journal Geological Society, London, vol. 43, p. 65.

1895. Schizoporella biaperta MacGillivray, Monograph Tertiary Polyzoa of Victoria, Transactions Royal Society of Victoria, vol. 4, p. 85, pl. 11, fig. 20.

1895. Schizoporella biaperta Neviani, Briozoi fossili della Farnesina e Monte Mario presso Roma, Paleontographica Italica, vol. 1, p. 110 (sep. 34).

1896, Schizoporella biaperta Neviani, Briozoi Postplioceniei di Spilinga (Calabria), Atti Accademia Gioenia di Scienze Naturali in Catania, ser. 4, vol. 9, p. 33, fig. 16.

1896–1898. Schizoporella biaperta Neviani, Briozoi neozoici di alcune localita d'Italia, Bollettino della Societa Romana per gli Studi Zoologica, pt. 3, p. 111 (sep. 10); pt. 5, pp. 4, 7.

1897. Schizoporella biaperta Neviani, Corallari e Briozoi neogenici di Sardigna, Bolletino della Società Geologica Italiana, vol. 15, p. 588 (sep. 20).

1901. Schizoporella biaperta Neviani, Briozoi neogenici della Calabrie Paleontographia Italiana, vol. 6, p. 195 (sep. 8) (regional bibliography).

1905. Schizoporella biaperta Neviani, Briozoi fossili di Carrubare (Calabria), Bollettino Societa Geologica Italiana, vol. 23, p. 532 (sep. 30).

1919. Stephanosella biaperta Canu and Bassler, Geology and Paleontology of the West Indies, Paleontology, Publications Carnegie Institution of Washington, no. 292, p. 91, pt. 1, figs. 16-18.

1920. Stephanosella biaperta Canu and Bassler, North American Early Tertiary Bryozoa, Bull 106, U. S. National Museum, p. 344.

 $\label{eq:measurements} \textit{Measurements.} - \Delta \text{pertura} \begin{vmatrix} ha = 0.10 \text{ mm.} \\ la = 0.10 \text{-}0.12 \text{ mm.} \end{vmatrix}$ Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Variations. This species is quite variable in its external aspects and in its micrometric measurements. The ovicelled zooecia are oriented in all directions; this absolutely inexplicable phenomenon characterizes this species and contributes much to making it often absolutely unrecognizable.

We give figures of specimens chosen from different formations which will suffice to enable the reader to avoid errors of determination. The specimens with large tremopores should possibly be separated as a variety, in which case Gabb and Horn's name of cornuta would be available. This is one of the few species common to both the Atlantic and Pacific.

Occurrence. Miocene (Bowden marl): Bowden, Jamaica (rare). Miocene (Duplin marl): Wilmington, North Carolina (rare). Pleistocene: Los Angeles (rare). Dead Mans Island off San Pedro (very rare), and Santa Monica, California (rare); Vero, Florida (rare).

Geological distribution.—Miocene of Australia (MacGillivray); Helvetian of France (Michelin), of Sardinia (Neviani), and Italy (Seguenza); Zanclean of Italy (Seguenza, DeStefani); Sahelian of Oran (Collection Canu); Plaisancian of England

(Busk), of Italy (Manzoni); Sicilian of Italy (Neviani, Waters); Quaternary of Italy (Neviani); Pliocene of New Zealand (Waters).

Habitat.—Arctic Ocean: Kara Sea, Spitzberg, Greenland, Jean Mayen (160–180 meters), Alaska (circumpolar species). Eastern Atlantic: North Sea, English Channel, England, Brittany, Gulf of Gascony (166 meters), Spain, Madeira (48 meters), Azores (130 meters). Western Atlantic: Woods Hole region (5–32 meters), Florida (14–97 meters). Eastern Pacific: Queen Charlotte Islands, California (213 meters). Western Pacific: Japan. Southern Pacific: Australia, New Zealand. Mediterranean: Nice, Naples, Corse (22–110 meters), Ajaccio (280 meters), Bonifacio (55–77 neters), Algeria (121 meters), and north of Morocco (370 meters).

Plesiotypes.—Cat. Nos. 68544-68546, U.S.N.M.

Genus STYLOPOMA Levinsen, 1909.

1909. Stylopoma Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, pl. 18 (name only).

1920. Stylopoma Canu and Bassler, Monograph North American Early Tertiary Bryozoa, Bull. 106, U. S. National Museum, p. 359.

The ovicell is hyperstomial; it covers the apertura and the avicularia entirely. The frontal is a tremocyst. The apertura is provided with a small linear rimule.

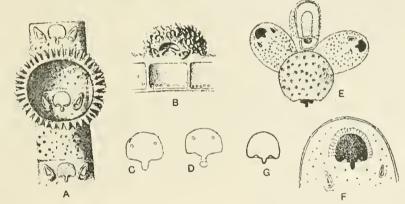


Fig. 15.—Genus Stylopoma Levinsen, 1909.

A.-H. Stylopoma spongites Pallas, 1766. A. Zooecium, × 40, with an ovicell, the frontal half of which is cut away. B. drawing, × 6, showing an ovicell from the proximal end and distal walls with septulae. C, D. Opercula, × 100. (A-D, after Levinsen, 1909.) E. Zooecia, × 25, showing ovicell and vicarious avicularium. F. A zooecium, × 85, showing the oral denticles from the front. G. Operculum, × 85. (E-G, after Waters, 1918.)

Genotype.—Stylopoma (Eschara) spongites Pallas, 1766. Range: Miocene-Recent.

Levinsen proposed this genus for Eschara spongites and Schizoporella longirostris Hincks, 1886, but having recognized that the size of the rimule upon which he based it is not a generic character, he suppressed the name. We reestablish the genus, basing it upon the function of the passage of the eggs, which operates in a very different manner from that in Schizopodrella, where the species would otherwise have been placed. The great development of the ovicell above the aperture assures this function. Schizoporella longirostris Hincks, 1886, has the same type of orifice, but not being provided with this ovicell it must be classified in Schizopodrella.

STYLOPOMA SPONGITES Pallas, 1766.

Plate 17, figs. 1-12.

- 1766. Eschara spongites Pallas, Elenchus Zoophytorum, p. 45.
- 1797. Cellepora spongites Esper, Die Pflanzenthiere, p. 242, pl. 111.
- 1803. Eschara spongites Moll, Die Seerinde aus der Ordnung der Pflanzenthiere, Wien, pl., figs. 3A, 3B.
- 1821. Cellepora spongites Lamouroux, Exposition methodique des genres de Polypiers, p.41, fig. 3.
- 1845. Cellepora informata Lonsdale, Report on the Corals from the Tertiary formations of North America, Quarterly Journal Geological Society London, vol. 1, p. 505 (text fig.).
- 1857. Reptocelleporaria informata Tuomey and Holmes, Pliocene Fossils of South Carolina, p. 15, pl. 4, figs. 11, 12.
- 1862. Reptocelleporaria informata Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences of Philadelphia, ser. 2, vol. 5, p. 132.
- 1872. Hippothoa spongites Smitt, Floridan Bryozoa, pt. 2, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 11, no. 4, p. 42, pl. 8, figs. 161–163.
- 1889. Schizoporella spongites Jelly, Synonymic Catalogue of Marine Bryozoa, p. 234 (bibliography).
- 1904. Schizoporella informata Ulrich and Bassler, Bryozoa, Maryland Geological Survey, Miocene, p. 419, pl. 114, figs. 6-10.
- 1909. Schizoporella spongites Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, p. 323, pl. 18, figs. 3, 4 (Stylopoma spongites on plate).
- 1914. Schizoporella spongites Osnurn, The Bryozoa of the Tortugas Islands, Publication Carnegie Institution of Washington, no. 182, p. 207.
- 1919. Stylopoma spongites Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 91, pl. 1, fig. 13.

Measurements. — Apertura
$$\begin{cases} ha = 0.14 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.36 \text{ mm.} \end{cases}$

Historical.—The bibliography published by Miss Jelly in 1889 was determined by Smitt, 1872, and appears inexact to us. This species is absolutely restricted to the Gulf of Mexico. By what chance could it have fallen into the hands of the eighteenth century authors? Certainly the species which they figured is the Schizopodrella unicornis Johnston, 1847, or Schizopodrella longirostris Hincks, 1886, which also present large multilamellar and spongy zoaria. We believe that the name of Lonsdale, 1845, would be preferable.

Variations.—The ovicell of this species is quite remarkable; a superb study was made by Levinsen, 1909. An ordinary polypide forms the zooccium. It degenerates and is replaced by a female polypide which forms the ovicell above the distal zooccium and the apertura. It is deprived of tentacles which could scarcely issue from the special orifice of the ovicell. The passage of the eggs is thus assured in a positive fashion. The escape of the larvae operates as in the species of the genus Schizopodrella by the rupture of the membrane which closes the ovicell.

The variations of this species are extraordinary. The avicularium is present or absent, large or small. The zooccia are long or short, narrow or swollen, even transverse. Frequently they are oriented in all directions.

The zoaria contain a large number of lamellae piled one on another; their dimensions reach a length of 5 centimeters. The larvae are fastened on shells, on other bryozoa, and more frequently on the fronds of small algae.

Occurrence.—Miocene (Yorktown formation): Yorktown and 3 miles southwest of Petersburg, Virginia (common). Miocene (Duplin marl): Wilmington, and other localities in North Carolina (common); Darlington, Muldrows Mills, 5 miles south of Maysville, and other localities, South Carolina (rare). Pliocene (Caloosahatchee marl): Shell Creek, De Soto County and Munroe County, Florida (common). Pleistocene: Simmons Bluff, Yonge's Island, Charleston County, South Carolina (rare); Mount Hope, Panama Canal Zone (rare). Pleistocene or Recent: Vero, Florida (rare).

Habitat.—Gulf of Mexico off Florida (21–56 meters), Tortugas (29 meters), Bermuda, St. Thomas, St. John, and perhaps at Malacca (Levinsen).

Plesiotypes.—Cat. Nos. 68547-68553, U.S.N.M.

STYLOPOMA PROJECTA, new species.

Plate 45, fig. 6.

Description.—Zoarium incrusting. Zooccia distinct, separated by a furrow, regularly oriented, sub-rectangular; frontal convex and garnished with numerous tremopores. Aperture small, transverse, with a small, narrow rimule; rounded at its extremity; peristome thick, salient, smooth. A small oral avicularium with pointed and very salient beak is adjacent to the rimule and to the aperture. The ovicell is enormous, globular ornamented with tremapores; it hides the aperture. The interzooccial avicularium is large, convex, with rounded and somewhat enlarged beak.

$$\label{eq:measurements} \textit{Measurements.} - \text{Aperture} \begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.11 \text{ mm.} \end{cases}$$
 Zooecia
$$\begin{cases} Lz = 0.50 - 0.55 \text{ mm.} \\ lz = 0.30 - 0.40 \text{ mm.} \end{cases}$$

Affinities.—This species is very well characterized by its small oral avicularium arranged to the right or to the left of the rimule, an arrangement not observed in other known species.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare). Plesiotype.—Cat. No. 68554, U.S.N.M.

STYLOPOMA MAGNIPOROSA, new species,

Plate 47, fig. 4.

Description.—The zoarium is bilamellar. The zooecia are large, elongated, distinct, separated by a furrow, elliptical; the frontal is convex and perforated by large expanded tremopores. The apertura is elongated; the anter is orbicular and very large; the poster is formed by a small narrow rectilinear rimule; the peristome is thick, little salient, smooth. On each side of the apertura there are two triangular oblique avicularia with the beak directed toward the base. The ovicell is enormous, globular, placed on the distal zooecium, covering and hiding the aperture.

Affinities.—This species differs from Stylopoma isabelleana D'Orbigny in its large frontal tremopores and in the beak of the avicularia directed toward the base. We have observed a giant aperture corresponding probably to a special or abnormal mandible.

Occurrence.—Miocene (Bowden horizon): Santo Domingo. Holotype.—Cat. No. 68555, U.S.N.M.

STYLOPOMA MINUTA, new species.

Plate 3, fig. 8.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by deep furrows, elongated, small; the frontal is convex and provided with small tremopores. The apertura is small, transverse, semielliptical with a narrow rectangular rimule on the proximal, rectilinear border; the peristome is thin, sharp, salient. The ovicell is very large, globular, provided with protuberances and entirely covers the apertura. In the vicinity of the apertura there is a small triangular avicularium with pivot, the beak of which is turned obliquely toward the top.

Measurements.—Apertura $\begin{cases} ha = 0.05 \text{ mm.} \\ la = 0.07 \text{ mm.} \end{cases}$ Zooecium $\begin{cases} Lz = 0.40 - 0.45 \text{ mm.} \\ lz = 0.25 - 0.30 \text{ mm.} \end{cases}$

This is the smallest known species of the genus *Stylopoma*. *Occurrence*.—Miocene (Bowden marl): Bowden, Jamaica (rare). *Holotype*.—Cat. 68556, U.S.N.M.

Genus SCHIZOPODRELLA Canu and Bassler, 1917.

(For description, see Bulletin 106, U.S. National Museum, p. 338.)

SCHIZOPODRELLA ACULEATA, new species.

Plate 31, figs. 13, 14.

Description.—The zoarium is massive, very large and formed of many superposed lamellae. The zooecia are little distinct, elongated, separated by an irregular furrow, elliptical or rectangular; the frontal is a tremocyst with large, widened pores. The apertura is suborbicular and bears a wide, rounded rimule of little depth; the peristome is thin, complete and salient on the zooecia of the internal lamellae. The ovicell is globular, salient, covered with tremopores. The avicularium with pivot is placed in the vicinity of the apertura; it is very long and terminates in a point.

Measurements.—Apertura $\begin{cases} ho = 0.15 \text{ mm.} \\ lo = 0.12 - 0.15 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.45 \text{ mm.} \\ lz = 0.25 \text{ mm.} \end{cases}$

Affinities.—This species differs from Schizoporella cumulata Ulrich and Bassler, 1904, in the length of its avicularium (L=0.40 mm.). It differs from Schizopodrella longirostris Hincks, 1886, which also bears a large avicularium, in the much greater width of its rimule. When the large avicularium is not much developed S. aculeata much resembles Stylopoma spongites Pallas, 1766; it differs from it in its rimule which is wide and concave and not small and rectangular and also in the different form of the ovicell. It differs from Schizopodrella unicornis Johnston, 1847, in which the apertura is identical, in the presence of a single very large avicularium.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (very rare).

Cotypes.—Cat. No. 68557, U.S.N.M.

SCHIZOPODRELLA MUTABILIS Canu and Bassler, 1919.

Plate 6, figs. 3-5.

1919. Schizopodrella? mutabilis Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 90, pl. 6, figs. 3-5.

Description.—The zoarium is unilamellar, tubular; it incrusts the fine radicells of algae. The zooccia are little distinct, elongate, elliptical; the frontal is little convex, surrounded by areolar pores and garnished with some large tremopores. The apertura is elongate, oval, with a wide rimule separated from the anter by two salient condyles.

Measurements.—Apertura $\begin{cases} ha = 0.18-0.20 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.70 \text{ mm.} \\ lz = 0.34 \text{ mm.} \end{cases}$

Affinities.—The very thick incrustation of the tremocyst renders the aspect of this species quite changeable and sometimes absolutely unrecognizable. We have not discovered the ovicell, so the classification naturally remains doubtful. It is possible that our specimens represent the incrusting base of some branching form like Gemelliporella punctata Canu and Bassler, 1919, but until the discovery of more specimens this point must remain in doubt.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (rare).

Holotype.—Cat. No. 68558, U.S.N.M.

SCHIZOPODRELLA UNICORNIS Johnston, 1847.

Plate 17, figs. 13, 14.

1847. Lepralia unicornis Johnston, History of British Zoophytes, ed. 2, p. 320, pl. 57, fig. 1.

1880. Schizoporella unicornis Hincks, British Marine Polyzoa, p. 238, pl. 35, figs. 1-5.

1889. Schizoporella unicornis Jelly, Synonymic Catalogue of Marine Bryozoa, p. 236 (general bibliography).

1904. Schizoporclla subquadrata Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 420, pl. 114, fig. 1; pl. 118, figs. 5, 6.

1906. Schizoporella unicornis Canu, Bryozoaires fossiles des Terrains du Sud-Ouest de la France, I Aquatanien, Bulletin de la Société géologique de France, p. 516 (biliography).

1907. Schizoporella unicornis Calvet, Expéditions scientifiques du Travailleur et du Talisman, vol. 8, p. 417 (bibliography).

1914. Schizoporella unicornis Osburn, The Bryozoa of the Tortugas Islands, Publications Carnegie Institute of Washington, No. IS2, p. 205 (American bibliography).

1918. Schizoporella unicornis Waters, Marine Fauna fo the Cape Verde Islands, Linnean Society's Journal Zoology, vol. 34, p. 14, pl. 2, figs. 14-17, 22 (bibliography).

1919. Schizopodrella unicornis Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publication of the Carnegie Institution of Washington, no. 291, p. 90.

This species may easily be confused with *Stylopoma spongites* Pallas, 1766; it is distinguished from it by the wide, proximal sinus of its apertura and by the quite different form of its ovicell.

S. unicornis is quite diffused through the European Tertiary formations, but on the contrary is quite rare in America, although its geologic distribution here appears to be almost identical.

Occurrence.—Lower Miocene: (Bowden formation): Bowden, Jamaica (rare). Miocene (Choptank formation): Governor Run, Maryland (rare). Miocene (Duplin

12184-23-Bull, 125--8

marl): Muldrows Mills, 5 miles south of Maysville, South Carolina (very rare); Wilmington, Natural Well, 2 miles southwest of Magnolia, etc., North Carolina (rare). Miocene (Yorktown formation): 1 mile west of Fort Nonsense, Gloucester County and other localities in Virginia (rare). Miocene (St. Mary's formation): Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia (rare). Pliocene (Caloosahatchee marl): Monroe County, Florida (rare). Pleistocene: Simmons Bluff, Younges Island, Charleston County, South Carolina; Daytona, Florida (rare).

Geologic distribution.—In Europe since the Stampian. Habitat.—Atlantic, Mediterranean, and boreal seas. Plesiotype.—Cat. Nos. 68559-68565, U.S.N.M.

SCHIZOPODRELLA FLORIDINA Osburn, 1914.

Plate 16, figs. 11-15.

1914. Schizoporella floridina Osburn, The Bryozoa of the Tortugas Islands, Florida, Publications Carnegie Institute of Washington, No. 182, p. 206, text figs. 17, 18.

Variations.—The zoarium is massive and formed of a number of superposed lamellae. Sometimes the zooecia are not oriented and are distributed in all directions. The large avicularium is very fragile and easily altered by fossilization. The aperture in our fossil specimens is not exactly similar to that figured by Osburn differing slightly in its rimule not placed at the level of the frontal. We have not observed moreover the large interzooecial avicularium.

Occurrence.—Miocene (Duplin marl): Wilmington and Lake Waccamaw, North Carolina (rare). Miocene (Choctawhatchee marl): Jackson Bluff, Ocklockonee River, 25 miles southwest of Tallahassee, Florida (very rare).

Cotypes.—Cat. Nos. 68566, 68567, U.S.N.M.

SCHIZOPODRELLA PUSILLA, new species.

Plate 17, figs 18, 19.

Description.—The zoarium is free and bilamellar or incrusting. The zooecia are small, little distinct, elongated, claviform; the frontal is somewhat convex, perforated by some large tremopores. The apertura is as high as wide; the rimule is wide, rounded, little deep. The ovicell is globular, hyperstomial. There are generally two small triangular avicularia with very salient beak placed on each side of the apertura.

 $\label{eq:measurements} Measurements. -- \text{Apertura} \begin{cases} ha = 0.10 - 0.12 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.54 \text{ mm.} \\ lz = 0.24 \text{ mm.} \end{cases}$

Affinities.—This spe ies has much resemblance to Schizopodrella patagonica Waters, 1905, in the arrangement of its avicularia; it differs from it in its larger avicularia, placed lower, salient and always inclined toward the aperture. The zooecia, although usually plain, are sometimes margined by a slightly salient thread.

Occurrence.—Miocene (Choctowhatchee marl): Jackson Bluff, Ocklockonee River, 25 miles southwest of Tallahassee, Florida (rare).

Holotype.—Cat. No. 68568, U.S.N.M.

SCHIZOPODRELLA MARGINATA, new species.

Plate 30, figs. 4, 5.

Description.—The zoarium incrusts oysters. The zooecia are distinct, oval, wide, bordered by a wide thread, little salient joined to the peristome; the frontal is little convex and ornamented with large tremopores. The apertura is orbicular; the rimule is wide, of little depth, triangular. The ovicell is little salient, much embedded in the distal zooecium with a frontal area. On each side of the apertura there are two small round avicularia without pivot.

$$Measurements. — Apertura \begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.08 - 0.10 \text{ mm.} \end{cases}$$
 Zooecia
$$\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.28 - 0.40 \text{ mm.} \end{cases}$$

Affinities.—This small species is quite well characterized and easy to determine. It differs from Schizopodrella pusilla, new species, in its great zooecial width and in the triangular form of the rimule.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare).

Holotype.—Cat. No. 68569, U.S.N.M.

SCHIZOPODRELLA DOVERENSIS Ulrich and Bassler, 1904.

Plate 18, figs. 1-3.

1904. Schizopodrella doverensis Ulrich and Bassler, Bryozoa, Geological Survey Maryland, Miocene, p. 421, pl. 117, fig. 1.

Description.—Zoarium a thin sheet on foreign bodies, the figured specimen growing on the reverse of Retepora doverensis. Zooecia well distinguished from each other but with the surface rather flat; irregularly arranged, though their elongate form gives some prominence to the longitudinal rows; average length 0.6 mm., width 0.35 to 40 mm. Orifice terminal, slightly transverse, broadly notched on the proximal side, on the whole nearly circular. Surface reticulate, only slightly convex, the central portion appearing flattened. Avicularia rather small, prominent, one on either or both sides of the orifice, rarely wanting, situated close to the peristome; apparently both divided by a septum. Ovicells not observed. (Ulrich and Bassler.)

Affinities.—The zooecia are not always regularly oriented. The zoarium incrusts shells or other bryozoa.

This species differs from Schizopodrella unicornis Johnston, 1847, whose apertura is identical, in its larger dimensions and in the absence of the frontal protruberance. It differs from Schizobrachiella (Escharella) sanguinea Smitt, 1872, in its single very large avicularium and in the smaller and more numerous tremopores. It differs from Stylopoma (Hippothoa) isabelleana Smitt, 1872, in the place of its avicularium not situated below the apertura.

Occurrence.—Miocene (Choptank formation): Dover bridge, Maryland (rare). Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida (very rare). Plesiotypes.—Cat. No. 68570, U.S.N.M.

Genus SCHIZOLAVELLA Canu and Bassler, 1920.

1920. Schizolavella Canu and Bassler, North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 358.

SCHIZOLAVELLA VULGARIS Moll, 1803.

Plate 35, fig. 10.

- 1803. Eschara vulgaris Moll, Die Seerinde, p. 55, pl. 3, fig. 10.
- 1867. Lepralia botterii Heller, Die Bryozoen des Adriatischen Meeres, Verhandlungen der k. k. zoologisch-botanischen Gesellschaft in Wien, vol. 7, p. 30, pl. 2, fig. 4.
- 1880. Schizoporella vulgaris Hincks, British Marine Polyzoa, p. 244, pl. 37, fig. 7; pl. 40, figs. 5, 6.
- 1880. Lepralia vulgaris Seguenza, Le formazioni terziarie della Provincia di Reggio (Calabria), Reale Accademia dei Lincei, ser. 3, vol. 6, pp. 202, 295, 369, (not Lepralia otophora of the same author).
- 21885. Pachykraspedon otophorum Koschinsky, Ein Beitrag zur Kenntnis der Bryozoenfauna der älteren Tertiärschichten des südlichen Bayerns, Paleontographica, vol. 32, p. 44.
- 1889. Schizoporella vulgaris Jelly, A Synonymic Catalogue of Recent Marine Bryozoa, p. 238 (bibliography) (not Lepralia cognata Reuss).
- 1889. Schizoporella vulgaris Pergens, Notes succinctes sur les Bryozoaires, du Miocène de la Russie méridionale, Bulletin des Séances de la Société Royale Malacologique de Belgique, vol. 24, p. 5.
- 1895. Schizoporella vulgaris Neviani, Briozoi fossili della Farnesina, Palaeontographica Italica, vol. 1, p. 113 (3), pl. 6, fig. 13.
- 1896. Schizoporella vulgaris Neviani, Briozoi Postpliocenici di Spilinga (Calabria), Atti Accademia Gioenia di Scienze Naturali in Catania, ser. 4, vol. 9, p. 32.
- 1896. Schizoporclla vulgaris Calvet, Campagne du Caudan, Bryozoaires, Annales de l'Université de Lyon, p. 258.
- 1898-1900. Schizoporcila vulgaris Neviani, Briozoi neozoici di alcune localita d'Italia, Bolletino della Società Romana per gli Studi Zoologici, pt. 4, vol. 5, p. 11; pt. 5, vol. 7, pp. 4, 7, 13; pt. 6, vol. 8, p. 67 (sep. 10), 1900.
- 1899. Schizoporella vulgaris Waters, Bryozoa from Madeira, Journal Royal Microscopical Society, pp. 11, 16, pl. 3, figs. 9-11.
- 1900. Schizoporella vulgaris Neviani, Briozoi neogenici della Calabria, Palacontographia Italica, vol. 6, p. 196 (sep. 82) (Bibliography regional).
- 1902. Schizoporella vulgaris Calvet, Bryozoaires marins des cotes de Corse, Travaux de l'Institut Zoologique de Montpellier, ser. 2, mem. 12, p. 21.
- 1902. Schizoporella vulgaris Calvet, Bryozoaries marins de la region de Cette, Travaux de l'Institut Zoologique de Montpellier, ser. 2, mem. 8, p. 41.
- 1903. Schizoporcila vulgaris Jullien, Bryozoaires provenant des Campagnes de l'Hirondelle, pp. 79, 136.
- 1905. Schizoporella vulgaris Neviani, Briozoi fossili di Carrubare (Calabria), Bollettino della Società Geologica Italica, vol. 21, pt. 1, p. 533 (sep. 31), fig. 12.
- 1907. Schizoporclla vulgaris Calvet, Expédition scientifiques du Travailleur et du Talisman, p. 417 (bibliography).
- 1909. Escharina vulgaris Norman, Polyzoa of Madeira, Linnean Society's Journal, vol. 30, p. 302.
- 1912. Schizoporclla vulgaris Canu, Etude comparée des Bryozoaires Helvétians de l'Egypte avec les Bryozoaires vivants de la Méditerranée et de la mer Rouge, Memoires de l'Institut Egyptien, vol. 6, p. 213, pl. 11, fig. 6 (palaeontological, illustrated bibliography).

This species has never been noted except in the eastern Atlantic from the English Channel to the Cape Verde Islands and in the Mediterranean. It is not known in the Pacific. However, Waters has noted the fossil form in the Miocene of Australia. Our specimen from the Pleistocene is undoubtedly of this species.

It is therefore probable that its geographic extension is greater than we had supposed hitherto.

Occurrence.—Pleistocene: Santa Barbara, California (very rare).

Geological distribution.—Stampian of Germany (Reuss); Casselian of Germany (Reuss); Miocene of Australia (Waters); Burdigalian of Gard (Canu collection); Helvetian of Italy (Seguenza, Neviani), of Gard, of Herault, and of Touraine (Canu collection); Tortonian of Italy (Seguenza), of Austria-Hungary (Reuss); Sahelian of Oran (Canu collection); Zanclean of Italy (Seguenza); Plaisancian of Italy (Manzoni, Neviani); Astian of Italy (Seguenza); Sicilian of Italy (Neviani, Waters); Quaternary of Italy (Neviani, Seguenza).

Habitat.—Eastern Atlantic; English Channel, Gulf of Gascony (155–180 meters) Madeira, Cape Verde Islands, Azore Islands (89–130 meters). Mediterranean, Black Sea, Adriatic, Corse (5–77 meters), Cette (20–40 meters), Oran (75–121 meters).

Plesiotype.—Cat. No. 68571, U.S.N.M.

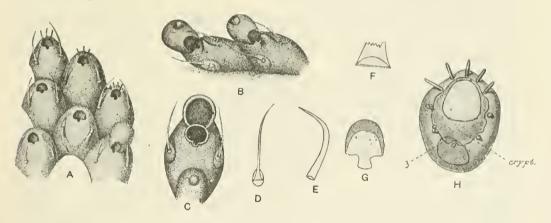


Fig. 16.—Genus Schizolavella Canu and Bassler, 1920.

A-H. Schizolavella vulgaris Moll, 1803. A. Portion of zoarium with an ovicelled zooecium × 25. B. Two zooecia, × 40, showing the umbonate ovicell and a mucro on the front wall. C. A zooecium, × 40, showing structure. D. Avicularian mandible. (A-D, after Hincks, 1880.) E. Avicularian mandible, × 85. F. Base of the avicularian mandible, × 250. G. Operculum, × 85. (E-G, after Waters, 1898.) H. Ancestrula. The region inside the marginal spines is partly occupied by a calcareous plate (crypt) which is probably a cryptocist; b. brown body (after Harmer, 1902).

Genus SCHIZOMAVELLA Canu and Bassler, 1920.

(For description see Bulletin 106, U. S. National Museum, p. 353.)

SCHIZOMAVELLA LONGIROSTRATA Hincks, 1883.

Plate 35, fig. 11.

1883. Schizoporella longirostrata Hincks, Report on the Polyzoa of the Queen Charlotte Islands, Annals Magazine Natural History, ser. 5, vol. 11, p. 447, pl. 17, fig. 4 (not Robertson, 1908).

Variations.—The rimule bears two salient condyles on which are supported the small proximal tongue of the operculum. The ovicell is globular, elongated, large; it bears in front a small concavity; it bears small tremopores like the frontal, it is closed by the operculum.

The large avicularium is not median; it is eccentric and curved.

The species which Miss Robertson has figured under the same name appears to be different for the apertura of the ovicelled zooccia is much larger than that of the ordinary zooccia. This is not the case in the figure of Hincks and of our specimen.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Habitat.—Queen Charlotte Islands.

Plesiotype.—Cat. No. 68572, U.S.N.M.

Genus GEMELLIPORELLA Canu and Bassler, 1920.

1920. Gemelliporella Canu and Bassler, North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 372.

The ovicell is hyperstomial and deeply embedded in the distal zooecium. The operculum does not close the ovicell and operates in a locella. The form of the apertura is like a keyhole. The frontal is garnished with lateral areolar pores and with a granular pleurocyst.

Genotype.—Gemelliporella vorax, new species. Range: Pliocene.

The genus Gemellipora Smitt, 1872, founded solely on the aspect of the apertura, is not a natural one. The first species described, G. eburnea, has been selected by Levinsen, 1909, as the type of the genus and classified in his family Liriozoidae. Gemellipora striatula Smitt, 1872, is a synonym for Trypostega venusta Norman, 1869. Gemellipora glabra Smitt appears to have a smooth frontal, but it is necessary to examine specimens anew. G. limbata Smitt is serial and appears to belong to the Phylactellidae. The keyhole form of the aperture does not appear to furnish a sufficient generic character because it does not correspond to an important modification of the hydrostatic function. In establishing the new genus Gemelliporella we have employed the characters furnished by the ovicell and the frontal.

GEMELLIPORELLA ASPER, new species.

Plate 18, figs. 5, 6.

Description.—The zoarium incrusts oysters. The zooecia are little distinct, irregularly elliptical; the frontal is small, hardly convex, surrounded by large, crowded, areolar pores. The apertura is deep, buried, elongate, shaped like a keyhole, with two lateral symmetrical condyles limiting the rimnle; the peristomice is elliptical and bears a little salient mucro. The ovicell is globular, salient, punctate; its orifice is very large and can not be closed by the operculum. On each side of the apertura there are two large orbicular avicularia with pivot, very salient; the one between them is much developed and becomes onychocelliform; its pivot bears a wide rectangular denticle.

Measurements.—Apertura $\begin{cases} ha = 0.14 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.40 - 0.50 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Affinities.—The avicularia by their saliency cover the zoarial surface with a great number of asperities which characterize this species, but which render it

very irregular. The denticle placed on the pivot of the large avicularia is rather peculiar to this species, but it is not constant; it corresponds perhaps to a particular arrangement of the rachis. The known recent bryozoa never have this arrangement. The number and importance of the avicularia indicate calm waters.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (rare). Pliocene (Waccamaw marl):

Waccamaw River, Horry County, South Carolina (very rare).

Cotypes.—Cat. Nos. 68573, 68574, U.S.N.M.

GEMELLIPORELLA VORAX, new species.

Plate 19, figs. 1-9.

Description.—The zoarium incrusts shells, oysters, and bryozoa. The zooccia are little distinct, separated by a furrow, irregularly elliptical; the frontal is convex, surrounded by some large areolar pores which are scattered and covered over with a pleurocyst more or less granular. The apertura is deep, oval, the point below with rimule wide and notched; the peristomice is semilunar with a proximal mucro more or less developed. The ovicell is convex, little salient, deeply embedded in the distal zooccium; its orifice is very wide and can not be closed by the operculum. On each side of the apertura there is a round avicularium; very frequently the one between them becomes very long and fusiform.

Measurements.—Apertura $\begin{vmatrix} ha = 0.10 \text{ mm.} \\ la = 0.08 \text{ mm.} \end{vmatrix}$ Zooecia $\begin{vmatrix} Lz = 0.40 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{vmatrix}$

Variations.—This species is exceedingly variable and takes the most fantastic aspects. The zooecia are only distinct on the small zoaria or on the margins of the large ones; at the center of the latter they are absolutely indistinct. The ancestrula is a small zooecium; it engenders two distal and four proximal zooecia. There are often interareolar costules. The reduction of its zooecial dimensions and the great development of the avicularia seems to indicate that this species required much oxygen.

It is remarkable that the species has disappeared from the recent Gulf of Mexico after its existence in the same region throughout the Miocene and Pliocene.

Occurrence.—Lower Miocene (Chipola marl): Chipola River, Calhoun County, Florida (rare). Miocene (Duplin marl): Wilmington and Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina (common); Muldrows Mills, 5 miles south of Maysville, South Carolina (rare). Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (rare). Miocene (Yorktown formation): Yorktown, 3 miles southwest of Petersburg, and other localities in Virginia (rare). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (common). Pliocene (Caloosahatchee marl): Shell Creek, De Soto County (rare), and Monroe County, Florida (common).

Cotypes.—Cat. Nos. 68575-68580, U.S.N.M.

GEMELLIPORELLA PUNCTAȚA Canu and Bassler, 1919.

Plate 5, figs. 7-9.

1919. Gemelliporella punctata Canu and Bassler, Biology and Paleontology of the West Indies, Bryozoa. Publications of the Carnegie Institution of Washington, No. 291, p. 92, pl. 5, figs. 7-9.

Description.—The zoarium is free, cylindrical, bifurcated. The zooccia are very little distinct, elongate, convex; the frontal is granular and surrounded with areolar

pores and is sometimes covered with *punctations* of pleurocystal origin. The aperture is oval, clongate, formed of a large semilunar anter and of a wide proximal sinus. The ovicell is deeply embedded in the distal zooccium, little salient, closed by the operculum, ornamented by an orbicular and very fragile frontal area.

 $\label{eq:measurements} \textit{Measurements.} - \textit{Apertura} \begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases} \quad \textit{Zooecia} \begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$

Affinities.—The punctations of the frontal are quite variable; there is not a single zooecium similar to another. In a better preserved state we think that each zooecium is surrounded with areolar pores irregular in size and spacing. The frontal is a thick and compact olocyst, covered by a pleurocyst which is granular or punctured with large pores which do not perforate the zooecial walls. Unfortunately we have not been able to confirm this exterior aspect by sections.

The possible relationship of this species to Schizopodrella mutabilis Canu and

Bassler, 1919, has been mentioned under the discussion of that species.

Occurrence.—Lower Miocene (Bowden formation): Bowden, Jamaica (rare). Cotypes.—Cat. No. 68581, U.S.N.M.

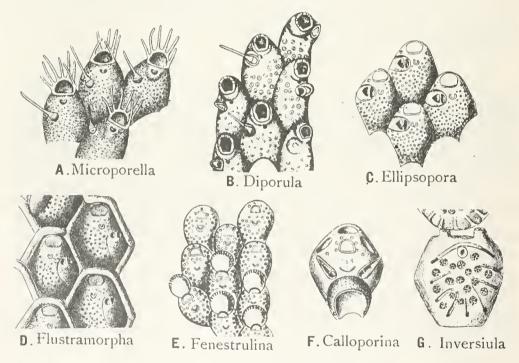


Fig. 17.—Genera of the Microporellae.

A. Microporella Hincks, 1887; M. eiliata Pallas, 1766, \times 43. B. Subgenus Diporula Hincks, 1880; D. verrucosa Peach, 1868, \times 25. C. Subgenus Ellipsopora, new; E. flabellaris Busk, 1852, \times 40. D. Subgenus Flustramorpha Gray, 1848; F. marginata Krause, \times 40. E. Fencstrulina Jullien, 1888; F. malusi Savigny-Audouin, 1826, \times 20. F. Calloporina Neviani, 1895; C. decorata Reuss. 1847, \times 40. G. Inversiulu Jullien, 1888; I. inversa Waters, 1889, \times 50.

Group 2. MICROPORELLAE Canu and Bassler, 1917.

(See Bulletin 106, U. S. National Museum, p. 417, for description and illustration).

In this group the orifice of the compensatrix (frontal pore, micropore or ascopore) is distinct and removed from the apertura. The ovicell is hyperstomial and always closed by the operculum. Dietellae are present.

Historical.—The presence of a frontal micropore was considered by Hincks as an important family character, but the study of the larvae does not permit the adoption of his conclusion. According to Jullien, the ascopore is the orifice of a horn-shaped body, allowing the exterior to communicate with the tentacular sheath. Harmer and Levinsch believed that this was an optical illusion and that the ascopore was really the orifice of the compensatrix.

Levinsen, 1909, did not recognize the different genera established by authors according to the form of the apertura. We again follow the same principles of generic distinction by considering the great variations of the function of calcification. Many species considered at first to belong to the Microporellae have since been recognized as true representatives of the Adeonidae. According to the nature of the calcification and the form of the aperture, we class the following genera in this group:

Fenestrulina Jullien, 1888. Mieroporella Hincks, 1877. Calloporina Neviani, 1895. Inversiula Jullien, 1888.

Genus FENESTRULINA Jullien, 1888.

1888. Fenestrulina Jullien, Mission scientifique du Cap Horn, 1882-83, VI, Zoologie, Bryozoaires, p. 37.

The frontal is garnished with stellate tremopores. The operculum closes the ovicell. No avicularia. Fourteen to fifteen tentacles.

Genotype.—Fenestrulina (Cellepora) malusi Savigny-Audouin, 1826.

Range.—Helvetian-Recent.

The known species belonging to this genus are:

Fenestrulina (Cellepora) malusi Audouin, 1826.

Fenestrulina (Microporella) proxima Waters, 1904.

Fenestrulina (Microporella) exigua Waters, 1904.

Fenestrulina (Mieroporella) parvipora Waters, 1904.

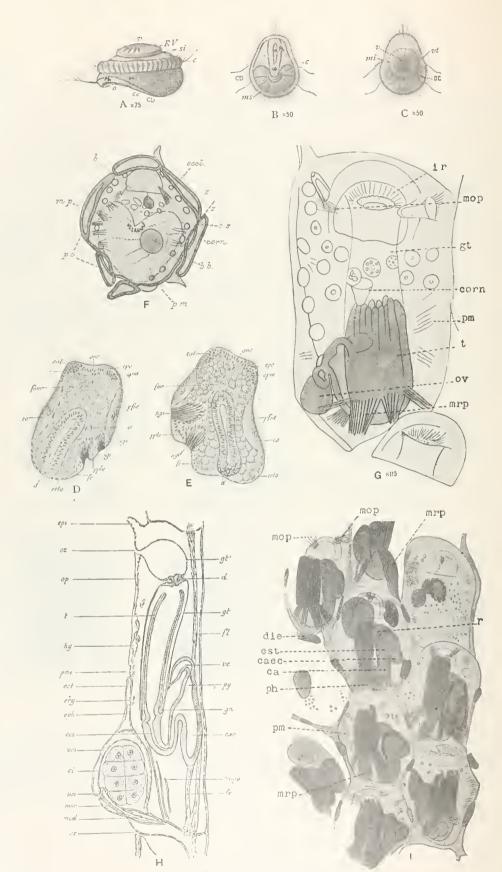


Fig. 18. Anatomy of the Microporellae.

Fig. 18.—Anatomy of the Microporellae.

- A-D. Microporcila ciliata Pallas, 1766. A. Profile view of free larva, × 75. B. Free larva, oral view, showing the large radiating elements of the oral mesoderm, × 50. C. Free larva, aboral view, showing radiating aspect of the calotte, 50. (A-C after Barrois, 1877.) c, corona; cal, calotte (terminal bud); CD, digestive cavity; cc, obscure part between the two branches of the stomach; mi, aboral mesoderm; ms, oral mesoderm; O, ciliated cleft; oc, oculiform points; ph, pharynx; RV, border of the calotte. D. Median sagittal section of an embryo in one of the last stages of development. (After Calvet, 1900.)
- E-I. Fenestrulina malusi Audouin, 1826. E. Median sagittal section of an embryo just before it emerges. (After Calvet, 1900.) In the course of the neuro-muscular bundles passing from the central nerve organs to the pyriform organ some cellules are seen. The internal sack has a simple form almost regularly cylindrical. c, mantle; cal, calotte (terminal bud); co, corona; d, tampon or inner sac; ecto, ectoderm; epc, ectoderm thickening; epm, mesoderm thickening; fe, ciliated cleft; fum, neuro-muscular bundle; one, central nervous organ of the embryo; pfse, sub-ectoderm nerve plexus; pplv, papilla of the vibratile plume; si, internal sac; sqs, superior glandular system; sqi, inferior glandular system. F. Basal view of a zooecium which has lost its polypide. (After Harmer, 1902.) b, polypide bud; bb, brown body; corn, cornicula; cs. compensatrix opening by the ascopore (m. p.); occl, operer muscles; p. c., dietellae; z, neighboring zooccia. G. Anterior view of a zooccium containing only a young polypide, × 115. (after Jullien, 1888.) ca, cardiac region of the stomach; caec, caecum of the stomach; corn, cornicula; die, dietella; est, stomach; qt, tentacular sheath; ir, irisoid; mop, occlusor muscles of the operculum; mrp, large retractor muscles of the polypide; ov, ovary; ph, pharynx and esophagus; pm, parietal muscles; r, intestine and rectum; t, tentacles. H. Longitudinal section of a bryozoid. (After Calvet, 1900.) caec, stomachic caecum; ci, incubation eavity; cry, cryptocyst, or skeleton; d, diaphragm; eph, hypostegal epithelium; epi, spine; fe, central funicular cord; fl, lateral funicular cord; gn, nervous ganglion; qt, sub-diaphragm region of the teutacular sheath; qt', subdiaphragm region; hy, hypostege; mud, dilator muscles of the incubation cavity; mugr, large retractor muscles; mur, retractor muscles of the frontal walls of the lower ovicellarian vesicle; ocs, esophagus; op, operculum; oz, zooecial orifice; pme, median frontal pore; py, pyloris; re, rectum; t, tentacles; voi, lower ovicellarian vesicle; vos. upper ovicellarian vesicle. I. Zoarium decalcified, seen posteriorly, × 45. (See G. for explanation of letters.)

FENESTRULINA MALUSI Savigny-Audouin, 1826.

Plate 36, figs. 2, 3.

1809. Cellepora malusii Savigny, Description de l'Egypte, Polypes, pl. 8, fig. 8.

1826. Cellepora malusii Audouin, Explication des planches de Savigny, p. 239.

1862. Cellepora californiensis Gabb and Horn, Monograph of the fossil Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy Natural Sciences Philadelphia, ser. 2, vol. 5, p. 130, pl. 19, fig. 12.

1895. Microporella malusi MacGillivray, Monograph of the Tertiary Polyzoa of Victoria, Transactions Royal Society of Victoria, vol. 4, p. 65, pl. 9, fig. 1 (regional bibliography).

1895. Microporella (Fenestrulina) malusi Neviani, Briozoi fossili delle Farnesina, Palacontographia Italica, vol. 1, p. 104 (sep. 28).

1895-1896. Microporella (Fenestrulina) malusi Neviani, Briozoi neozoici, di alcune localitia d'Italia, Bollettino della Societa Romana per gli Studi Zeologici, pt. 1, p. 115 (sep. 7); pt. 2, p. 229 • (sep. 5); pt. 3, p. 109 (sep. 8).

1898. Microporella malusi De Angelis, Los primeros antozoos y briozoos miocenicos recogidos en Cataluña, p. 22.

1901. Microporella (Fenestrulina) malusi Neviani, Briozoi neogenici della Calabric, Palaeontographia italica, vol. 6, p. 175 (sep. 61) (Regional bibliography).

1904. Microporella malusi Canu, Les Bryozoaires du Patagonien, Memoires de la Societe geologique de France, vol. 12, No. 33, p. 11, pl. 3, fig. 27.

1905. Microporella (Fenestrulina) malusi Neviani, Bryozoi fossili de Carrubare (Calabrio), Bollettino delle Societa geologica italiana, vol. 23, p. 524, fig. 8.

1907. Microporella malusi Calvet, Expeditions scientifiques du Travailleur et du Talisman, p. 404 (bibliography).

1908. Microporella malusi Canu, Bryozoaires fossiles de l'Argentine, Anales del Museo Nacional de Buenos Aires, vol. 16, p. 280.

1908. Microporella malusi ROBERTSON, The incrusting cheilostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 4, no. 5, p. 282, pl. 18, figs. 35, 36.

1909. Fenestrulina malusi Norman, The Polyzoa of Madeira, Journal Linnean Society, Zoology, vol. 30, p. 297.

1910. Microporella malusi Waters, Reports on the marine biology of the Sudanese Red Sea, Journal Linnean Society of London, Zoology, vol. 32, p. 168.

1912. Microporella malusi Barroso, Briozoos de la estacion maritima de Sautander, Trabajos del Museo de ciencias naturales, no. 5, p. 27.

It is remarkable that this very cosmopolitan species has never been observed in the western Atlantic, fossil as well as recent.

 $\label{eq:measurements} \textit{Measurements.} - \textit{Apertura} \begin{cases} ha = 0.08 - 0.10 \, \text{mm.} \\ la = 0.16 - 0.18 \, \text{mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.50 \, \text{mm.} \\ lz = 0.36 \, \text{mm.} \end{cases}$

Occurrence.—Pleistocene: Santa Barbara (common), and Dead Mans Island, off San Pedro, California (very rare).

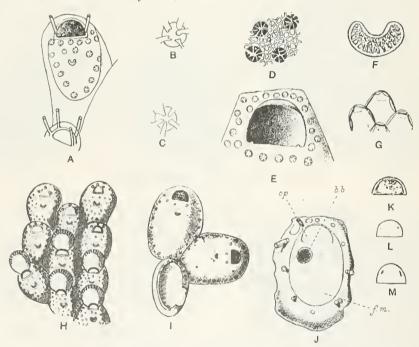


Fig. 19.—Genus Fenestrulina Jullien, 1888.

A-J. Fenestrulina malusi Savigny Audouin, 1826. A. A zooecium × 50 showing all the characters. B, C. Stellate frontal porcs, × 250. (A-C, after Waters, 1903.) D. Portion of surface of a zooecium, × 175. E. Aperture, × 100. (After Levinsen, 1909.) F. Detailed structure of the ascopore (fenestrule of Jullien) × 175. G. Dorsal face of the zooecia, showing the dietellae × 18. H. Zooecia × 21, several showing the ovicell. (D, F-H, after Levinsen, 1894.) I. Ancestrula and ancestrular zooecia, × 25. J. Ancestrula. The frontal membrane is typically Flustrine, its calcareous margin bearing ten spines, of which three are oral spines; op, operculum; b. b., brown body; f. m., frontal membrane (=ectocyst). (After Harmer, 1902.)

K. Operculum, × 85. (After Waters, 1877.) L. Fenestrulina parvipora Waters, 1903. Operculum, × 85. M. Fenestrulina proxima Waters, 1903. Operculum, × 85. (I, L, M, after Waters, 1903)

Geological distribution.—Burdigalian of Catalogne (De Angelis); Patagonian of Argentina (Canu); Miocene of Australia (MacGillivray) and New Zealand (Waters); Plaisancian of England (Busk) and of Italy (Manzoni); Astian of Italy (Seguenza); Sicilian of Italy (Seguenza, Neviani); Quaternary of Italy (Seguenza, Neviani).

Habitat.—Arctic Ocean: Finmark (16–48 meters). North Sea: Norway, Denmark (29–45 meters), England. Eastern Atlantic: England, English Channel, Gulf of Gascony (10–180 meters), Madeira. Mediterranean: Cette, Corse, Naples, Oran. Adriatic (32–89 meters). Southern Atlantic: Tristan da Cunha (228–243 meters), Falkland Islands (8–34 meters). Eastern Pacific: Queen Charlotte Islands, California (32–48 meters). Southern Pacific: Tahiti, Australia (5–13 meters), New Zealand. Western Pacific: Japan, Cape Tizard (China Sea) (44 meters), Indian Ocean, Red Sea. Antartic Ocean: Cape Horn (5–300 meters), Malouines Islands, Terra del Fuego.

Plesiotypes.—Cat. No. 68582, U.S.N.M.

FENESTRULINA POROSA, new species.

Plate 37, fig. 4.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a deep furrow, very little elongated; the surface is convex, very porous and perforated in its middle portion by a crescentric ascopore. The apertura is semilunar, transverse, surrounded by a little salient, very thin peristome. The ovicell is globular, covered with scarcely visible tuberosities and surrounded by a salient ring.

Affinities.—The peristome frequently bears two spines. The frontal is often covered with a double pellicule.

This species is very close to *Fenestrulina malusi*, but differs from it in its greater number of frontal pores and in its triple instead of double rows of pores present between the aperture and the ascopore.

Occurrence.—Pleistocene: Santa Monica (Long Wharf Canyon), California (rare).

Holotype.—Cat. No. 68583, U.S.N.M.

Genus MICROPORELLA Hincks, 1877.

1877. Microporella Hincks, On British Polyzoa, Annals Magazine Natural History, ser. 4, vol. 20, p. 526.

The operculum closes the ovicell and is semielliptical. The frontal is an ordinary tremocyst. Avicularia are present. Thirteen to fourteen tentacles.

Genotype.—Microporella (Eschara) ciliata Linnaeus, 1759.

Range.—Miocene-Recent.

Under this well-known genus we recognize three subgenera, as follows:

Diporula Hincks, 1879, characterized by a horseshoe-shaped orifice, slightly contracted by two lateral projections.

Ellipsopora, new subgenus, characterized by a transverse, elliptical apertura.

Flustramorpha Busk, 1884, characterized by the presence of radicular fibers and vibracula.

The known species belonging to this genus and its subgenera are as follows:

Microporella ciliata Linnaeus, 1759.

Microporella flabelligera Levinsen, 1909.

Microporella divaricata Waters, 1903.

Microporella hyadesi Jullien, 1888.

Microporella personata Busk, 1852.

Microporella rudis McGillivray, 1895.

Microporella rugosa Maplestone, 1900.

Microporella marginata Maplestone, 1909.

Microporella coronata Audouin, 1896.

Mieroporella (Flustramorpha) marginata Krauss, 1837.

Microporella (Diporula) verrucosa Peach, 1868.

Microporella (Diporula) hastigera Busk, 1884.

Microporella (Ellipsopora) flabcllaris Busk, 1852.

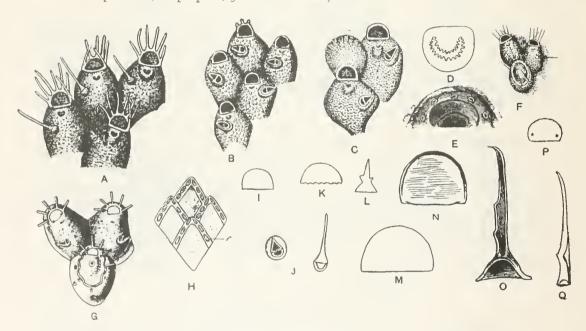


Fig. 20.—Genus Microporella Hineks, 1877.

A-I. Microporella ciliata Pallas, 1766. A. Zooecia with spines and mandible, × 43. B. Zooecia with avicularia, × 29. C. Ovicelled zooecia, × 40. E. Aperture, × 200. F. Ancestrula and ancestrular zooecia, × 29. (A-C, after Hineks, 1880.) G. The same part of the zoarium, × 55. (E, F, after Levinsen, 1909.) H. Zooecia from the basal surface, × 26, showing the dietellae. (After Levinsen, 1894.) I. J. Opercula and mandibles. (After Waters, 1877, and Hineks, 1880.)

D. Structure of the ascopore of Microporella hyadesi Jullien, 1888.

K-Q. Opercula and mandibles. K, L. Microporella hyadesi Jullien, 1888. (After Waters, 1877.)
M. Microporella divaricata Canu, 1902. (After Waters, 1903.) N, O. Microporella personata Busk, 1852.
(After Busk, 1885.) P, Q. Microporella coronata Audouin, 1826. (After Waters, 1909.)

MICROPORELLA CILIATA Linnaeus, 1759.

Plate 20, figs. 1-6; plate 36, figs. 4, 5.

1759. Cellepora ciliata LINNAEUS, Systema Naturae, ed. 12, p. 1286.

1889. Microporella ciliata Jelly, A synonymic catalogue of marine Bryozoa, p. 179 (bibliography).

1891. Microporella ciliata Neviani, Briozoi postpliocenici di Livorno, Bollettino della Societa geologica italiana, vol. 10, p. 117 (sep. 21).

1895. Microporella (Fenestrulina) ciliata Neviani, Briozoi fossili della Farnesina Palaeontographia italica, vol. 1, p. 105 (sep. 29), pl. 5, fig. 25.

1895-1900. Microporella (Fenestrulina) ciliata Neviani, Briozoi neozoici di alcune localita d'Italia, Bollettino della Societa Romana per gli Studi Zoologici, pt. 1, vol. 4, p. 7; pt. 2, vol. 4, p. 234 (sep.10); pt. 3, vol. 5, p. 122 (sep. 21); pt. 5, vol. 7, pp. 13, 15; pt. 6, vol. 8, p. 3, 9, 10.

1896. Microporella (Fenestrulina) ciliata Neviani, Briozoi postpliocenici di Spilinga, Atti Academia Gioenia di Scienze Naturali in Catania, vol. 9, p. 22.

1900. Microporella (Fenestrulina) ciliata Neviani, Briozoi neogenici della Calabrie, Palaeontographia italica, vol. 6, p. 176 (sep. 62) (regional bibliography).

1900. Microporella (Fenestrulina) ciliata Neviani, Briozoi terziari et posterziari della Toscana, Bollettino della Societa geologica italiana, vol. 19, p. 368 (sep. 19).

1904. Microporella inflata Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 416, pl. 110, fig. 7.

1904. Microporella praeciliata Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 415, pl. 110, fig. 6, pl. 113, fig. 3.

1905. Microporella (Fenestrulina) ciliata Neviani, Briozoi fossili di Carrubare (Calabria), Bollettino della Societa geologica italiana, vol. 23, p. 324 (sep. 22).

1907. Microporella ciliata Calvet, Expeditions scientifiques du Travailleur et du Talisman, p. 403, (regional bibliography).

1908. Microporella ciliata Waters, The Bryozoa of the Sudanese Red Sea, Journal Linnean Society, Loudon, vol. 31, p. 143.

1911. Microporella ciliata Guerin-Ganivet, Bryozoaires provenant de la Rade de Brest, Travaux du Laboratoire de Zoologie de Concarneau, p. 2.

1912. Microporella ciliata Guerin-Ganivet, Contributions a l'etude des Bryozoaires des côtes armoricaines, III, Travaux scientifiques du Laboratorie de Zoologie de Concarneau, p. 12.

1912. Microporella ciliata Osburn, Bryozoa of Woods Hole Region, Bulletin Bureau of Fisheries, vol. 30, p. 233, pl. 24, fig. 44; pl. 30, fig. 90.

1912. Microporella ciliata Barroso, Briozoos de la estacion de biologia maritima de Santander, Trabajos del museo de ciencias naturales, p. 26.

1912. Microporella ciliata Osburn, Bryozoa from Labrador, Newfoundland, and Nova Scotia, Proceedings U. S. National Museum, vol. 43, p. 279.

1913. Microporella ciliata Guerin-Ganivet, Bryozoaires de la mission arctique commandee par Ch. Bénard, Société d'oceanographie du Golfe de Gascogne, vol. 7, p. 23.

1913. Microporella ciliata Canu, Contribution a l'etude des Bryozoaires fossiles, IV, Pliocene d'Alger, Bulletin Société Geologique France, vol. 13, p. 125.

1915. Microporella ciliata Barroso, Contribucion al conocimiento de los Briozoos marinos de España, Boletin de la Sociedad española de Historia natural, vol. 15, p. 414.

1915. Microporella ciliata Waters, Marine biology of the Sudanese Red Sea, Journal Linnean Society, London, vol. 31, p. 443.

1917. Microporella ciliata Barroso, Notas sobre Briozoos, Boletin de la R. Sociedad española de Historia natural, vol. 17, p. 4.

The great variations in the calcification of the frontal in this species have been known for a long time. They have been carefully noted by the zoologists and depend solely on the nature of the water and the age of the zoarium. When the granulations are important, the tremopores appear very small and even may disappear leaving the frontal smooth (forma *inflata*). If the granulations are little

developed, the tremopores appear large (forma praeciliata). A specimen from Carteret County, North Carolina, is very suggestive. The incrusting portion shows normal zooecia with granulations and tremopores (fig. 1); a bilamellar expansion shows, on the contrary, zooecia without granulations but with large tremopores (fig. 2). A specimen from Santa Barbara, California, has very small avicularia as in Microporella umbonata, but nevertheless its dimensions are absolutely identical with Microporella ciliata.

 $Measurements. - Apertura \begin{bmatrix} ha = 0.04 - 0.06 \text{ mm.} \\ la = 0.10 - 0.12 \text{ mm.} \end{bmatrix}$ Zooecia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

The coast of California is the Elysium of the genus *Microporella*, for a wealth of species occurs there. They are very closely related and their determination is not made without difficulty because of their polymorphism. It is necessary especially to take into consideration the reciprocal place occupied by the avicularium and the ascopore; the micrometric measurements are of value only in extreme cases.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Ocklockonee River, 25 miles southwest of Tallahassee, Florida (rare). Miocene (St. Mary's formation): Cove Point, Maryland (rare). Miocene; Kuhns, Carteret County, North Carolina (rare). Miocene (Choptank formation); Jones Wharf, Maryland (rare). Pleistocene: Santa Monica (rare); Dead Mans Island, off San Pedro (rare); and Santa Barbara, California (rare).

Geological distribution.—Stampian of Germany (Schreiber); Chatian of Germany (Reuss); Miocene of Australia (Waters), of Tunis (Canu); Burdigalian of Gard in France (Canu collection); Helvetian of Italy (Seguenza Neviani), of France (Canu collection); Tortonian of Italy (Seguenza), of Austria-Hungary (Reuss); Sahelian of Oran (Canu collection); Zanclean of Italy (Seguenza); Pliocene of New Zealand (Waters); Plaisancian of Algeria (Canu), of Italy (Manzoni), of England (Busk); Astian of Italy (Seguenza); Sicilian of Italy (Seguenza, Manzoni, Neviani), of Rhodes (Manzoni); Quaternary of Italy (Waters, Seguenza, Neviani, De Stefani).

Habitat.—Cosmopolitan.

Plesiotypes.—Cat. Nos. 68584-68590, U.S.N.M.

MICROPORELLA HEXAGONA, new species.

Plate 20, figs. 14, 15.

Description.—The zoarium incrusts shells. The zooecia are little distinct, separated by a furiow, short, hexagonal, convex, gibbose; the frontal is convex, irregular, and formed of a tremocyst with large pores detachable from a subjacent, finely perforated olocyst. The apertura is deeply buried, semilunar; the peristome is somewhat salient, very thin, and bears four to six large, hollow spines. The ovicell is little salient and is covered by an incomplete tremocyst. The ascopore is large and is surrounded by a very salient peristome. There are two small avicularia or a single one very large and salient.

Measurements.—Apertura $\begin{bmatrix} ha = 0.07 \text{ mm.} \\ la = 0.12 \text{ 0.15 mm.} \end{bmatrix}$ Zooecia $\begin{bmatrix} Lz = 0.45 - 0.50 \text{ mm.} \\ lz = 0.45 \text{ mm.} \end{bmatrix}$

Affinities.—The intensity of the calcification is remarkable in this species and gives to it an aspect very characteristic and difficult to figure. The large avicularium is triangular, the beak pointing upward; the pivot is almost always broken. The distal border of the apertura is often denticulated by the rupture of the hollow spines much wider than the peristome. In spite of its double protective envelope the ovicell is often broken. The exterior aspect is absolutely unique. This species must have lived in a strong current.

Occurrence.—Miocene (Duplin marl): Darlington Courthouse, South Carolina

(rare).

Holotype.—Cat. No. 68591, U.S.N.M.

MICROPORELLA BIFOLIATA Ulrich and Bassler, 1904.

Plate 20, figs. 7-11.

1904. Microporella? bifoliata Ulrrich and Bassler, Maryland Geological Survey, Miocene, p. 417, pl. 113, figs. 6-8.

The original description is as follows:

Zoarium erect, bifoliate, not known to branch. Zooecia subquadrate or hexagonal, arranged in regular longitudinal and diagonally intersecting series, four longitudinally and five diagonally in 2 mm. Appearance of surface varying with age. In young examples the zooecia are more or less convex and separated, especially transversely, by an impressed line containing one or more rows of pores; the orifice is somewhat transverse and subovate with the proximal side straightened, the peristome but little elevated, the front sparsely punctated, the avicularia of which there is usually one to each zooecium placed some distance beneath and to one side of the orifice, rather large, subcircular and divided into two nearly equal parts by a thin partition. In old examples the oral part is sunken and the rest of the surface abundantly punctate, while the avicularia have been somewhat reduced in size. Immediately behind the orifice there is always a small (? peristomial) pore. Ovicells large, rather strongly convex, punctate.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.11 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.55 \text{ mm.} \\ lz = 0.25 - 0.35 \text{ mm.} \end{cases}$

We have nothing to add to the original description except to call attention

to the fact that the ascopore is crescent shaped.

This species differ from *Microporclla ciliata* Linnaeus, 1759, in the absence of frontal granules, in the presence of larger tremopores and in its nontransverse aperture which is almost as high as wide. The erect form of the zoarium is rather rare in the genus *Microporclla* and it appears to form a good character for this species. Nevertheless we have observed bilamellar expansions in *Microporella ciliata*.

Occurrence.—Miocene (Choptank formation): Cordova, Maryland (rare). Cotypes.—Cat. No. 68592, U.S.N.M.

MICROPORELLA FISSURIFERA, new species.

Plate 19, figs. 12-13.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a deep furrow, elliptical, little elongated, wide; the frontal is convex and perforated by a large number of tremopores. The apertura is semilunar, transverse; it is surrounded by a thin peristome bearing four large spines. The ascopore is placed

12184-23-Bull, 125---9

on a median prominence of the frontal. The avicularium is short, triangular, without pivot, with its beak turned toward the top. The dietellae are not entirely covered over, and appear between the zooecia in the form of small slits. The ovicell is large, globular, buried in the distal zooecium, covered with a tremocyst.

Measurements.—Apertura $\begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.10 - 0.12 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.50 \text{ mm.} \end{cases}$

Variations.—In this genus, where the species are so difficult of determination, the present species is very well characterized by the dietellae not entirely covered over by the tremocyst. We are ignorant as to the cause of this remarkable peculiarity, which has been observed hitherto only in the Membraniporae. The distal diatellae are visible on the marginal zooecia. The two proximal spines are always larger and more salient than the distal spines. Our micrometric measurements are those of the larger zooecia for the variations are considerable.

Occurrence.—Miocene (Yorktown): Near Macedonia Church, Essex County, (very rare); 1 mile northeast of Suffolk, 3 miles southwest of Petersburg, and other localities in Virginia. Miocene (Duplin marl): 10 miles south of Greenville, North Carolina (rare).

Cotypes.—Cat. Nos. 68593, 68594, U.S.N.M.

MICROPORELLA TESSELLATA Tuomey and Holmes, 1857.

Plate 30, figs. 2, 3.

1857. Cellepora tesselata Tuomey and Holmes, Pliocene fossils of South Carolina, p. 13, pl. 4, fig. 7.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.06 \text{ mm.} \\ la = 0.11 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.75 \text{ mm.} \\ lz = 0.60 \text{ mm.} \end{cases}$

Variations.—There are often six very short and very inconstant spines. The vestibular arch is always visible. The apertura is relatively small for so large a species, which appears to indicate extremely fine tentacles. The frontal is perforated with rather large tremopores. The ascopore is surrounded by a salient peristome. In the vicinity of the ancestrula the avicularium is small and placed at the level of the ascopore. The large marginal zoeccia have their avicularia larger and placed in the wider part of the zooecium well below the ascopore. This character is quite visible on the figure of Tuomey and Holmes and we believe that it is sufficient to recognize this long-forgotten species of these American authors. There are six lateral and one distal dietellae.

Affinities.—This species differs from Microporella eustomata Gabb and Horn, 1862, and from Microporella vibraculifera Hincks, 1883, in its smaller avicularium, more distant from the zooecial axis and of a more irregular position. It differs from Microporella ciliata Linnaeus, 1759, in the large zooecial dimensions and its larger avicularium.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, and Giles Bluff, Peedee River, South Carolina (Tuomey and Holmes).

Pleisotypes.—Cat. No. 68595, U.S.N.M.

MICROPORELLA UMBONATA Hincks, 1882.

Plate 36, figs. 6, 7.

1882. Microporella ciliata forma umbonata Hincks, Report on the Polyzoa of the Queen Charlotte Islands, Annals and Magazine of Natural History, ser. 5, vol. 2, p. 15, pl. 17, fig. 1.

$$\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{bmatrix} ha = 0.08 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{bmatrix} \quad \text{Zooecia} \begin{bmatrix} Lz = 0.52 - 0.60 \text{ mm.} \\ lz = 0.30 - 0.40 \text{ mm.} \end{bmatrix}$$

Variations.—There is one small oral avicularium (rarely two). The frontal is a tremocyst with small pores. The ovicell is costulated. The frontal gibbosity (umbo) is always placed below the ascopore. Our specimens bear no spines.

Affinities.—This species differs from Microporella ciliata Linnaeus, 1759, in the presence of the frontal umbo and in its larger micrometric dimensions. It differs from Microporella fallax Canu, 1904 from the post-Pampean of Argentina, which also bears a frontal gibbosity, in its avicularium which is placed not much below the ascopore but always at the same level with it.

Occurrence.—Pleistocene: Santa Monica (rare), Santa Barbara (very rare), and Dead Mans Island, off San Pedro (very rare), California.

Habitat.—Queen Charlotte Islands. Plesiotype.—Cat. No. 68596, U.S.N.M.

MICROPORELLA CALIFORNICA Hincks, 1883.

Plate 36, figs. 8-10.

1858. Lepralia californica Busk, Zoophytology, Quarterly Journal Microscopical Science, vol. 4, p. 310, pl. 11, fig. 6.

1883. Microporella ciliata forma californica Hincks, Report on the Polyzoa of the Queen Charoltte Islands, Annals aud Magazine Natural History, ser. 5, vol. 2, p. 444, pl. 17, fig. 3.

1908. Microporella californica Robertson, The incrusting chilostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 4, No. 5, p. 281, pl. 18, figs. 32-34.

Variations.—Our specimens are somewhat larger than those living to-day off the coast of California. There are six spines but on our fossils only two, three, or four alone persist. The frontal is a tremocyst with large pores superposed on an olocyst perforated by very small corresponding pores; it is detachable.

The two avicularia are rather constant erect, rather long, very sharp, with a very fragile pivot; their slender beak is easily broken. In some cases one avicularium is smaller than the other. There is a salient frontal gibbosity bearing the ascopore or placed below it. The ascopore is surrounded by a smooth peristone.

Hincks, 1883, thought that his specimens belonged to Busk's species, 1856. Waters, 1908, affirmed that Busk's species is *Microporella coronata* Savigny-Audouin, 1826.

Affinities.—This species much resembles Microporella coronata Savigny-Audouin, 1826, in its costulate ovicell; it differs from it in the presence of six spines (and not four) and in its larger avicularia. It differs from Microporella ciliata Linnaeus, 1759, in the constant presence of two large avicularia, and in its frontal with large tremopores.

Occurrence.—Pleistocene: Santa Monica (Long Wharf Canyon) (rare), Santa Barbara (common), and San Pedro (rare), California.

Habitat.—Queen Charlotte Islands and coast of California.

Plesiotypes.—Cat. Nos. 68597, 68598, U.S.N.M.

MICROPORELLA VIBRACULIFERA Hincks, 1883.

Plate 36, figs. 11, 12.

1883. Microporella ciliata forma vibraculifera Hincks, Report on the Polyzoa of the Queen Charlotte Islands, Annals and Magazine of Natural History, ser. 5, vol. 11, p. 15, pl. 17, fig. 2.

1890. Microporella ciliata, var. vibraculifera Ortmann, Die japanische Bryozoen-Fauua, Archiv für Naturgeschichte, vol. 50, pt. 1, p. 38, pl. 3, fig. 5.

l'ariations.—There are six large hollow spines. The ovicell and the frontal bear large tremopores. The avicularium is large, salient, tuberous, placed laterally and lower than the ascopore; the mandible is placed transversally. The ovicell is costulate. The organ originally called vibraculum by Hincks is in reality an avicularium with very long and setiform mandible. There is often a pivot.

Affinities.—This species differs from Microporella ciliata Linnaeus, 1759, in the presence of six large hollow spines and in larger micrometric measurements. It differs from Microporella fallax Canu, 1904, in its larger avicularium placed very near the ascopore. It differs from Microporella eustomata Gabb and Horn, 1862, in its transverse avicularium.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon) (rare), Dead Mans Island off San Pedro (rare), and Santa Barbara (very rare), California.

Habitat.—Queen Charlotte Islands and Japan (113-405 meters).

Plesiotypes.—Cat. Nos. 68599,68600, U.S.N.M.

MICROPORELLA EUSTOMATA Gabb and Horn, 1862.

Plate 36, fig. 13.

1862. Reptoporina eustomata Gabb and Horn, Monograph of the fossil Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy of Natural Sciences of Philadelphia, ser. 2, vol. 5, p. 144, pl. 20, fig. 26.

$$Measurements. - \Lambda \text{pertura} \begin{vmatrix} ha = 0.06 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{vmatrix}$$
 Zooecia
$$\begin{bmatrix} Lz = 0.60 - 0.80 \text{ mm.} \\ lz = 0.50 \text{ mm.} \end{bmatrix}$$

Affinities.—Gabb and Horn's figure is very characteristic; the mandible of the avicularium is oblique in relation to the zooccial median axis so that the whole avicularium convexity appears bent toward this same axis. This is the exact case in our specimens. However, they bear only six large hollow spines instead of eight shown on the figures of the American authors. The frontal and the ovicell are garnished with large tremopores. The ovicell is preceded by a sort of tubular turret, at the base of which is buried the operculum. The avicularium is curved and always has a pivot.

This species differs from *Microporella vibraculifera* Hincks, 1883, in its oblique and nontransverse avicularium. It differs from all the other species of *Microporella* in its large avicularium.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon) (rare) and Santa Barbara, California (Gabb and Horn).

Plesiotype. -Cat. No. 68601, U.S.N.M.

MICROPORELLA HEERMANNI Gabb and Horn, 1862.

Plate 37, figs. 1, 2.

1862. Reptescharellina heermanni Gabn and Horn, Monograph of fossil Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy Natural Sciences of Philadelphia, ser. 2, vol. 5, p. 147, pl. 20, fig. 30.

Description.—The zoarium incrusts shells. The zooccia are large, elliptical, swollen, separated by a furrow, convex; the frontal is formed by a granular tremocyst perforated by numerous small pores. The apertura is semilunar, transverse; the peristome is thin, somewhat salient, deprived of spines resistant to fossilization.

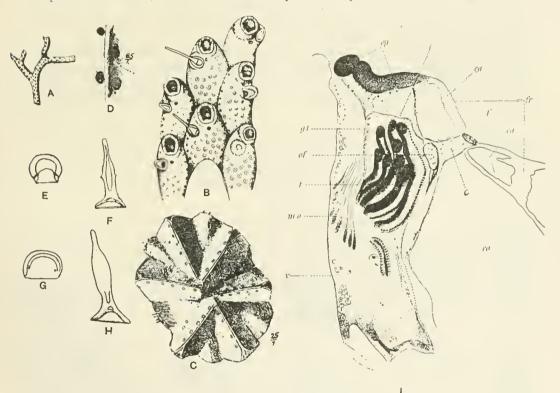


Fig. 21.—Subgenus Diporula Hincks, 1879.

A-F. Diporula verrucosa Peach, 1868. A. Zoarium, natural size. B. Zooecia with avicularia and mandibles, \times 25. (A, B, after Hincks, 1880.) C, D. Growing extremity of a zoarium. The ends of the last formed row of zooecia have a double slope, like the roof of a house with a raised, rounded, slightly overlapping ridge at the top. On each side of this are usually five tubular holes and one or two lower down on the side. At the bottom of these tubes is a membrane which has one, two, or even more minute perforations. D shows the ridge of the roof and rosette pores, \times 85. E. Operculum, \times 85. (C-E, after Waters, 1878.) F. Mandible, \times 85. (After Waters, 1885.)

G-I. Diporula hastigera Busk, 1884. G. Operculum, \times 85. II. Mandible, \times 85. (After Waters, 1889.) I. Section of zooecium showing anatomical structure. (After Jullien, 1903.) ar, avicularium; c, corniculum; ca, calcite; co, compensatrix; f, fenestrule (=ascopore); fr, frontal; gt, tentacular sheath; i, irisoid; mo, opercular muscles; of, inferior orifice of the corniculum; op, operculum; t, tentacles; v, dorsal. The corniculum is not a tube allowing communication of the ascopore with the tentacular sheath as Jullien thought, but it is a simple fold of the compensatrix probably determined by the size of the tentacles.

The ovicell is globular, placed on the distal zooecium, and of the same nature as the frontal. One or two small triangular oblique avicularia are located on each side of the ascopore.

me ascopore.

Measurements.—Apertura $\left\{ \begin{array}{l} ha=0.10 \text{ mm.} \\ la=0.16 \text{ mm.} \end{array} \right.$ Zooecia $\left\{ \begin{array}{l} Lz=0.70\text{--}1.00 \text{ mm.} \\ lz=0.50\text{--}0.60 \text{ mm.} \end{array} \right.$

Affinities.—This large species is well characterized. It differs from Microporella californica Hincks, 1883, in its smaller tremopores, its avicularia twice as short, its granular frontal, and its larger dimensions. The vestibular arch is as large as in Microporella ciliata Linnaeus, 1759; it differs from it in its micrometric measurements, which are twice as large (Lz = 1.00 mm. and not 0.50 mm.). It differs from Microporella gibbera in the regular form of its zooecia and in its small avicularia placed at the level of the ascopore.

Although we have not found this species at Santa Barbara, Gabb and Horn's type locality, we do not hesitate to compare our specimens with Reptescharella heermanni Gabb and Horn. Their figure resembles Microporella ciliata Linnaeus, 1759, as much as it does the present one, but we believe it best to adopt their name instead of suggesting a new one.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon) (rare), and Santa Barbara, California (Gabb and Horn).

Plesiotype.—Cat. No. 68602, U.S.N.M.

MICROPORELLA GIBBERA, new species.

Plate 37, fig. 3.

Description.—The zoarium is unilamellar. The zooecia are distinct, separated by a furrow, very irregular, gibbose; the frontal is convex and perforated by numerous and rather large spines. The apertura is semilunar, transverse; the peristome is thin, a little salient, deprived of spines, resistant to fossilization. ascopore is very small and placed in the immediate vicinity of the apertura. ovicell is large, very globular, irregular, of the same nature as the frontal. two avicularia are very small and placed lower than the ascopore.

Measurements.—Apertura $\begin{bmatrix} ha = 0.12 \text{ mm.} \\ la = 0.14 - 0.16 \text{ mm.} \end{bmatrix}$ Zooecia $\begin{bmatrix} Lz = 0.90 - 1.00 \text{ mm.} \\ lz = 0.60 \text{ mm.} \end{bmatrix}$ Affinities.—The two avicularia are not constant; there is often only a single

one, somewhat a little larger. The form of the zooecia defies description.

This species differs from Microporella heermanni Gabb and Horn, 1862, in which the dimensions are close, in the irregularity of its zooecia and in its small avicularia placed below the ascopore.

Occurrence.—Pleistocene: Rustic Canyon, Santa Monica, California (very rare). Holotype.—Cat. No. 68603, U.S.N.M.

Subgenus DIPORULA Hincks, 1879.

1879. Diporula HINCKS, On the classification of the British Polyzoa, Annals and Magazine Natural History, ser. 5, vol. 3, p. 156.

The operculum is semilunar and closes the ovicell. The frontal is a tremocyst. Avicularia are present. The apertura is horseshoe shaped and is slightly contracted by two lateral projections. The concavity of the ascopore is denticulated.

Genotype. —Diporula (Eschara) verrucosa Peach, 1873.

Range.—Pliocene-Recent.

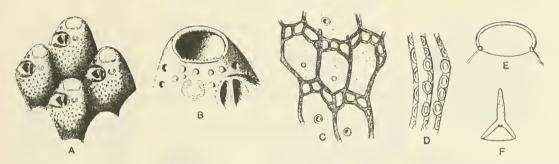


Fig. 22.—Subgenus Ellipsopora, new.

A-F. Ellipsopora (Microporella) flabellaris Busk, 1852. A. Zooecia with avicularia, \times 40. B. The distal end of a zooecium, \times 100. C. Zooecia from the basal surface, \times 40. Besides the basal wall of the dietellae, the small triangular basal surface of the vibracular chamber is seen lowest down to the right on the four zooecia. On some zooecia the basal surface shows a septula and on others an opening corresponding with a septula in an opposite zooecium. D. Radical fibers, \times 140. E. Operculum, \times 140. F. Mandible, \times 55. (A-F, after Levinsen, 1909.)

ELLIPSOPORA, new subgenus.

The aperture is transverse, elliptical. The operculum is elliptical and closes the ovicell. The frontal is a tremocyst. There is a vibraculum on each zooecium. Genotype.—Ellipsopora (Eschara) flabellaris Busk, 1852. Recent.

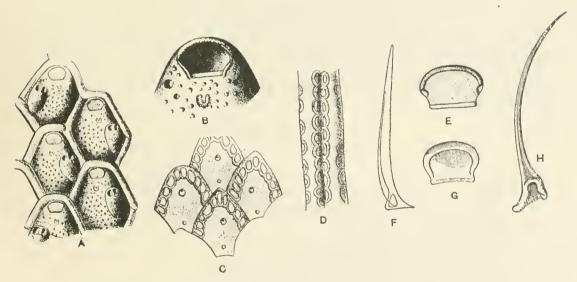


Fig. 23.—Subgenus Flustramorpha Gray, 1848.

A-H. Flustramorpha marginata Krauss, 1837. A. Zooecia with their avicularium, \times 40. B. The distal end of a zooecium, \times 100. C. Four zooecia, from the basal surface, \times 40. In addition to the marginal dietellae, each hasal zooecial surface shows a rosette plate (=septula) and an opening for communication with zooecia in the opposite layer. D. Radical fibers, \times 40. E. Operculum, \times 140. F. Avicularian mandible, \times 55. (A-F, after Levinson, 1909.) G, II. Operculum and mandible. (After Busk, 1885.)

Subgenus Flustramorpha Gray, 1848.

1848. Flustramorpha Gray, List of British Animals in British Museum, pt. 1, Centroniae or Radiated animals.

The operculum closes the ovicell; it is trapezoid. The zoarium is chitinous. "Zoarium erect, radicate, bilaminar, composed of irregular lobes, bordered and loosely connected by chitinous tubes; mouth coarctate; a lateral pouch-like vibracularium." (Busk.)

Genotype.—Flustramorpha (Flustra) marginata Krauss, 1837. Recent.

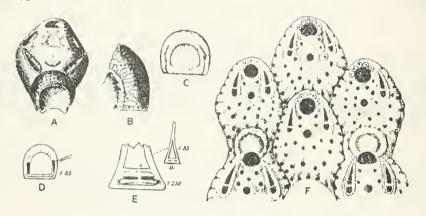


Fig. 24.—Genus Calloporina Neviani, 1895.

A-F. Calloporina decorata Reuss, 1847. A. The zooecium is furnished with three distal dietellae, and the curved belts on the ovicell, at the base of which are seen the fine pores of the endooecium, are canal like cavities between the endooecium and the distal calcified part of the ectooecium. Between these canals, which open through a circle of pores, the two layers of the zooecium have united; × 40. B. A portion of the zooecium, magnified × 75. C. Operculum, × 100. (A-C, after Levinsen, 1909.) D. Operculum, × 85. E. Mandible, × 85. (D, E, after Waters, 1887.) F. Portion of a fossil zoarium. (After Manzoni, 1875.)

Genus CALLOPORINA Neviani, 1895.

1895. Calloporina Neviani, Briozoi fossili della Farnesina e Monte Maria presso Roma Paleontographia Italica, Pisa, vol. 1, p. 107.

The frontal is garnished with lateral areolar pores. Costules and avicularia are present. The ascopore is circular.

Genotype.—Calloporina (Cellepora) decorata Reuss, 1847.

Range.—Helvetian-Recent.

The known species of the genus are:

Calloporina (Cellepora) decorata Reuss, 1847.

Calloporina (Lepralia) diadema MacGillivray, 1868.

Calloporina (Microporella) renipuncta MacGillivray, 1882.

Calloporina (Microporella) scandens MacGillivray, 1884.

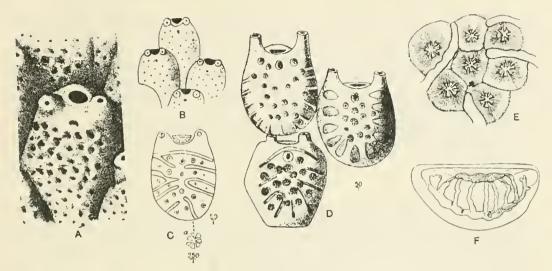


Fig. 25.—Genus Inversiula Jullien, 1888.

A. Inversiula nutrix Jullien, 1888. Zooecium. (After Jullien, 1888.)

B-F. Inversiala inversa Waters, 1889. B. Zooecia, \times 25. C. Decalcified zooecia, \times 50. Distinct tubes occupy the place of the grooves. The interior membrane of the zooecial wall is not perforated by the stellate pores. A stellate pore \times 250 is represented at a. D. Three stages of growth, \times 50. There are deep grooves between the stellate pores. (B-D, after Waters, 1889.) E. Structure of the stellate pores, \times 200. (After Levinsen, 1909.) F. Operculum, \times 250. (After Waters, 1887.)

Genus INVERSIULA Jullien, 1888.

1888. Inversiula Jullien, Mission scientifique du Cap Horn. 1882-83, VI, Zoologie, Bryozoaires, p. 41.

No ovicell. The frontal is a tremocyst with stellate pores. The aperture is elliptical and transverse. The convexity of the ascopore is turned toward the aperture. Avicularia are present. No spines.

Genotype.—Inversiula nutrix Jullien, 1888.

Range.—Miocene-Recent.

The known species of this genus are:

Inversiula nutrix Jullien, 1888.

Inversiula (Microporella) inversa Waters, 1889.

Inversiula (Microporella) airensis Maplestone, 1910.

Inversiula (Microporella) quadricornis Maplestone, 1910.

Group 3. HIPPOPORAE Canu and Bassler, 1917.

Genus HIPPOPORINA Neviani, 1895.

(For description see Bulletin 106, U. S. National Museum, p. 374.)

HIPPOPORINA PUSILLA, new species.

Plate 45, figs. 8, 9.

Description.—The zoarium incrusts sponges and corals. The zooccia are distinct, separated by a furrow, small, clongate, elliptical; the frontal is an absolutely smooth olocyst. The apertura is very small, surrounded by a thick peristome which

is salient tuberous, or garnished with six spines; two very small cardelles placed on the lower third separate the anter from the concave poster. The ovicell is globular, salient; it can not be closed by the operculum.

Measurements.—Apertura $\begin{cases} ha = 0.07. \\ la = 0.05 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.25 \text{ mm.} \end{cases}$

Affinities.—The saliency of its peristome and its small dimensions characterize this species quite well.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (very rare). Holotype.—Cat. No. 68604, U.S.N.M.

HIPPOPORINA GIBBOSA, new species.

Plate 18, fig. 10.

· Descriptions.—The zoarium incrusts shells. The zooccia are distinct, separated by a furrow of little depth, hexagonal, elongate; the frontal is smooth, little convex, garnished with a frontal gibbosity. The apertura is elongate and formed of a large anter separated by two cardelles from a small poster with proximal border almost straight. The ovicell is large globular, widely open.

Measurements.—Apertura $\begin{cases} ha = 0.14 \text{ mm.} \\ la = 0.11 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.45 - 0.50 \text{ mm.} \\ lz = 0.30 - 0.35 \text{ mm.} \end{cases}$

Variations.—No other known species of the genus Hippoporina presents a frontal gibbosity; it is therefore easy to distinguish this one and no error is possible. The micrometric measurements of the zooccia are quite variable and our figure shows a zooccium measuring 0.60 mm. in width. This irregularity is difficult to explain, for it is not occasioned only by calcification; it is real and easy to ascertain on the worn zooccia whose frontal is broken. In most of the other Cheilostomatous bryozoa the irregularity is more apparent than real and examination of the interiors reveals the zooccia to be practically equal.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (rare). Holotype.—Cat. No. 68605, U.S.N.M.

HIPPOPORINA(?) VESTITA, new species.

Plate 18, figs. 7-9.

Description.—The zoarium incrusts oysters. The zooccia are distinct, hexagonal, separated by a furrow of little depth, with little convex frontal, smooth and covered by two or three superposed calcareous pellicles. The apertura is elliptical, elongate. The ovicell is little salient; it bears a very fragile frontal area.

Variation.—This is a strange species, the generic position of which is very uncertain; but we are unable to create a special genus for such incomplete specimens. The latter bear only broken ovicells and we can not obtain an exact idea of their nature. The exterior calcification is very remarkable. The deposit of two or three calcareous pellicles can only be explained by an endocystal covering, immediately adjacent to the ectocyst.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (rare).

Holotype.—Cat. No. 68606, U.S.N.M.

HIPPOPORINA LATA(?) Smitt, 1872.

Plate 1, fig. 11.

1872. Gemellipora lata Smitt, Floridan Bryozoa, pt. 2, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 11, No. 4, p. 36, pl. 7, fig. 157.

1919. Hippoporina lata Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution at Washington, No. 291, p. 93, pl. 1, fig. 14.

This recent species was described from specimens from the Floridan waters and the species itself is confined to this region. A very mediocre fossil specimen of doubtful determination has been found in the strata of Antigua.

Occurrence.—Oligocene (Antigua formation): Rifle Butts, Antigua, Leeward Islands (rare).

Plesiotype.—Cat. No. 68607, U.S.N.M.

Genus HIPPODIPLOSIA Canu, 1916.

(For description see Bulletin 106, U. S. National Museum, p. 393.)

HIPPODIPLOSIA BACCATA Canu and Bassier, 1920.

Plate 3, fig. 1.

1920. Hippodiplosia baccata Canu and Bassler, Monograph of the Early Tertiary Bryozoa of North America, Bulletin 106, U. S. National Museum, p. 397, pl. 87, figs. 5, 6.

This species, which was based on specimens from the Vicksburgian of Mississippi, is represented in Lower Miocene strata at Bowden, Jamaica. In these Miocene specimens the tremocyst is separable and the intensity of the calcification is very active in the ancestrular region.

Occurrence.—Miocene (Bowden marl): Bowden, Jamaica (rare).

Geologic distribution.—Vicksburgian of Mississippi.

Plesiotype.—Cat. No. 68608, U.S.N.M.

HIPPODIPLOSIA BIGIBERRA, new species.

Plate 12, fig. 10.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a furrow, clongated, subrectangular: the frontal is somewhat convex, perforated by a dozen of small tremopores and ornamented with two gibbosites symmetrically placed in the vicinity of the apertura, which they hide and partially deform. The apertura is elliptical, elongated; two small, deep cardelles separate a large anter from a small poster.

Measurements.—Apertura $\begin{bmatrix} ha = 0.17 - 0.19 \text{ mm.} \\ la = 0.15 - 0.17 \text{ mm.} \end{bmatrix}$ Zooccia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.35 - 0.45 \text{ mm.} \end{cases}$

Structure.—The aspect of the apertura is very deceiving in a photograph. It appears to resemble that of Gemellipora, but this is an optical illusion arising from the projection on the same plane of the different zooecial elevations. In reality the apertura is elliptical, but its proximal portion is deformed by two frontal gibbosites which appear thus separated by a false rimule. By inclining specimens the true form of the apertura is visible.

Occurrence.—Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia (rare).

Holotype.—Cat. No. 68609, U.S.N.M.

Genus HIPPOMENELLA Cann and Bassler, 1917.

(For description see Bulletin 106, U. S. National Museum, p. 379.)

HIPPOMENELLA INFRATELUM Canu and Bassler, 1919.

Plate 6, fig. 2.

1919. Hippomenella infratelum CANU and BASSLER, Geology and Paleontology of the West Indies Bryozoa, Publications of the Carnegie Institution of Washington, No. 291, p. 92, pl. 6, fig. 2.

Description.—The zoarium is a narrow Eschara borne on an expanded base. The zooecia are elongate, distinct, elliptical; the frontal is convex, surrounded by a line of small areolar pores and formed of a pleurocyst very finely granulated. The apertura is elliptical, elongate, with two very small cardelles. The avicularium is salient, elliptical with pivot, placed on the line of pores in the lower part of the zooecium.

Measurements.—Apertura
$$| ha = 0.16 - 0.20 \text{ mm.} | Lz = 0.60 - 0.70 \text{ mm.} | la = 0.14 \text{ mm.} | Lz = 0.40 - 0.50 \text{ mm.} | L$$

Affinities.—The only specimen found has been figured. The species is absolutely characterized by its elliptical avicularium placed inferiorly, a character which does not exist in the other species of the genus. We have observed a rather rare case of regeneration of a zooccium by an avicularium (zr).

Occurrence.—Lower Miocene (Bowden marl): Cercado de Mao, Santo Domingo (very rare).

Holotype.—Cat. No. 68610, U.S.N.M.

Genus HIPPOPORELLA Canu and Bassler, 1920.

(For description see Bulletin 106, U. S. National Museum, p. 377.)

HIPPOPORELLA SPINOSA, new species.

Plate 19, fig. 10.

Description.—The zoarium incrusts other bryozoa. The zooecia are distinct, separated by a deep furrow, somewhat elongate, elliptical; the frontal is smooth, very convex, garnished laterally with small areolar widely spaced pores. The apertura is elongate, the proximal border is almost straight; the two small cardelles are placed very low; the peristome is little salient and bears six spines. There is a vestibular arch. The ovicell is large, globular, salient, never closed by the operculum.

Measurements.—Apertura
$$\begin{cases} ha = 0.12 \text{ mm.} \\ la = 0.10 - 0.12 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.35 \text{ mm.} \end{cases}$

Variations.—The zooccia are rather variable in aspect and measurements; however, the number of spines, the vestibular arch, and the place of the cardelles are very constant characters which permit no confusion with the species which we have described from the American Eccene.

Occurrence.—Miocene (Yorktown): Yorktown, Virginia (rare). Holotype.—Cat. No. 68611, U.S.N.M.

HIPPOPORELLA(?) PAPULIFERA, new species.

Plate 19, fig. 14.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a deep furrow, elongated, elliptical; the frontal is very convex, perforated by a double row of areolar pores, ornamented with little salient costules and covered by a more or less granular pleurocyst. The apertura is oblique, deep, located at the base of a peristomie, the peristome of which is spinous, very thin and little salient; the two cardelles are quite low. A hollow mucro in the form of a pimple hides the proximal part of the apertura.

Affinities.—We have not discovered the ovicell of this species. The frontal granulations are rare or numerous according to the place of the zooecia. On account of the obliquity of the aperture the two cardelles are visible only when the specimen is inclined to an angle of 45°.

This species differs from Lepralia montifera Ulrich and Bassler, 1904, in its nontransverse aperture and from Hippoporella costulata in the absence of small adventitious avicularia.

Occurrence.—Miocene (Choctowhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (very rare).

Holotype.—Cat. No. 68612, U.S.N.M.

HIPPOPORELLA COSTULATA, new species.

Plate 45, fig. 14.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a deep furrow, hexagonal, widened; the frontal is convex and formed of an olocyst surmounted by a pleurocyst garnished laterally with arcolar pores and with costules converging toward the oral mucro. The apertura bears two very small cardelles placed in the lower third; the proximal border is somewhat concave and denticulated. The ovicell is globular, smooth, little salient, much embedded in the distal zoccium; it is never closed by the operculum. On the line of pores there are small irregularly placed avicularia.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{bmatrix} ha = 0.12 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{bmatrix} \quad \text{Zooecia} \begin{bmatrix} Lz = 0.45 - 0.50 \text{ mm.} \\ lz = 0.35 - 0.45 \text{ mm.} \end{bmatrix}$

Variations.—The line of arcolar pores is often doubled by another more interior one. The passage of the eggs, as in all the species of this genus, is assured by the embedding of the ovicell in the distal zooecium and in the presence of the oral mucro formed by the coalescence of the interarcolar costules. In the vicinity of the ancestrula the zooecia have some large frontal pores. The species has neither peristome nor spines.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (very rare). Holotype.—Cat. No. 68613, U.S.N.M.

Genus LEPRALIA Johnston, 1847.

The name Lepralia is retained for those species of the Hippoporae which show no ovicell and which cannot therefore be more definitely placed.

LEPRALIA MONTIFERA Ulrich and Bassler, 1904.

Plate 18, fig. 11.

1904. Lepralia montifera Ulrich and Bassler. Maryland Geological Survey, Miocene, p. 424, pl. 116, fig. 5.

The original description is as follows:

Zoarium parasitic, in one or more layers. Zooecia not regularly arranged, subovate, averaging 0.5 mm. or a trifle more in length and about 0.35 mm. in width. Orifice oblique, depressed in front, transversely subovate, broadly sinuate below; peristone scarcely thickened. Central portion of surface very high, the slopes traversed by rows of large punctures in radially disposed furrows. Ovicells not observed; nor avicularia, unless certain elongate-acuminate, curved depressions, with a pore at the broader lower extermity, that sometimes may be observed close to the rim of the orifice, are of that nature. This rather highly ornamented form reminds in certain respects of *Cribrilina*, but on the whole it agrees better with *Lepralia*. The strikingly monticular elevation and strongly puncto-radiate marking of the surface of the zooecia will, we believe, serve very well in distinguishing the species.

We are unable to classify this species generically on account of the absence of ovicell on the type and only specimen.

The interarcolar costules are larger than in *Hippoporella costulata*, but the present species differs in the absence of a hollow infraoral mucro and of the small adventitious avicularia.

Occurrence.—Miocene (St Mary's formation): St. Marys River, Maryland (very rare).

Holotype.—Cat. No. 68614, U.S.N.M.

LEPRALIA CRIBROSA? Maplestone, 1900.

Plate 36, fig. 1.

1900. Lepralia cribrosa Maplestone, Further descriptions of the Tertiary Polyzoa of Victoria, pt. 6, Proceedings Royal Society, Victoria, vol 13, pt. 2, p. 210, pl. 35, fig. 16.

We are not certain of our determination, although the specimen studied is a superb one. If the magnification indicated by Maplestone be exact our species would be somewhat smaller. The absence of an ovicell does not allow the species to be classified generically. Two very small cardelles separate a large anter from a smaller poster.

Occurrence.—Pleistocene: Santa Monica, (Long wharf Canyon), California (very rare).

Geological distribution.—Miocene of Australia (Maplestone).

Plesiotype.—Cat. No. 68615, U.S.N.M.

Group 4. PERISTOMELLAE Canu and Bassler, 1917.

Genus TRYPEMATELLA Canu and Bassler, 1920.

1920. Trypcmatella Canu and Bassler, Monograph North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 417.

The ovicell is hyperstomial and closed by the operculum for the passage of the eggs. The apertura is semilunar with proximal border a little concave. The frontal bears some lateral areolar pores (pleurocyst on olocyst). Two large lateral avicularia are placed below the apertura.

Genotype.—Trypematella papulifera, new species. Pleistocene.

TRYPEMATELLA PAPULIFERA, new species,

Plate 35, figs. 12-14.

Description.—The zoarium is free and cylindrical; it incrusts fine algae. The zooecia are little distinct, short, wide; the frontal is little convex and perforated laterally by large arcolar pores. The apertura is semiclliptical; the peristome is salient and very thin with four distal spines; the ovicell is very large, placed on the distal zooecium, globular, salient, closed by the operculum for the passage of the eggs; it is costulate and granulated and bears a smooth prominence at its summit. On each side of the apertura there is a small round avicularium without pivot. Laterally, on the line of the arcolar pores, on each side of the zoecium, there is a large triangular avicularium, with pivot, transverse or turned toward the top.

Measurements.—Apertura $\begin{vmatrix} ha = 0.09 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{vmatrix}$ Zooecia $\begin{vmatrix} Lz = 0.40 - 0.45 \text{ mm.} \\ lz = 0.30 - 0.45 \text{ mm.} \end{vmatrix}$ (With the avicularia.)

Variations.—This fine species is very irregular in aspect on account of the irregularities of calcification. This is very active and gives a great thickness to the zooecial walls. The size and number of the avicularia, the organs of oxygenation, seem to indicate that the species lived in very calm waters.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (rare). Cotypes.—Cat. No. 68616, U.S.N.M.

Group 5. DIVERS GENERA.

Genus CYCLOCOLPOSA Canu and Bassler, 1920.

1920. Cyclocolposa Canu and Bassler, Monograph North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 431.

The apertura is suborbicular or elliptical without cardelles. The frontal is an olocyst, perforated by a double row of areolar pores, and covered by a granular, detachable pleurocyst. The ovicell is hyperstomial, never closed by the operculum, embedded in the distal zooecium.

Genotype.—Cyclocolposa perforata, new species.

Range.—Miocene-Pliocene.

In spite of appearances this genus is very different from Cyclicopora Hincks, 1884. The frontal pores are really areolar pores and not tremopores, for they are separated by short costules; the granulations reveal also the detachable pleurocyst, moreover, and are often visible on the altered zooecia. In Cyclicopora the operculum always closes the ovicell to assure the passage of the eggs; here this function is assured by the embedding of the ovicell in the distal zooccium, and in the great thickness of the frontal; the orifice is thus arranged in the locella in front of the tentacular sheath.

CYCLOCOLPOSA PERFORATA, new species.

Plate 30, figs. 6-14.

Description.—The zoarium incrusts oysters over large surfaces. The zooecia are distinct, separated by a furrow, elliptical or hexagonal, short, wide; the frontal is convex, surrounded by a double row of areolar pores, and formed of an olocyst supporting a granular and detachable pleurocyst. The apertura is oblique, subor-

bicular, deeply embedded. The ovicell is globular, smooth, little salient, deeply embedded in the distal zooccium. The walls are very thick and perforated by a large number of dietellae. The ancestrula is small and reduced to only the apertura; it emits five zooccia.

Measurements.—Apertura $\begin{cases} ha = 0.17 \text{ mm.} \\ la = 0.17 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.50 - 0.55 \text{ mm.} \\ lz = 0.50 \text{ mm.} \end{cases}$

Variations.—The pleurocyst is detachable by alteration during fossilization (fig. 10). The ancestrula, normal in figure 8, is often covered by the pleurocyst of the adjacent zooecia (fig. 9). Sometimes there are nonoriented zooecia. The calcified zooecia are provided with tremopores. The structure of the olocyst is that of Cyclicopora. The very thick walls with numerous dietellae characterize this species (fig. 14); their number is quite variable from 12 to 20.

Mueronella laqueata Norman, 1864, presents the same dietellae; it differs from

it in its perforated frontal and in the absence of lyrule.

Occurrence. Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (very common).

Cotypes.—Cat. No. 68617, U.S.N.M.

CYCLOCOLPOSA TENUIPARIETIS, new species.

Plate 21, figs. 1-4.

Description.—The zoarium incrusts pelecypod shalls. The zooecia are distinct, clongated, elliptical, separated by a deep furrow; the frontal is convex, surrounded by a double row of large, scattered, areolar pores, separated by pleurocystal granules. The apertura is elongate, oblique, elliptical, without cardelles. The ovicell is small, hardly salient, much embedded in the distal zooecium, with frontal smooth and fragile. Rarely a round oral avicularium is present.

 $\label{eq:measurements} \textit{Measurements.} - \Delta \text{pertura} \begin{cases} \hbar a = 0.16 \text{ mm.} \\ la = 0.14 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.54 - 0.58 \text{ mm.} \\ lz = 0.46 - 0.50 \text{ mm.} \end{cases}$

Affinities.—The presence of the ovicell absolutely disfigures this species; isolated portions of the zoarium containing only ovicelled zooecia can not be determined with certainty. The fragility of the superior wall is extreme; it does not resist fossilization; it results in a fantastic aspect difficult to comprehend. Exteriorly the pleurocyst is much reduced, and the abundance of arcolar pores seems to indicate a tremocyst; the study of the interior does not permit this supposition.

This species differs from Cyclocolposa perforata in its elongate zooccia and in its

very thin zooecial walls.

Occurrence.—Miocene (Duplin marl): Harvey Mills, Leon County, Florida (rare). Cotypes.—Cat. No. 68618, U.S.N.M.

CYCLOCOLPOSA(?) SPINIFERA, new species.

Plate 20, figs. 12, 13.

Description.—The zoarium incrusts oysters. The zooecia are distinct, separated by a deep furrow, somewhat elongated, much swollen; the frontal is very convex, garnished laterally with a double row of arcolar pores and formed of an olocyst supporting a detachable pleurocyst. The apertura is orbicular or somewhat elongated and is surrounded by six large hollow spines. The ovicell is hyper-

stomial, very globular, smooth, costulated on its borders; it is closed by the operculum. A very small median avicularium opens in the apertura.

Variations.—The exterior aspect of this species is rather variable. Figure 12 represents the young zoarium which has not had time to attain its complete development. The apertura appears as elliptical and elongated; the frontal is still not completely covered by the pleurocyst and the deposit of calcite is clearly visible around each areolar pore. In figure 13 the calcification is complete; the frontal is very thick and the form of the apertura here becomes undefinable. The small median avicularium appears to be excavated in the thickness of the pleurocyst.

Occurrence.—Miocene (Yorktown formation): 1 mile west of Fort Nonsense, Gloucester County (rare), Beulahland, King and Queen County, and other localities in Virginia (rare).

Cotypes.—Cat. No. 68619, U.S.N.M.

Genus CYCLOPERIELLA Canu and Bassler, 1920.

1920. Cycloperiella Canu and Bassler, Monograph North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 43.

The ovicell is hyperstomial, globular, not embedded in the distal zoecium, and entirely covers the apertura. The apertura is formed of a semilunar anter and of a very concave poster. The frontal is formed of a very thin olocyst supporting a tremocyst with large widened pores.

Genotype.—Cycloperiella rubra, new species.

Range.—Miocene-Pliocene.

This genus offers the same frontal and apertural characters as *Cyclicopora* Hineks, 1884. It differs from it in its ovicell never closed by the operculum. The passage of the eggs is assured by the form of the ovicell itself, which entirely covers the apertura.

CYCLOPERIELLA RUBRA, new species.

Plate 21, figs. 5-9.

Description.—The zoarium incrusts shells and frequently presents two superposed lamellae. The zooccia are elongate, a little elliptical, separated by a deep furrow; the frontal is thick, convex, and covered with large, widened tremopores. The apertura is semilunar, with a concave proximal border, and surrounded by a sort of thick and little salient peristome. The ovicell is globular, salient, covered with tremopores; it entirely covers the apertura. Its exterior orifice is irregular.

Measurements.—Apertura
$$\begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.15 \text{ mm.} \end{cases}$$
 Zooccia $\begin{cases} Lz = 0.55 - 0.60 \text{ mm.} \\ lz = 0.35 \text{ mm.} \end{cases}$

Affinities.—The zoarium often covers large surfaces; it was strongly pigmented, all the fossils are purple, indicating that their primitive tint was red. In the interior the very thin and pellicular olocyst scarcely resists fossilization, and the tremopores are clearly visible. Two very small lateral condyles serve as axis to the operculum. The orifice of the ovicell is irregular, oblique to the zooceial surface. It is difficult to comprehend how the polypide could extend its tentacles.

12184-23-Bull, 125---10

It is therefore probable that after the passage of the eggs the polypide disappears by histolysis. The zooecia with calcified orifice are not rare. Sometimes a small gibbosity occurs on the frontal.

Occurrence.—Miocene (Duplin marl): Wilmington, 8 miles east of Snow Hill, 10 miles south of Greenville, and Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina (common). Miocene (Yorktown formation): Suffolk, 3 miles southwest of Petersburg, and other localities in Virginia (common). Miocene (Bowden marl): Bowden, Jamaica (rare). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (common).

Cotypes.—Cat. Nos. 68620-68625, U.S.N.M.

Genus CYCLICOPORA Hincks, 1884.

(For description, see Bulletin 106, U.S. National Museum, p. 424.)

CYCLICOPORA MULTILAMELLOSA, new species.

Plate 46, figs. 3-6.

Description.—The zoarium is free, irregularly orbicular, striated concentrically on the inferior face and formed of many superposed lamellae. The zooecia are heaped upon each other, little raised, nonoriented, distinct, elongated, hexagonal or rectangular; the frontal is little convex, tuberous, and formed of a tremocyst with scattered pores, resting on an olocyst. The apertura is elliptical, transverse, with a concave proximal border; it is placed at one of the zooecial angles. In the immediate vicinity of the apertura there is a small orbicular avicularium sometimes transformed into a long transverse avicularium. The ovicell is globular, salient, very large, partially embedded in the distal zooecium and always closed by the operculum; it is covered by the tremocyst.

by the operculum; it is covered by the tremocyst.

Measurements.—Apertura
$$\begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.20 \text{ mm.} \end{cases}$$
Zooecia $\begin{cases} Lz = 0.80 \text{ mm.} \\ lz = 0.60 - 0.80 \text{ mm.} \end{cases}$

Variations.—The zooccial dimensions are quite variable; the zooccia are elongated or transverse. The characteristic of this species is the excentric position of the apertura; this and the avicularium occupy the distal portion of the zooccium. In the interior the aperture occupies its usual median position; it is therefore the irregularities of calcification which causes its exterior aspect.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare). Cotypes.—Cat. No. 68626, U.S.N.M.

CYCLICOPORA? MANSFIELDI, new species.

Plate 19, fig. 11.

Description.—The zoarium incrusts Cellepore bryozoa. The zooecia are distinct, separated by a furrow of little depth, elongated, elliptical; the frontal is little convex and covered with large, widely spaced tremopores. The apertura is suborbicular, the proximal border being simply concave; neither cardelles nor peristome are present.

Measurements.—Apertura
$$\begin{cases} ha = 0.12 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.35 \text{ mm.} \\ lz = 0.25 - 0.30 \text{ mm.} \end{cases}$

As our single specimen is without an ovicell, the generic determination must remain doubtful. This species is named in honor of Mr. Wendell C. Mansfield, of the United States Geological Survey, who has collected many interesting specimens of bryozoa for our study.

Occurrence.—Miocene (Yorktown formation): 1½ miles southwest Reed's

Ferry, Virginia (very rare).

Holotype.—Cat. No. 68627, U.S.N.M.

CYCLICOPORA(?) GIGANTEA, new species.

Plate 37, fig. 5.

Description.—The zoarium incrusts pebbles. The zooecia are distinct, gigantic, ogival or hexagonal; the frontal is convex and formed of a tremocyst with small scattered pores surmounting an olocyst perforated by very minute pores corresponding to the tremopores. The apertura is elongate, elliptical, without lyrule or cardelles, with a concave proximal border; it is surrounded by a salient peristome, more or less thick and widened. The ovicell is hyperstomial, not embedded in the distal zooecium.

Measurements.—Apertura $\begin{cases} ha = 0.30 - 0.35 \text{ mm.} \\ la = 0.30 - 0.35 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz = 1.50 - 1.60 \text{ mm.} \\ lz = 0.90 - 1.40 \text{ mm.} \end{cases}$

Variations.—The micrometric measurements are variable. The tremocyst is detachable and finely granular between the pores. The ovicells of our specimens are broken and we are therefore unable to classify the species without doubt.

Occurrence.—Pleistocene: Santa Monica (Tremochal Canyon), California (rare).

Holotype.—Cat. No. 68628, U.S.N.M.

Genus AIMULOSIA Jullien, 1888.

(For description, see Bulletin 106, U. S. National Museum, p. 428.)

AIMULOSIA ACULEATA, new species.

Plate 21, figs. 10-14.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a furrow, ovoid, wide; the frontal is convex, smooth, perforated laterally by small, scattered, areolar pores; it bears on its median line a small avicularium very salient, oblique, orbicular, without pivot. The apertura is semilunar, a little elongate, with a somewhat concave proximal border; the peristome is thick, little salient, and garnished by two distal spines. The ovicell is globular, smooth, quite salient, transverse, very widely open in front of the median avicularium. Frequently there is a small triangular avicularium at the side of the apertura.

 $\begin{array}{lll} \textit{Measurements.} & -\text{Apertura} \\ la = 0.10 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{array} \end{array} \quad \text{Zooeeia} \\ \begin{array}{ll} Lz = 0.50 \text{ mm.} \\ lz = 0.45 \text{ mm.} \end{array}$

Variations.—In all the species of this genus there is a very great micrometric difference between the ancestrular zooccia and the marginal ones. Our measurements are the largest that we have observed. The oral avicularium replaces an areolar pore. This is the rule in all the cheilostomatous Bryozoa. This organ is nourished in fact from the interior of the zooccia and must necessarily be in relationship with the mesenchymatous tissue. This avicularium is inconstant in its presence; sometimes it is lacking; sometimes there are two. The irregularity

of the avicularia is one of the most tantalizing problems of bryozoology. It will remain unsolved until we know the exact physiological functions of these organs.

The apertura has no fixed measurements; it may be clongate or transverse, large or small. It appears to be in rapport with the size of the zooecium itself. The ancestrula is very small; it is reduced to its apertura; it engenders four normal zooecia and a zooeciule bearing only a salient median avicularium. The presence of the median avicularium is a vital necessity for the zooecium and the first bud of the ancestrula is really this avicularium. The specimens from the Pliocene are still more irregular than those of the Miocene.

Affinities.—This species differs from Aimulosia brevis in its large micrometric dimensions and in the presence of arcolar pores. It differs from Aimulosia radiata in its greater zooccial width, in the absence of interarcolar costules, and in the

much smaller areolar pores.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (rare); Harvey's Mills, Leon County, Florida (rare); and Muldrow's Mills, 5 miles south of Maysville (rare), South Carolina. Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare).

Cotypes.—Cat. Nos. 68629, 68630, U.S.N.M.

AlMULOSIA BREVIS, new species.

Plate 3, figs. 5-7.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a furrow, ovoid, very short; the frontal is convex, smooth, generally without arcolar pores, terminated on the median line by a small, round, very salient avicularium. The apertura is semielliptical; the peristome is little salient, wide, and bears two hollow spines. The ovicell is globular, salient, smooth, transverse with a very large orifice. Frequently there is a triangular avicularium at the side of the peristome.

Measurements.—Apertura $\begin{cases} ha = 0.08 \text{ mm.} \\ la = 0.08 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.30 \text{ mm.} \\ lz = 0.20 \text{ mm.} \end{cases}$

Affinities.—The arcolar pores are very small; they are easily closed by fossilization and the zooccia appear deprived of them.

This species differs from Aimulosia aculcata in its smaller micrometric

measurements and in the apparent absence of areolar pores.

Occurrence.—Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida (rare). Lower Miocene (Bowden marl): Bowden, Jamaica (common). Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (rare).

Cotypes.—Cat. Nos. 68631-68633, U.S.N.M.

AIMULOSIA RADIATA, new species.

Plate 32, fig. 5.

Description. The zoarium incrusts shells. The zooccia are distinct, separated by a furrow, elongate, claviform; the frontal is convex, garnished laterally with large areolar pores and with interareolar costules arranged in radiating rows, and formed of a pleurocyst, slightly granular, surmounting a smooth olocyst; the median avicularium is very salient, oblique, and surmounts a mucro. The apertura is semilunar, located at the base of a short peristomie. The peristome is thick and bears two hollow spines. The ovicell is globular, very salient, smooth, very widely open before the median avicularium.

 $\begin{array}{lll} \textit{Measurements.} & -\text{Apertura} | ha = 0.08 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{array} \\ & \text{Zooecia} \\ \begin{array}{lll} Lz = 0.40 \text{ mm.} \\ lz = 0.20 - 0.30 \text{ mm.} \end{array}$

Affinities.—This species is very easy to recognize by its interareolar radiating costules; however, this character is much less apparent on the zooccial margins where the pleurocystal calcification is incomplete.

Occurrence.—Pliocene (Caloosahatchee marl): Shell Creek, De Soto County,

Florida (very rare).

Holotype.—Cat. No. 68634, U.S.N.M.

Family EURYSTOMELLIDAE Levinsen, 1909.

Levinsen's original description is as follows:

No spines. The thick-walled and strongly calcified zooecia lack a covering membrane and have either no pores at all or 2-5 extremely large feuestrae. The broad aperture, which has a concave proximal rim, is provided with a more or less strongly chitinized operculum surrounded by a continuous marginal thickening. No avicularia. The ooecium is enclosed in a kenozooecium, the frontal wall of which is provided with a very large uncalcified part. Pore-chambers [dietellae] or groups of uniporous rosette-plates [septulae].

The two known species classed in this family are Lepralia foraminigera Hincks, 1883, and Lepralia bilabiata Hincks, 1884. Levinsen believed that the second species is probably the type of a special genus while the first is the type of the genus Eurystomella.

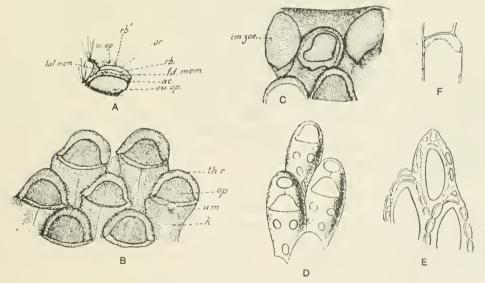


Fig. 26.—Genus Eurystomella Levinsen, 1909.

A-C. Eurystomella bilabiota Ilincks, 1884. A. Sketch showing the operculum dissected out; rb, outer rib; rb', inner rib; in. op., inner operculum; or, orifice; lat. mem., lateral membrane; fd. mem., folded membrane; oc, accessory lip; ou. op., outer operculum. B. A few adult zooecia, \times 30; th. r, thickened rim; op, operculum; um, umbo; k, keel. C. A few zooecia at the growing edge of the colony showing the membranous aperture of the immature zooecia (im. zoe.) (A-C, after Robertson, 1908.)

D-F. Eurystomella foraminique Ilineks, 1883. D. Two gonozooccia and an ordinary zooccium, \times 40. E. Three zooccia, from the basal surface, which has a large uncalcified central part, \times 40. F. A sagittal section through a zooccium with ovicell, \times 40. The orifice of the ovicell is closed by the ectocyst. (D-F, after Levinsen, 1909.)

Genus EURYSTOMELLA Levinsen, 1909.

1909. Eurystomella Levinsen, Morphological and systematic studies on the cheilostomatous Bryozoa, p. 314.

Characters same as for the family. The operculum closes the ovicell.

EURYSTOMELLA BILABIATA Hincks, 1884.

Plate 37, fig. 6.

1884. Lepralia bilabiata Hineks, Polyzoa Queen Charlotte Islands, Annals and Magazine Natural History, ser. 5, vol. 13, p. 49, pl. 3, fig. 1.

1908. Lepralia bilabiata ROBERTSON, The incrusting Cheilostomatous Bryozoa of the West coast of North America, University of California Publications, Zoology, vol. 4, no. 5, p. 298, pl. 21, figs. 61-64.

Our fossil specimens of this species are mediocre and do not permit us to add any useful contribution to the knowledge of this remarkable species. We therefore have no modifications to add to Levinsen's description.

Occurrence.—Pleistocene: Dead Man's Island, off San Pedro, California (rare).

Habitat.—Pacific: California and Queen Charlotte Islands.

Plesiotype.—Cat. No. 68635, U.S.N.M.

Family STOMACHETOSELLIDAE Canu and Bassler, 1917.

Genus LEIOSELLA Cann and Bassler, 1917.

(For description see Bulletin 106, U. S. National Museum, p. 448.)

LEIOSELLA EDAX, new species.

Plate 22, figs. 1-6.

Description.—The zoarium is free, unilamellar, often tubular; it was attached to the delicate radicells of algae. The zooecia are indistinct. The peristomice is very large, elongate, provided with a large rimule spiramen; the peristomic is deep; the apertura (in the interior) is semilunar with a concave proximal border. The ovicell is placed in the peristomic; it is widely open above the operculum; it bears a circular, very fragile frontal area. Around the peristomice there are three or four small, round, salient avicularia, with two denticles for pivot; they are sometimes transformed into large avicularia with spatulate mandibles. Another avicularium opens into the peristomic.

Measurements.—Apertura $\begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.125 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.30 - 0.35 \text{ mm.} \end{cases}$ (interior) $\begin{cases} lz = 0.30 - 0.35 \text{ mm.} \end{cases}$

Affinities.—The aspect of this species is quite variable and depends solely on the number and development of the frontal avicularia. It appears very greedy for oxygen, a fact which is supported by the great thickness of its frontal wall and the activity of the calcareous secretion. Sometimes two small avicularia are replaced by a much larger one with large spatulate mandible.

The peristomic is the real seat of life of this species. An avicularum (visible from the interior) opens into the lateral portion. The distal portion is reserved for the ovicell. Finally, the pseudo-rimule spiramen in the proximal portion is the orifice of the hydrostatic system. It is probable that the female polypide disappears by histolysis at the moment of the development of the embryos and the escape of the larva.

Occurrence.—Lower Miocene (Chipola marl): Chipola River, Calhoun County, Florida (rare).

Cotypes.—Cat. No. 68636, U.S.N.M.

Family SMITTINIDAE Levinsen, 1909.

Genus SMITTINA Norman, 1903.

(For description see Bulletin 106, U. S. National Museum, p. 456.)

SMITTINA TRISPINOSA Johnston, 1838.

Plate 22, figs. 7-14.

1838. Lepralia trispinosa Johnston, History of British Zoophytes, p. 324, pl. 57, fig. 7.

1889. Smittia trispinosa Jelly, A synonymic catalogue of marine Bryozoa, p. 252 (general bibliography).

1894. Smittia trispinosa Levinsen, Zoologica Danica (Danske Dyr), Heft 9, p. 70, pl. 6, figs. 10, 11. 1896. Smittia trispinosa Neviani, Briozoi Postpliocenici di Spilinga (Calabria), Atti Accademia

Gioenia di Scienze Naturali in Catania, ser. 4, vol. 9, p. 41, fig. 22.

1899. Smittia trispinosa Waters, Bryozoa from Maderia, Journal Royal Microscopical Society, p. 16.
1900. Smittia trispinosa Robertson, Papers from the Harriman Alaskan Expedition, VI, The Bryozoa, Proceedings Washington Aacademy Sciences, vol. 2, p. 327.

1902. Smittia trispinosa Calvet, Bryozoaires marins de la region de Cette, Travaux de l'Institut de Zoologie de Montpellier, Mem. 11, p. 56.

1902. Smittia trispinosa Calvet, Bryozoaires marins des cotes de Corse, Travaux de l'Institut de Zoologie de Montpellier, Mem. 12, p. 30.

1907. Smittina trispinosa Nordgaard, Bryozoen von dem norwegischen Fischereidampfer "Michael Sars," Bergens Museums Aarbog, no. 2, p. 15.

1908. Smittia trispinosa Robertson, The incrusting cheilostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 4, pl. 22, figs. 68-70.

1910. Smittia trispinosa Osburn, Bryozoa of the Woods Hole region, Bulletin Bureau of Fisheries, vol. 30, p. 246, pl. 27, fig. 65 (American bibliography).

1912. Smittina trispinosa Nordgaard, Revision av Universitets museets samling av norske Bryozoer, Kgl. norske Videnskabers Selskabs Skriften, no. 3, p. 22.

1914. Smittina trispinosa Osburn, The Bryozoa of the Tortugas Islands, Florida, Publications of the Carnegie Institution of Washington, no. 182, p. 208.

Variations.—This species is extremely variable and many varieties are known. The varieties arborea Levinsen and lamellosa Smitt appear to be peculiar to the boreal regions. The varieties munita Hincks, bimucronata Hincks, nitida Verril, japonica Ortmann, etc., have no particular geographic extension and they generally accompany the type. The varieties, of which we have not cited the bibliography, are established either on the zoarial aspect or on the form of the avicularian mandibles; that is to say, on very secondary characters. Calvet, 1902, has shown that these characters themselves are rather characters of variations.

The detailed study of the variations would be desirable and would permit perhaps the discovery of the cause of them, which would be very important from the biologic point of view. It is probable that the richness of the waters in planktonic elements, the swiftness of the currents, the nature of the substratum, the agitation of the waters, and the nature of the bottom are the causes of the variations of which it is important to establish exact limits.

The geographic distribution of this species is universal; it has been discovered in all the explored seas. According to the known rules, the geological distribution must be correlative. But it is not so; only Waters and Neviani have mentioned fossil specimens.

The constant characteristic of this species is its very regular mode of gemmation (fig. 10) operating alternately between the walls which are absolutely recti-

linear and deprived of septules. The pseudo-rimule of the peristomice has no avicularium as in almost all other species of the genus. This avicularium is generally placed on the line of the areolar pores and sometimes on the frontal; it is unequally developed and almost always triangular on our fossils. The lyrule is very small; it is more or less visible according to the development of the lateral lips of the peristomice.

Measurements.—Apertura $\begin{cases} ha = 0.08-0.10 \text{ mm.} \\ la = 0.08-0.10 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz = 0.36-0.50 \text{ mm.} \\ lz = 0.20-0.30 \text{ mm.} \end{cases}$

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (common). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare). Pliocene (Caloosahatchee marl): Shell Creek, De Soto County (common), and Monroe County, Florida (common). Pleistocene: Vero (rare), Simmons Bluff, Yonges Island, Charleston County, South Carolina (common), and Daytona, Florida (rare.)

Geological distribution.—Miocene of Australia (Waters); Quaternary of Sicily (Neviani).

Habitat.—Cosmopolitan to a depth of 160 meters. Plesiotypes.—Cat. Nos. 68637-68640, U.S.N.M.

SMITTINA MALEPOSITA, new species.

Plate 31, figs. 10, 11.

Description.—The zoarium incrusts shells. The zooccia are distinct, separated by a salient thread, little elongate, hexagonal, poorly oriented; the frontal is little convex and formed of a granular pleurocyst surrounded by a row of areolar pores separated by very short costules. The apertura is semilunar with a very short proximal lyrule; the peristomic is of little depth and the peristomice is suborbicular. The ovicell is large, globular, finely perforated. On the median line of the frontal there is a small thin avicularium the beak of which is directed toward the bottom.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \left\{ \begin{matrix} ha = 0. \ 10 \ \text{mm.} \\ la = 0. \ 10 \ \text{mm.} \end{matrix} \right. \quad \text{Zooccia} \left\{ \begin{matrix} Lz = 0. \ 40 \ \text{mm.} \\ lz = 0. \ 32 \ \text{mm.} \end{matrix} \right.$

Affinities.—This species is quite close to Smittina trispinosa Johnston, 1838. It is distinguished from it by its somewhat larger peristomice and by the place of its avicularium, always placed in the vicinity of the median line of the zooecium. The position of this avicularium is everywhere rather variable, but its presence is constant. There is therefore a group of Smittina in which the avicularium is not directly supported on the lyrule. We do not believe that this character should be generic, for it does not appear to correspond to an essential and different function.

Occurrence.—Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida (very rare). Pleistocene or Recent: Vero, Florida (rare).

Cotypes.—Cat. Nos. 68641, 68642, U.S.N.M.

SMITTINA OPIIIDIANA Waters, 1877.

Plate 3, fig. 3.

1879. Smittina reticulata, var ophidiana Waters, On the Bryozoa (Polyzoa) of the Bay of Naples, Annals and Magazine of Natural History, ser. 5, vol. 3, p. 40, pl. 9, fig. 1.

1885. Smittina reticulata, var. ophidiana Waters, On the use of the avicularium mandible in the determination of Chilostomatous Bryozoa, Journal Royal Microscopical Society, ser. 2, vol. 5, pl. 14, fig. 6.

1889. Smittia ophidiana Waters, Bryozoa from New South Wales, Annals and Magazine of Natural History, ser. 6, vol. 4, pl. 3, fig. 19.

1903. Smittia ensifera Jullien, Bryozoaries provenant des Campagnes de l'Hirondelle (1886-1888), Resultats des Campagnes Scientifiques du Prince de Monaco, fasc 23, p. 102, pl. 12, fig. 4.

1904. Smittia ophidiana Waters, Resultats Voyage Belgica, Zoologie Bryozoa, p. 66.

1907. Smittia ophidiana Calvet, Expédition scientifiques du Travailleur et Talisman, Bryozoaires, p. 433 (bibliography).

Affinities.—This species is very close to Smittina marmorea but differs from it in the presence of costules and of areolar pores close together (and not scattered punctations) and the constant presence of two frontal pores.

Occurrence.—Miocene (Bowden marl): Bowden, Jamaica (very rare).

Geographic distribution.—Mediterranean at Naples; Atlantic at the Azore Isles (80–130 meters).

Plesiotype.—Cat. No. 68643, U.S.N.M.

SMITTINA GRANDICELLA, new species.

Plate 37, figs. 10-12.

Description.—The zoarium incrusts shells in one or more layers. The zooecia are distinct, separated by a furrow, elongate, elliptical, large; the frontal is convex and formed of an olocyst surmounted by a pleurocyst. The apertura is semi-elliptical; the lyrule is short and wide; the peristome is little salient, hardly indented in the proximal portion, and bears three distal spines.

Affinities.—This species much resembles Smittina trispinosa Johnston, 1838, for it often bears a large avicularium on the line of areolar pores; it is distinguished from it in its large zooccial dimensions and in the much less regular arrangement of the zooccia. The gemmation is identical, but the lines of the nonseptular walls are less rigorously rectilinear and often sinuous (fig. 12). The beak of the avicularium is round and salient.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (rare). Cotypes.—Cat. No. 68644, U.S.N.M.

SMITTINA DISCOIDEA, new species.

Plate 38, figs. 1-7.

Description.—The zoarium is unilamellar, discoidal, very large; the lower face is concentrically wrinkled and the nonseptular walls are visible in the form of radial costules somewhat salient. The zooecia are little distinct, elongate, more or less marginate; the frontal is very little convex and formed of an olocyst surmounted by a detachable pleurocyst with large interareolar costules. The apertura is semilunar; the lyrule is short and lamellose; the peristome is not salient, and the peristomic is excavated in the thickness of the frontal wall. The ovicell is globular, salient, deeply embedded in the distal zooecium; it is surrounded by a little salient collar. On the line of the arcolar pores there is an enormous triangular avicularium with pivot or with condyles of which the beak is acuminate, salient, and pointed upward.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.09 \text{ mm.} \\ la = 0.10 - 0.12 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.50 - 0.60 \text{ mm.} \\ lz = 0.25 \text{ mm.} \end{cases}$

Affinities.—This species is of the group of Smittina trispinosa Johnston, 1838, in all its characters; it differs from it in the size, the form, and the direction of its avicularium. This avicularium is sometimes interzooecial (fig. 2). We are ignorant of the causes which result in the variations in size, direction, and form of the avicularia. These biologic variations are still to be studied. On the ovicelled zooecia the large avicularium is often transformed into two smaller ones (fig. 3). In the interior (fig. 7) the lyrule is laterally accompanied by two kinds of condyles, probably replacing the small cardelles.

Occurrence.-Pleistocene: Santa Monica (Rustie Canyon), California (common).

Cotypes.—Cat. No. 68645, U.S.N.M.

SMITTINA CALIFORNIENSIS Robertson, 1908.

Plate 37, figs. 7-9.

1998. Smittia californiensis Robertson, The incrusting cheilostomatous Bryozoa of the west coast of North America, University of California Publications Zoology, vol. 4, p. 303, pl. 22, fig. 71.

Measurements.—Apertura $\begin{cases} ha = 0.12 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.70 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Variations.—We do not know all the variations of Miss Robertson's species; we are therefore not exactly certain of our determination. However, our specimens bear the principal characters. The peristome is little salient; it frequently bears two large lateral lips, leaving between them a false rimule (fig. 7). We have found some zooecia bearing a large, lateral avicularium; the latter is replaced by two smaller, thin avicularia whose beak is turned toward the bottom (figs. 8, 9,) or indeed one toward the top and the other toward the bottom. The interareolar costules are thin (fig. 7) or very thick (fig. 8). The interzooecial avicularium bears a rectilinear pivot. The zoarium is bilamellar while that in Miss Robertson's species is incrusting.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Habitat.—Pacific, off California.

Plesiotypes.—Cat. No. 68646, U.S.N.M.

SMITTINA(?) BREVIS Canu and Bassler, 1919.

Plate 2, figs. 20, 21.

1919. Smittina? brevis Canu and Bassler, Geology and Palcontology of the West Indies, Bryozoa,
Publication of the Carnegie Institution of Washington, No. 291, p. 93, pl. 2, fig. 2.

Description.—The zoarium is bilamellar with fronds somewhat compressed. The zooeeia are little distinct, elongate, elliptical; the frontal is convex and very short and it bears below a large salient avicularium with rounded beak. The apertura is elliptical and deep. The ovicell is globular, little salient, ornamented by a fragile circular area; it opens into the peristomie.

Affinities.—This species is quite unique and difficult to classify. We have put it in Smittina because the ovicell opens into the peristomic formed by the thicken-

ing of the frontal walls; but it does not really belong to this genus because it has no lyrula nor oral avicularium and because the apertura is not semilunar.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (rare).

Holotype.—Cat. No. 68647, U.S.N.M.

SMITTINA PORIFERA Hincks, 1884.

Plate 38, fig. 9.

1884. Porella marsupium forma porifera Hincks, Report on Polyzoa of Queen Charlotte Islands, Annals Magazine Natural History, ser. 5, vol. 13, p. 50 (sep. 24), pl. 4, fig. 4.

1887. Porella marsupium, var. porifera Waters, Tertiary Cheilostomatous Bryozoa from New Zealand, Quarterly Journal Geological Society London, vol. 42, p. 63.

Description.—The zoarium is incrusting. The zooecia are distinct, swollen, separated by a deep furrow, very convex, perforated by some pores widely spaced. The frontal is a pleurocyst garnished laterally with six large areolar pores, of which the two superior ones are sometimes changed into avicularia. The apertura is deep, hidden by the peristomic and by a small, salient, orbicular, oral avicularium. The peristome is thin and bears four small spines which are often replaced by a small distal orbicular avicularium (rarely two). There is a very small lyrula hidden by the frontal avicularium. The ovicell is globular, salient, smooth; it opens above the operculum by a very large opening.

Measurements.—Apertura
$$\begin{bmatrix} ha = 0.12 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{bmatrix}$$
 Zooecia $\begin{bmatrix} Lz = 0.45 \text{ mm.} \\ lz = 0.28 \text{ mm.} \end{bmatrix}$ The avicularium chamber forms a frontal gibbosity perforated by four large

The avicularium chamber forms a frontal gibbosity perforated by four large pores. The latter characterize the species, for they do not exist in *Porella marsupium* MacGillivray.

Occurrence.—Pleistocene: Santa Monica (Long Wharf Canyon), California (rare). Geological distribution.—Pliocene of New Zealand (Waters).

Habitat.—Queen Charlotte Islands (Hincks).

Plesiotype.—Cat. No. 68648, U.S.N.M.

Genus PORELLA Gray, 1848.

(For description, see Bulletin 106, U.S. National Museum, p. 479.)

PORELLA BELLA Busk, 1860.

Plate 1, fig. 12.

1860. Lepralia bella Busk, Zoophytology, Quarterly Journal Microposcopical Science, vol. 8, pl. 27, fig. 2, 3.

1873. Escharella landsborovi Smitt, Floridan Bryozoa, pt. 2, Kongl. Seven. Vet. Akad. Hand., vol. 11, no. 4, p. 60, pl. 10, figs. 201, 202.

1880. Smittia bella Hincks, British Marine Polyzoa, p. 352, pl. 42, figs. 9, 10.

1919. Porella bella Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution, no. 291, p. 93, pt. 1, fig. 15.

We are not certain that Busk's species is identical with that of Smitt, as the former author represents it, but is evident that the characters of our Antigua fossil, although it is poorly preserved, are in harmony with Smitt's figures. Following are the measurements of this fossil.

$$\label{eq:apertura} \text{Apertura} \begin{cases} ha = 0.16 \text{ mm.} \\ la = 0.20 \text{ mm.} \end{cases} \quad \text{Zooccia} \begin{cases} Lz = 0.90 \text{ mm.} \\ lz = 0.70 \text{ mm.} \end{cases}$$

Occurrence.—Oligocene (Antigua formation): Rifle Butts, Antigua, Leeward Islands (rare).

Habitat.—Shetland Islands. Waters off Florida (185 meters). Plesiotype.—Cat. No. 68649, U.S.N.M.

PORELLA REVERSA Uirlch and Hassler, 1904.

Plate 23, figs. 5-10.

1904. Lepralia? reversa Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 426, pl. 113, figs. 1, 2.

The original description is as follows:

Zearium forming parasitic patches, several centimeters in diameter and composed of a single layer, en shells. Zeecia obleng quadrate er subhexagenal, generally arranged in rather regular lengitudinal and diagonally intersecting rows, each about 0.5 mm. in length and 0.3 mm. in width. Orifice rather large, rounded-quadrate, enclosed by a peristeme of mederate thickness and elevation. Peristemes divided into two parts, anterior and posterior, the former either straight or slightly arcuate and not so prominent as the horseshee-shaped pertion enclosing the sides and proximal margins of the crifice. Distal extremities of the latter portion of the peristome often a little thickened and projecting slightly inward. Just behind the preximal border of the orifice there is constantly a rather small but preminently elevated and thick-walled avicularium, opening obliquely forward. Remainder of front zeeccia with from one to three rows of large pores. Frequently adjoining zeeccia are separated by a thin raised line. Ovicells moderately convex, rather large, with a central pore and one or two somewhat radially disposed marginal rows of smaller peres. When broken they leave a sharply defined cencave space in front of the orifice, slightly exceeding the latter in size.

The division of the peristeme into two parts as described is unusual and produces the probably false appearance of a reversal of the ends of the operculum that has suggested the specific name. If it could be proved that the hinge of the operculum was really on the distal side of the orifice instead of the proximal, then this species would be distinct enough to justify the erection of a new genus for its reception; but until this unusual condition can be demenstrated we think it well to regard it as related to such species as Lepralia pallasiana. We know of none resembling it closely enough to require unusual care in its discrimination.

$$\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{vmatrix} ha = 0.15 \text{ mm.} \\ la = 0.13 \text{ mm.} \end{vmatrix}$$
 Zooccia
$$\begin{vmatrix} Lz = 0.45 - 0.55 \text{ mm.} \\ lz = 0.25 - 0.30 \text{ mm.} \end{vmatrix}$$

A reexamination of the type specimen shows this species should be classed in the genus *Porella*. It is well characterized by its embedded aperture, by its infraoral avicularium, and by its ovicell perforated like the frontal. Finally the interior shows the parietal arrangement habitual in the Smittinidae (fig. 10).

Occurrence.—Miocene (St. Mary's formation): Cove Point, Maryland (rare). Miocene; Kulins, Carteret County, North Carolina (rare).

Cotypes and Plesiotype.—Cat. Nos. 68650, 68651, U.S.N.M.

PORELLA COLLIFERA Robertson, 1908.

Plate 38, figs. 10-15.

1908. Porella collifera Robertson, The incrusting cheilestematous Bryozoa of the west coast of North America, University of California Publications, Zeolegy, vol. 4, no. 5, p. 304, pl. 23, fig. 72.

Measurements.—Apertura
$$\begin{vmatrix} ha = 0.24 \text{ mm.} \\ la = 0.20 \text{ mm.} \end{vmatrix}$$
 Zooecia $\begin{vmatrix} Lz = 0.80 - 1.00 \text{ mm.} \\ lz = 0.50 - 0.60 \text{ mm.} \end{vmatrix}$

Variations.—The zoarium incrusts pebbles. The peristome is very salient especially in the proximal portion. The oral avicularium rests on a wide but little salient lyrule; it is often only visible by a suitable inclination of the preparation

so that the bottom of the peristome can be seen. The zooecia are sometimes bordered by a salient thread (fig. 13). The ovicell is very small (fig. 11). In the the interior the olocyst is perforated by very small pores corresponding to the large widened pores of the superposed tremocyst.

Occurrence.—Pleistocene: Santa Monica (rare) and Santa Barbara, California (common).

Habitat.—Pacific: Coronado Islands (24-29 meters).

Plesiotypes.—Cat. No. 68652, U.S.N.M.

Genus PALMICELLARIA Alder, 1864.

1864. Palmicellaria Alder, Description New British Polyzoa, Quarterly Journal Microscopical Society, new ser., vol. 4, p. 100.

The apertura is orbicular, semicircular, or semielliptical. Neither lyrula nor cardelles are present. The ovicell opens in the peristomie. The frontal is a granular pleurocyst, surrounded with areolar pores. The peristomie is much developed. Often an avicularian mucro partially hides the apertura.

Genotype.—Palmicellaria elegans Alder, 1864.

Range.—Miocene to Recent.

The known species of this genus are:

Palmicellaria elegans Alder, 1864.

Palmicellaria (Millepora) skenei Solander, 1786.

Palmicellaria (Eschara) lorea Alder, 1864.

Palmicellaria inermis Jullien, 1882.

Palmicellaria tenuis Calvet, 1907.

Structure.—The species of this genus are garnished with areolar pores and they therefore do not belong to the genus Porella Gray, 1848, as we have limited it. The absence of the lyrula and of cardelles distinguishes it from Smittina.

Historical.—For a long time, on the authority of Waters, the species of this genus have been classed in the genus Porella Gray, 1848, because the known mandibles are identical. The opercula and mandibles of Palmicellaria skenei Solander, 1786, are indeed those of Porella. The mandible of P. lorea Alder, 1864, is also that of Porella, but the operculum is semielliptical and of quite a divergent type from other Porellas. The chitinous appendages of the genotype and of the other species are unfortunately unknown.

All of the species cited having without exception the same characteristic of calcification, namely, a granular pleurocyst surrounded by areolar pores, we believe that it is necessary to maintain Alder's genus, although it does not appear to us very well marked.

PALMICELLARIA cf. INERMIS Jullien, 1882.

Plate 4, fig. 13.

Our figured specimen is too incomplete to be named with certainty, but we are introducing it to call attention to the genus in the early Miocene. This specimen is deprived of ovicell and its aperture is invisible. It approaches *Palmiccllaria incrmis* Jullien, 1882, but it differs from it in its larger micrometric measurements and in its apertura, which does not appear transverse.

Occurrence. - Miocene (Bowden horizon): Santo Domingo.

Plesiotype.—Cat. No. 68653, U.S.N.M.

PALMICELLARIA COSTULATA, new species.

Plate 45, figs. 12, 13.

Description.—The zoarium incrusts corals. The zooccia are distinct, separated by a broad furrow, little elongated; the frontal is very convex, thick, covered by a tremocyst with large pores, garnished with broad radiating costules, and ending in a very salient avicularium. The apertura is semielliptical, transverse, deeply buried, partially hidden by the avicularian mucro. The ovicell is deeply embedded in the distal zooccium; it is large, little salient, similarly covered with large radiating costules.

Measurements.—Apertura
$$\begin{cases} ha = 0.09 \text{ mm.} \\ la = 0.13 \text{ mm.} \end{cases}$$
 Zooccia $\begin{cases} Lz = 0.45 - 0.50 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$

Affinities.—In the vicinity of the ancestrula the avicularian mucro is not constant; it is often replaced by small salient orbicular avicularia irregularly arranged around the apertura.

This beautiful species on account of its frontal irregularities is very difficult to photograph. Better preserved specimens, moreover, will probably show new characters necessitating its classification in another genus. The species is certainly shorter than *Lepralia corrugata* MacGillivray, 1895, which has some resemblance.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare).

Cotypes.—Cat. No. 68654, U.S.N.M.

PALMICELLARIA CONVOLUTA Ulrich and Bassler, 1904.

Plate 23, figs. 1-3.

1904. Palmicellaria convoluta Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 427, pl. 16, figs. 2-4.

The original description of this species is as follows:

Zoarium erect, forming loose masses 3 cm. or more in diameter, consisting of broad, bifoliate, convoluted, anastomosing leaves, 1.0 mm. or more in thickness. Zooecia prominent, distinct, oval, rhomboidal or hexagonal, arranged in irregular quincunx, averaging between 0.9 mm. and 1.0 mm. in length and about 0.45 mm. in width; surface rather coarsely punctured; orifice terminal, the proximal edge overhung by a prominent mucro containing an avicularium the sagittate opening of which is divided into two unequal parts by a septum and lies on the abrupt distal slope of the mucro so as to be nearly or entirely concealed in a front view. When the apex of the mucro is worn or broken away the cavity of the avicularium is exposed to view as a cell immediately behind the orifice and almost equaling it in size. Ovicells small, transverse, bulbous, closely united to the cell next above.

$$Measurements.$$
—Apertura $\begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.20 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz = 0.85 - 1.00 \text{ mm.} \\ lz = 0.45 - 0.50 \text{ mm.} \end{cases}$

The essential features of this species are noted in the original description. Occurrence.—Miocene (Calvert formation): Reeds, Maryland (rare). Cotypes.—Cat. No. 68655, U.S.N.M.

PALMICELLARIA PUNCTATA Ulrich and Bassler, 1904.

Plate 23, fig. 4.

1904. Palmicellaria punctata Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 428, pl. 116, fig. 1.

Only the type specimen of this species, a fragment of bifoliate zoarium, is known, and our studies upon it must remain incomplete. This specimen shows

punctate zooccia with a subcircular orifice and a mucro like the preceding, but its zooccia are much smaller than *Porella* (*Palmicellaria*) convoluta Ulrich and Bassler, 1904, of the Maryland Miocene, and its mucro is less prominent. The ovicells however are relatively larger and longer.

Measurements.—Apertura a = 0.10 mm. Zooecia, irregular.

Occurrence.—Miocene (Calvert formation): Reeds, Maryland (very rare). Holotype.—Cat. No. 68656, U.S.N.M.

Genus RHAMPHOSTOMELLA Lorenz, 1886.

(For description see Bulletin 106, U. S. National Museum, p. 476.)

RHAMPHOSTOMELLA LATICELLA Canu and Bassler, 1919.

Plate 5, fig. 13.

1919. Rhamphostomella laticella Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publicatious of the Carnegie Institution of Washington, no. 291, p. 94, pl. 5, fig. 12.

The figured specimen is the only one found; it is unfortunately incomplete and worn and does not permit a detailed description. It incrusts a shell; the areolar costules are rare or little visible. The oral avicularium is quite large, oblique, the point directed toward the bottom.

Occurence.—Lower Miocene (Bowden marl): Bowden, Jamaica (very rare). Holotype.—Cat. No. 68657, U.S.N.M.

RHAMPHOSTOMELLA GRANULOSA, new species.

Plate 3, fig. 2.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a deep furrow, elongated, elliptical; the frontal is very convex, covered with a granular pleurocyst and bordered with small arcolar pores. The apertura is large, elongated, and deformed by a small, elongated, triangular avicularium arranged obliquely. The ovicell is globular, very salient, and covered also with a granular pleurocyst.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.55 \text{ mm.} \\ lz = 0.25 - 0.30 \text{ mm.} \end{cases}$

Affinities.—The zooecia are very irregular in their micrometric measurements. The areolar pores are often hidden by the pleurocyst.

This species approaches *Rhamphostomella brendolensis* Waters, 1891, which is often deprived of costules, but differs from it in the presence of the frontal granules.

Occurence.—Miocene (Bowden marl): Bowden, Jamaica (very rare).

Holotype.—Cat. No. 68658, U.S.N.M.

Genus CYSTISELLA Canu and Bassler, 1917.

(For description, see Bulletin 106, U. S. National Museum, p. 479.)

CYSTISELLA AVICULIFERA, new species. '

Plate 38, fig. 8.

Description.—The zoarium incrusts pebbles or algae. The zooecia are distinct, clongate, subcliptical, swollen: the frontal is convex, smooth or bears an avicularian chamber rather large and somewhat salient. The apertura is semicircular and placed at the base of a deep peristomie; the peristome is thin, salient, and bears two or three orbicular avicularia. The ovicell is large, hyperstomial, convex, globular, smooth; it opens into the peristomie by a very large opening placed above the operculum.

 $\label{eq:measurements} \textit{Measurements.} - \text{Peristomice} \begin{cases} hpe = 0.15 - 0.18 \text{ mm.} \\ lpe = 0.16 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.65 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$

Affinities.—Our micrometric measurements are taken on the marginal zooecia; at the center of the zoarium they are much smaller, the frontal of the zooecia being much reduced there. The ancestrular zooecia appear to be erect as in the Celleporidae; this aspect is deceiving and is occasioned by the presence of small peristomial avicularia; they are perfectly oriented like the marginal zooecia but much smaller. The relative constancy of the peristomice permits us to utilize the micrometric measurements. The frontal avicularian chamber is much smaller than in the other species of the same genus; but the absence of the areolar pores leaves no doubt as to the exactness of our generic determination. The multiplicity of avicularia on a given species always reveals whether the water was calm or not very agitated.

Occurrence.—Pleistocene: Santa Barbara (rare) and Santa Monica, California (rare).

Holotype.—Cat. No. 68659, U.S.N.M.

Family RETEPORIDAE Smitt, 1867.

Genus RETEPORA Imperato, 1599.

(For description, see Bulletin 106, U. S. National Museum, p. 500.)

RETEPORA DOVERENSIS Ulrich and Bassler, 1904.

Plate 23, figs. 11-17.

1904. Retepora doverensis Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 422, pl. 111, figs. 5-7; pl. 115, figs. 1-5.

The original description follows:

Zoarium reticulate, fenestrae of variable size, averaging 0.8 mm. long and about 0.4 mm. wide. Branches varying from 0.2 mm. to 1.0 mm. wide, usually about 0.6 mm.; reverse solid, the surface generally smooth, occasionally minutely granulose, divided into irregular angular spaces by fine impressed or raised lines that may or may not correspond with the bases of the zooccial walls. Zooccia immersed, with oblique imbricating and slightly flaring mouths, and a narrow notch in the elevated and angular proximal border. The appearance of the celluliferous surface varies greatly in different specimens, the difference being due chiefly to the presence or absence and number of the avicularia and ovicells. The latter appear as bulbous inflations with a slit running from the center to one edge. When they occur at all it is usually in abundance. Of avicularia there are at least two sets, the larger ones occurring on the front of the zooccia, forming its highest part and causing it to appear inflated.

Those of the smaller set occur in the depressed spaces, usually close beside the zooecial orifice, and rarely on the reverse of the branches.

This common species of the Choptank formation apparently belongs to the *Retepora-cellulosa* group of Waters. Though resembling several of the species of the group rather closely we could not decide that it was any nearer to one than to another. Still, having found it difficult to make thoroughly satisfactory comparisons, we will not be greatly surprised should future study prove *R. doverensis* to belong to some previously described species.

The peristome bears a wide, very irregular, rimule spiramen. The ovicell is fissured.

Occurrence.—Miocene (Choptank formation): Dover Bridge and Jones Wharf, Maryland (common).

Cotypes and plesiotypes.—Cat. Nos. 68660, 68661, U.S.N.M.

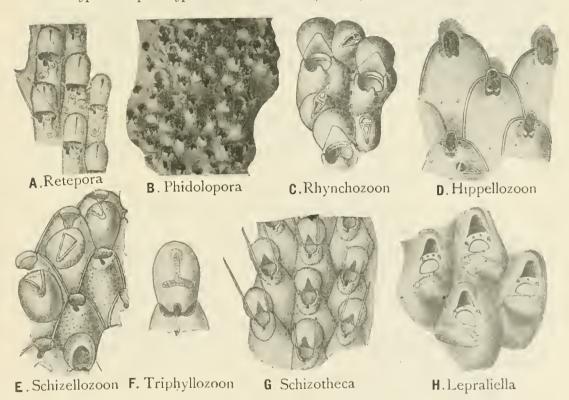


Fig. 27.—Genera of the family Reteporidae Smitt, 1867.

A. Retepora beaniana King, 1846, × 25. Recent. (After Hincks, 1880.) B. Phidolopora labiata Gabb and Horn, 1865, × 20. Pleistocene of California. C. Rhynchozoon angulatum Levinsen, 1909, × 55. Recent. (After Levinsen, 1909.) D. Hippellozoon novaezelandiae Waters, 1894, × 25, Recent. (After Waters, 1894.) E. Schizellozoon imperati Busk, 1884, × 25. Recent. (After Waters, 1885.) F. Triphyllozoon moniliferum MacGillivray, 1860, × 85. Recent. (After Waters, 1894.) G. Schizotheca fissa Busk, 1858, × 45. Recent. (After Hincks, 1880.) H. Lepraliella contigua Smitt, 1867. Recent. (After Levinsen, 1916.)

12184-23-Bull, 125-11

Genus PHIDOLOPORA Gabb and Horn, 1862.

1862. Phidolopora Gabb and Horn, Monograph Polyzoa of the Secondary and Tertiary Formations of North America, Journal Academy National Science Philadelphia, ser. 2, vol. 5, p. 138.

The frontal of the ovicell is not fissured. The aperture is semilunar, with a concave proximal border. The peristomice bears a rimule spiramen. The frontal is an oloeyst. No labial avicularium.

Genotype.-Retepora labiata Gabb and Horn, 1862. Pleistocene, Recent.

Affinities.—The apertura is identical with that of the genus Retepora Imperato, 1859, as Waters has limited it in 1913. The genus Phidolopora Gabb and Horn, 1862, differs from it in the absence of a fissure on the ovicell, in the replacement of the spiramen by a rimule of the peristomice, and in the absence of the labial avicularium. Moreover, the frontal is an olocyst, a character which does not exist notably in the group of Sertella Jullien, 1903. The definition of Gabb and Horn 1862, has no significance; it is even little comprehensible: "It differs from Retepora by the scattered special pores over the surface of the colony." Retepora pacifica Robertson, 1908, belongs absolutely to the same group and shows the generic characteristics clearly.

PHIDOLOPORA PACIFICA Robertson, 1908.

Plate 39, figs. 1-7.

1908. Retepora pacifica Robertson, The incrusting cheilostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 4, p. 310, pl. 24, figs.

$$\label{eq:loss_loss} \begin{aligned} & \textit{Measurements.} - \text{Peristomice: } lpe = 0.14-0.16 \text{ mm.} \\ & \textit{Zooecia} \begin{vmatrix} Lz = 0.40-0.60 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{vmatrix} \end{aligned} \end{aligned} \\ & \text{Fenestrae} \begin{vmatrix} Lf = 1.75 \text{ mm.} \\ lf = 0.50 \text{ mm.} \end{aligned}$$

Affinities.—This species differs from Phidolopora labiata Gabb and Horn, 1862, in its larger peristomice always wider than 0.14 mm.; thus the apertures appear closer together. The small tongue on the ovicell mentioned by Miss Robertson is too fragile to be preserved on the fossils. The zooecia deprived of the large frontal avicularium bear some small scattered pores. In tangential section the vibices appear clearly. They are therefore formed by a very dense calcification in the entire thickness of the zoarial dorsal. Sometimes the dorsal bears radicular pores. In transverse sections the lacunae appear very irregularly arranged.

Occurrence.—Pleistocene: Santa Monica (rare), and Dead Mans Island, off San

Pedro (rare), California.

Habitat.—Coast of California.

Plesiotypes.—Cat. No. 68662, U.S. N. M.

PHIDOLOPORA LABIATA Gabb and Horn, 1862.

Plate 39, figs. 13-17.

1862. Phidolopora labiata Gabb and Horn, Monograph Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy Natural Sciences, Philadelphia, ser. 2, vol. 5, p. 138, pl. 19, fig. 21.

Description.—The zoarium is reticulated; the fenestrae are narrow and elongated. The zooecia are distinct, margined by a salient thread, elongate, fusiform; the frontal is slightly convex and smooth. The apertura (visible only in the inte-

rior) is transverse, subelliptical; the peristomice bears a rimule spiramen arranged between two salient lips. The ovicell is large, convex, smooth, buried in the distal zooccium; its orifice is very large and located in front of the two lips. Many zooccia bear a very large avicularium arranged obliquely on the frontal and provided with a curved, very salient bcak. The zooecia without avicularia bear some small scattered pores.

$$\label{eq:lemma:lemma:decomposition} \begin{split} & \textit{Measurements.} - \text{Peristomice: } lpe = 0.10 \text{ mm.} \\ & \text{Zooecia} \begin{cases} Lz = 0.46 - 0.60 \text{ mm.} \\ lz = 0.20 - 0.30 \text{ mm.} \end{cases} \end{split}$$
 Fenestrae
$$\begin{cases} Lf = 2.00 \text{ mm.} \\ lf = 0.30 - 0.75 \text{ mm.} \end{cases}$$

Affinities.—This species differs from Phidolopora pacifica Robertson, 1908, in the smaller dimensions of its peristomice, which measure more than 0.10 mm. The general aspect is therefore that of bearing apertures much more scattered and in having a peristomice much smaller than the zooecial width.

The micrometric variations are extraordinary. On the same zoarium one may find zooecia 0.50 mm, by 0.20 mm, with zooecia 0.46 by 0.26-0.30 mm. Frequently the peristomice of the ovicelled zooecia is somewhat wider than that of the other zooecia. The vibices are also very irregular; thay are salient, but disappear easily in fossilization.

Occurrence.—Pleistocene: Dead Mans Island off San Pedro (rare), Santa Monica (common), and Santa Barbara, California (very common).

Plesiotypes.—Cat. No. 68663, U.S.N.M.

Genus RHYNCHOZOON Hincks, 1891.

(For description, see Bulletin 106, U.S. National Museum, p. 506.)

RIIYNCHOZOON VAUGHANI Cann and Bassler, 1919.

1919. Rhynchozoon vaughani Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 94, pl. 2, fig. 23; pl. 4, figs. 1, 2.

Description.—The zoarium is unilamellar. The zooecia are large, distinct, hexagonal or claviform; the frontal is smooth, convex, surrounded by a line of small scattered areolar pores. The apertura is large, suborbicular, with a wide proximal rimule; the peristomie is deep and widened; the very irregular peristome bears some large tubercles; the false rimule is compressed between two large tuberosities, one of which bears an oral avicularium transversely arranged.

Measurements.—Apertura
$$\begin{cases} ha = ? \\ la = 0.20 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.50 \text{ mm.} \end{cases}$

Affinities.—This species differs from Rhynchozoon solidum Osburn, 1914, which it much resembles, in its much larger micrometric measurements and in the absence of the small frontal avicularium.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (very rarc).

Cotypes.—Cat. No. 68664, U.S.N.M.

RHYNCHOZOON CURTUM, new species.

Plate 4, fig. 14.

Description.—The zoarium is unilamellar. The zooecia are distinct, separated by a deep furrow, elongated, very short, much narrowed behind; the frontal is very small, somewhat convex, surrounded with aereolar pores; it bears a small elliptical transverse avicularium. The apertura, hidden at the bottom of the peristomie, is little visible; it bears a wide rounded rimule. The peristome is salient, wide, formed of tuberosities smooth or perforated; it is notched in its proximal part by a triangular pseudorimule placed in the median axis of the zooecium. The ovicell is small, short, deeply buried, recumbent convex and transverse.

Measurements.—Opercula
$$\begin{cases} ha = 0.14 \text{ mm.} \\ la = 0.16 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Affinities—This species differs from Rhynchozoon verruculatum Smitt, 1872, in its very short zooecia, in its elliptical and always acuminate avicularium, the length of which never surpasses 0.12 mm. In its general aspect the peristome appears to occupy half of the zooecium, which thus reduces considerably the length of the frontal and which gives such a short appearance.

Occurrence.—Miocene (Bowden horizon): Santo Domingo.

Holotype.—Cat. No. 68665, U.S.N.M.

RHYNCHOZOON GRANDICELLA, new species.

Plate 47, figs. 7, 8.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a deep furrow, large, elongated, elliptical. The frontal is a costulated pleurocyst surrounded by small areolar pores; it is covered by an avicularian chamber forming a truncated tongue above the apertura. The apertura is very oblique, transverse, with a wide proximal rimule. The peristomice is very oblique and bears an eccentric pseudorimule to the right or left of the avicularian mucro. The ovicell is small, globular, opening into the peristomie by a wide orifice. The ancestrular zooecia are smaller than the ordinary ones.

Measurements.—Apertura
$$\begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.20 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.85 - 0.90 \text{ mm.} \\ lz = 0.60 - 0.70 \text{ mm.} \end{cases}$

Variations.—The apertura is visible only when the specimen is much inclined. The interareolar costules are little salient; on the zoarial margin they are visible only under special illumination.

Affinities.—This species differs from Rhynchozoon longirostris Hincks, 1881, also provided with large zooecia, in its dimensions greater than 0.70 mm. and in the absence of a frontal avicularium. It differs from Rhynchozoon corrugatum Thornely, 1905, also deprived of frontal avicularia in its much larger dimensions, in its interareolar costules, and in its ovicell wanting the two areas.

Occurrence.—Pleistocene: Rustic Canyon, Santa Monica, California (rare). Cotypes.—Cat. No. 68666, U.S.N.M.

RHYNCHOZOON(?) LEVIGATUM, new species. Plate 45, fig. 11.

Description.—The zoarium incrusts shells. The zooecia are distinct, elongate, ovoid, separated by a furrow; the frontal is very convex, smooth or very finely granular. The apertura is oval; the peristomice bears interiorly an excentric pseudolyrula; the peristome is thick and salient. The ovicell is globular, salient, attached to the peristome; its orifice is large, placed in the peristomie, and can not be closed by the operculum.

 $\begin{array}{lll} \textit{Measurements.} & -\text{Apertura} \; \left\{ \begin{matrix} ha = 0.12 \; \text{mm.} \\ la = 0.10 \; \text{mm.} \end{matrix} \right. & \text{Zooecia} \; \left\{ \begin{matrix} Lz = 0.50 - 0.75 \; \text{mm.} \\ lz = 0.25 \; \text{mm.} \end{matrix} \right. \\ & \text{Affinities.} & -\text{The excentric spinule in the peristome characterizes this species.} \end{array} \right.$

Affinities.—The excentric spinule in the peristome characterizes this species. It differs from Schizoporella argentea Hincks, 1882, in the absence of a large oral avicularium. The species differs from Mucronella contorta Busk, 1882, in its nonperforated frontal. It differs from Rhynchozoon tuberculatum Osburn, 1914, in the absence of scattered tuberosities on the frontal.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (very rare). Holotype.—Cat. No. 68667, U.S.N.M.

RHYNCHOZOON VERRUCULATUM Smitt, 1872.

Plate 3, figs. 11-13.

1872. Escharella verruculata Smitt, Floridan Bryozoa, pt. 2, Kongl. Svenska Vetenskaps-Akademiens Handlingar, no. 4, p. 50, pl. 8, figs. 170-172.

1879. Cellepora verruculata Waters, On the Bryozoa of the Bay of Naples, Annals and Magazine of Natural History, ser. 5, vol. 3, p. 193, pl. 14, figs. 1-7.

1884. Escharoides verruculata Busk, Report on Polyzoa collected by the Challenger, vol. 10, pt. 30, p. 150.

1885. Cellepora verruculata Waters, On the use of the avicularian mandible, with additional list of the Naples fauna, Journal Royal Microscopical Society, ser. 2, vol. 5, pl. 14, fig. 37.

1902. Cellepora verruculata Calvet, Bryozoaires marins de la région de Cette, Travaux de l'Institut de Zoologie de L'Université de Montpellier, mem. 11, p. 66, pl. 2, figs. 6-9.

1902. Cellepora verruculata CALVET, Bryozoaires marins des côtes de Corse, Travaux de l'Institut de Zoologie de L'Université de Montpellier, mem. 12, p. 37.

1914. Cellepora verruculatum Osburn, Bryozoa of the Tortugas Islands, Publication Carnegie Institution of Washington, no. 192, p. 214.

Measurements.—Apertura $\begin{cases} ha = ? \\ la = 0.12 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.45 - 0.50 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Variations.—The marginal zooecia are well oriented; the central zooecia are not, which has caused the species to be classified incorrectly in Cellepora. The frontal avicularium is transverse, much elongated, with triangular mandible. The peristome is lobed or crowned by from three to six tubercles. One of the two lips of the pseudorimule of the peristomice often bears a small avicularium. The irregularity of this species defies all description; the absence of limits between the zooecia appears to be the cause.

We are not entirely certain that the Mediterranean species is that of Smitt; the measurements taken from Calvet's figures appear larger.

Occurrence.—Miocene (Bowden marl): Bowden, Jamaica (rare). Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida (rare). Pleistocene: Simmons Point, Yonges Island, South Carolina (very rare).

Habitat.—Mediterranean at Naples, Cette, Corse. Atlantic: Gulf of Mexico and Tortugas Islands (24 meters). Indian Ocean: Heard Island (121 meters).

Plesiotypes.—Cat. No. 68668, U.S.N.M.

Family ADEONIDAE Jullien, 1903.

Genus ADEONA (Lamouroux, 1816) Levinsen, 1909.

(For description see Bulletin 106, U. S. National Museum, p. 560).

ADEONA POROSA Canu and Bassler, 1919.

Plate 6, figs. 22, 23.

1919. Adeona porosa Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa,
Publications of the Carnegie Institution of Washington, no. 291, p. 94, pl. 6, figs. 22, 23.

Description.—The zoarium is free, bilamellar. The zooecia are little distinct, very elongate, separated by a hardly visible, salient thread; the frontal is porous, little convex, surrounded by areolar, parietal pores of which one is larger than the others. The ascopore is large, orbicular, median. The apertura and peristomice are semilunar with the proximal border somewhat concave. The gonoecia have two very large symmetrical, areolar pores and irregular, oral gibbosities. The avicularian zooecia have their opesia arranged on the lower part of a pyriform area, calcified and deep.

Measurements.—Apertura
$$\begin{cases} ha = 0.12 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$ Affinities.—As we possess only the figured fragment the study of this species

Affinities.—As we possess only the figured fragment the study of this species is necessarily incomplete. The species is very peculiar, for there is no other one provided with a porous frontal and oral gibbosities at the genoecia.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo (rare).

Cotypes.—Cat. No. 68669, U.S.N.M.

ADEONA HECKELI Reuss, 1847.

Plate 24, figs. 1, 2.

- 1845. Cellepora umbilicata Lonsdale, Report Corals Tertiary formation North America, Quarterly Journal Geological Society, London, vol. 1, p. 507.
- 1847. Cellepora heckeli Reuss, Die fossilen Polyparien des Wiener Tertiarbeckens, Haidinger's naturwissenschaftliche Abhandlungen, vol. 2, p. 85, pl. 10, fig. 10.
- 1849. Lepralia violacca Johnston, A History of the British Zoophytes, p. 325, pl. 57, fig. 9.
- 1862. Multiporina umbilicata Gabb and Horn, Monograph of the fossil Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy of Natural Sciences of Philadelphia, ser. 2, vol. 5, p. 145, pl. 20, fig. 27.
- 1880. Microporella violacea Hincks, History of the British Marine Polyzoa, p. 216, pl. 30, figs. 1-4.
- 1889. Microporella heckeli Jelly, A synonymic catalogue of the recent marine Bryozoa, p. 184 (general synonymy).
- 1895. Microporella (Hcckelia) violacea Neviani, Briozoi fossili della Farnesina, Paleontographia italica, vol. 1, p. 106, (30), pl. 5, (1), figs. 27-29.
- 1900. Microporella heckeli Calver, Contribution a l'Histoire naturelle des Bryozoaires ectoprocts marins, Travaux de l'Institut de Zoologie de l'Université de Montpellier, mem. 8, pl. 7, fig. 1; pl. 8, fig. 15.
- 1901. Microporella (Heckelia) violacea Neviani, Briozoi neogenici della Calabrie, Paleontographia Italica, vol. 6, p. 177 (63), (local bibliography).
- 1902. Microporella heckeli Calvet, Bryozoaires marins de la region de Cette, Travaux de l'Institut de Zoologie de l'Université de Montpellier, vol. 2, mem. 11, p. 39.
- 1902. Microporella heckeli Calvet, Bryozoaires des côtes de Corse, Travaux de l'Institut de Zoologie de l'Université de Montpellier, vol 2, mem. 12, p. 20.
- 1904. Adeonellopsis umbilicata Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 417, pl. 113, figs. 4, 5; pl. 114, fig. 5.

1905. Microporella (Heckelia) violacea Neviani, Briozoi fossili di Carrubare (Calabrie), Bolletino della Societa geologica italiana, vol. 23, p. 525.

1907. Microprella heckeli Calvet, Expédition scientifique du Travailleur et du Talisman, vol. 8, p. 404 (bibliography).

1909. Adeona violacea Levinsen, Morphological and systematic studies on the cheilostomatous Bryozoa, p. 83, pl. 14, fig. 1.

1909. Reptadeonella violacea NORMAN, The Polyzoa of Madeira and neighboring islands, Journal Linnean Society of London, vol. 30, p. 296.

1914. Adeona violacea Osburn, The Bryozoa of the Tortugas Islands, Florida, Publication No. 182, Carnegie Institution of Washington, p. 199.

1915. Microporella heckeli Barroso, Contribucion al conociementa de los Briozoos marinos de l'Espana, Boletin de la real sociedad espanola de Historia naturale, p. 415.

1917. Adeona heckeli Barroso, Notas sobre Briozoos, Boletin de la real sociedad espanola de Historia naturale, p. 498.

1919. Adeona heckeli Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. I, p. 95.

This species is better known to zoologists under the name of Microporella violacea Johnston, 1849. In 1874 Reuss abandoned his right of priority because of his mediocre figure of 1847. Jelly, 1889, did not recognize his withdrawal of the name and authors have employed one or the other name according to their personal preference. On account of the size and position of the large frontal avicularium there has been created a variety, plagiopora Busk, 1859. Osburn, 1914, has shown that on the same specimen this avicularium may be straight or oblique and that it does not necessitate the establishment of a special variety.

The American fossil specimens are very rare; they belong to the normal form with straight avicularium, long since described by Gabb and Horn under the name of Multiporina umbilicata. Of this species we have found only a single specimen in the Bowden marl which is moderately well preserved. In the Duplin marl of the Upper Miocene, however, it is more common but not so abundant as in the

European Miocene and Pliocene.

Occurrence.—Lower Miocene (Bowden marl): Bowden, Jamaica (rare). Lower Pliocene (Caloosahatchee formation): Shell Creek, De Sota County, Florida (common). Upper Miocene (Duplin marl): Wilmington, North Carolina; Natural well, 2 miles southwest of Magnolia, Duplin County, North Carolina; 3 miles southwest of Petersburg, and near Powcan, King and Queen County, Virginia (common).

Geological distribution.—Stampian of Germany (Reuss); Aquitanian of Gironde (Duvergier); Miocene of Australia (Waters); Helvetian of Italy (Seguenza), of Touraine (Canu); Tortonian of Austria-Hungary (Reuss), of Italy (Seguenza, Neviana), of England (Busk); Sicilian of Rhodes (Manzoni, Pergens), of Italy

(Seguenza, Neviani); Quaternary of Italy (Seguenza, Neviani).

Habitat.—Mediterranean: Adriatic (32-89 meters), Naples (54 meters), Algeria, Cette (40-97 meters), Bonifacio (89-92 meters), Balearic Islands, shores of Spain. Atlantic: England, English Channel, Gulf of Gascony, Bay of Cadex (60-97 meters), Cape Verde Islands (118-180 meters), Madeira, Florida (56-97 meters), Tortugas (8-29 meters), Bernuda. Indian Ocean: Burmah. Pacific: Australia, China Sea and Cape Tizard (43 meters).

This species does not extend beyond the fifty-second parallel in Europe and the twenty-second in America. It is never abundant in any locality.

Plesiotypes.—Cat. Nos. 68670-68673, U.S.N.M.,

Genus BRACEBRIDGIA MacGillivray, 1886.

(For description, see Bulletin 106, U. S. National Museum, p. 557.)

BRACEBRIDGIA DEFORMIS Canu and Bassler, 1919.

Plate 8, figs. 11-16.

1919. Bracebridgia deformis Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 97, pl. 3, fig. 11-16.

Description.—The zoarium is free, bilamellar, compressed, bifurcated. The zooecia are distinct, elongate, separated by a deep furrow, claviform or elliptical; the peristomice is oval or elliptical; an oblique avicularium is buried in the peristomie; the apertura (visible only from the interior) is semilunar; the frontal is surrounded by arcolar, parietal pores and on the zooccia it bears a large pore which does not perforate the wall.

Measurements.—Peristomice $\begin{cases} hpie=0.15 \text{ mm.} \\ lpie=0.12 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz=0.45-0.60 \text{ mm.} \\ lz=0.25-0.30 \text{ mm.} \end{cases}$

Affinities.—This species has the exterior aspect of an Adeonellopsis. The large frontal pore, however, is not a perforated area; it is invisible in the interior and its nature is unknown. The few specimens which we have studied were, it is true, rather poorly preserved. The peristomial avicularium is rarely visible at the exterior; it is on the contrary quite constant in the interior. This interior is that of Bracebridgia; it is therefore, indeed, in this genus that it must be classified, but it is a deformed Bracebridgia. The presence of a large frontal pore clearly characterizes this species and clearly differentiates it from the other known species of the same genus.

Occurrence.—Lower Miocene (Bowden marl): Bowden, Jamaica (rare). Cotypes.—Cat. No. 68674, U.S.N.M.

Genus LAMINOPORA Michelin, 1842.

1842. Laminopora Michelin, Iconographie zoophytoolgique.

The apertura is very elongate; the anter is semielliptical; the poster bears a long, rounded rimule, two small cardelles separate the two latter parts; the operculum bears laterally two lateral denticles articulating on the cardelles. The frontal is a tremocyst with pores not starred. The gonoecia have no special form. Thirteen to fifteen tentacles.

Genotype.—Laminopora contorta Michelin, 1842.

Range.—Miocene-Recent.

Gemellipora arbuscula Calvet, 1907, belongs to this genus.

LAMINOPORA MIOCENICA, new species.

Plate 47, fig. 5.

Description.—The zoarium is bilamellar. The zooccia are distinct, separated by a deep furrow, elongated, oval, much narrowed behind, surrounded by a row of parietal dietellae; the frontal is convex, perforated by large pores; it bears a small elongated avicularium variable in form and in position. The apertura is long and narrow; the rimule is long and spear shaped.

 $\label{eq:measurements} \textit{Measurements.} - \text{Aperture} \begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.06 \text{ mm.} \end{cases} \quad \text{Zooccia} \begin{cases} Lz = 0.65 \text{ mm.} \\ lz = 0.40 - 0.45 \text{ mm.} \end{cases}$

Affinities.—The present new species differs from Laminopora contorta Michelin, 1842, in the position of its avicularium below the aperture. It differs from Laminopora arbuscula Calvet, 1907, in the greater length of its apertura (0.15 and not 0.11 mm.), in its bilamellar zoarium and in its apertura not occupying two-fifths of the zooccial length.

The avicularium is very irregular, sometimes straight with the beak directed

above, sometimes transverse, triangular, or elliptical.

Occurrence.—Miocene (Bowden horizon): Santo Domingo (very rare). Holotype.—Cat. No. 68675, U.S.N.M.

Fig. 28.—Genus Laminopora Michelin, 1842.

A-F. Laminopora contorta Michelin, 1842. A. Sketch of aperture, × 85, showing elongated sinus. B. Zooecia, × 25. C. Mandible, × 85. E. Lateral view of a zoarium showing large vicarious vibracula. F. Zoarium, natural size, illustrating branching at right angles after forming very short branches only. (A-F, after Waters, 1919.)

G-J. Laminopora arbuscula Calvet, 1902. G. Zooecia, × 45. H. Operculum, × 90, viewed on the outer side. I. Operculum, × 90, viewed on the inner side. J. Zoarium, natural size. (G-J, after Calvet, 1907.)

Genus ADEONELLOPSIS MacGillivray, 1886.

(For description, see Bulletin 106, U. S. National Museum, p. 563.)

ADEONELLOPSIS COCCINELLA, new species.

Plate 24, figs. 5-8.

Description.—The zoarium is bilamellar with small irregular fronds. The zooecia are distinct, little elongate, elliptical, little convex; the frontal is perforated by four to seven stellate pores. The apertura is terminal, semilunar, with a slightly

eoneave proximal border; the peristome is thin, little salient, notched, and bears a small very inconstant supraoral avicularium. On each side of the apertura there are two small straight avicularia, without pivot, the point above.

Measurements.—Apertura $\begin{cases} ha = 0.06 \text{ mm.} \\ la = 0.04 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz = 0.40 \text{ mm.} \\ lz = 0.20 \text{ mm.} \end{cases}$

Affinities.—This species is quite close to Escharipora stellata Smitt, 1872, a recent species off the coast of Florida, in the ensemble of its characters. It differs from it in the vertical direction of the oral avicularia, in the less low position of the latter, and in the much smaller number of frontal perforations. Escharipora stellata MacGillivray, a recent species from Australia, is not the species of Smitt, 1872, but it is undoubtedly of the same genus.

We are absolutely ignorant of the hydrostatic system of this group called *Escharipora* by Smitt, 1872. MaeGillivray believed that it belonged to the Adeonidae. Levinsen considered the stellate pores as ascopores. It has seemed to us that one of our specimens had a gonoecium with transverse apertura, but it is necessary to await more detailed zoological study in order to classify this group of species.

Occurrence.—Lower Miocene (Chipola marl): Chipola River, Calhoun County, Florida (rare).

Cotypes.—Cat. No. 68676, U.S.N.M.

Genus ANARTHROPORA Smitt, 1867.

(For description, see Bulletin 106, U. S. National Museum, p. 430.)

Levinsen classified this genus doubtfully in the Escharellidae, where we also placed it in our monograph on the North American Early Tertiary Bryozoa. The absence of the ovicell, the nature of the frontal, and the analogy with certain species of Adeonellopsis indicate that the genus is better placed in the Adeonidae.

Family HIPPOPODINIDAE Levinsen, 1909.

Genus METRARABDOTOS Canu, 1914.

(For description, see Bulletin 106, U. S. National Museum, p. 533.)

METRARABDOTOS COLLIGATUM Canu and Bassler, 1919.

Plate 4, figs. 3-12.

1919. Metrarabdotos colligatum Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 95, pl. 4, figs. 3-12.

Description.—The zoarium is free, bilamellar, attached to algae by a small, expanded base and bent upward like a erank; the fronds are large, bifurcated, but narrow. The zooecia are distinct, separated by a salient thread, long and narrow; the frontal is smooth, convex, surrounded by a line of large areolar pores often separated by short costules. The apertura is suborbicular; the peristomice is oval with a proximal pseudorimule. There are sometimes two quite inconstant, small,

oral avicularia. The ovicell is endozooecial, enormous, borne by a considerably enlarged zooecium; its frontal is garnished with radiating and granulated costules.

Measurements.—Young zooecia: Apertura $\begin{cases} ha = 0.14 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases}$ Zooecia: $\begin{cases} Lz = 0.60 \text{ mm.} \\ lz = 0.28 - 0.30 \text{ mm.} \end{cases}$ Old zooecia: Apertura $\begin{cases} ha = 0.24 \text{ mm.} \\ la = 0.16 \text{ mm.} \end{cases}$ Zooecia: $\begin{cases} Lz = 0.70 \text{ mm.} \\ lz = 0.24 - 0.30 \text{ mm.} \end{cases}$

Affinities.—The genus Metrarabdotos contains more species in America than in Europe. The present species differs from Metrarabdotos moniliferum Milne-Edwards,

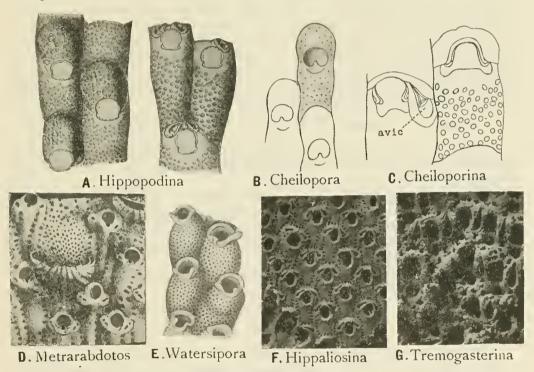


Fig. 29.—Genera of the Hippopodinidae Levinsen, 1909.

A. Hippopodina Levinson, 1909; H. feegeensis Busk, 1884, \times 35. B. Cheilopora Levinson, 1909; C. sincera Smitt, 1867, \times 20, C. Cheiloporina, new genus; C. haddoni Harmer, 1902, \times 44. D. Metrarabdotos Canu, 1914; M. moniliferum Milne-Edwards, \times 25. E. Watersipora Neviani, 1895; W. cucullata Busk, 1853, \times 35. F. Hippaliosina Canu; H. rostrigera Smitt, 1872, \times 20. G. Tremogasterina Canu 1911; T. horrida, new species, \times 20.

1836, common in the French Miocene, in its granulated costules on the ovicell, in the absence of a pseudolyrule at the peristomice, and in the absence of oral avicularia. It differs from *Metrarabdotos grandis* of the Vicksburgian, which also forms large fronds, in its greater zooecial width and in much less deep intercostular spaces

of the ovicell. The species is quite variable and we have figured the principal variations.

Occurrence.—Lower Miocene (Bowden horizon): Cercado de Mao and Rio Cana Santo Domingo (common).

Cotype.—Cat. No. 68677, U.S.N.M.

METRARARDOTOS LACRYMOSUM Canu and Rassler, 1919.

Plate 8, figs. 1-10.

1919. Metrarabdotos lacrymosum Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 96, pl. 3, figs. 1-10.

Description.—The zoarium is free, bilamellar, bifurcated, narrow, with rectangular fronds, and fastened by a little expanded base. The zooecia are distinct, little elongate, rectangular, surrounded by large arcolar pores more or less separated by the costules. The apertura is oblique, semielliptical, divided in front by a rounded and perforated sinus. The ovicelled zooecia are quite wide; the ovicell is endozooecial; it is little salient, finely punctate, and ornamented with interareolar costules. On each side of the apertura there are two large, triangular, salient avicularia with their point directed upward.

Measurements.—Apertura $\begin{cases} ha = 0.18 \text{ mm.} \\ la = 0.14 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz = 0.70 - 0.74 \text{ mm.} \\ lz = 0.28 - 0.30 \text{ mm.} \end{cases}$ Affinities.—The characteristics of this species are the two large, oral avicularia

Affinities.—The characteristics of this species are the two large, oral avicularia which hang from each side of the apertura like two large tears. They are very irregular.

This species differs from *Metrarabdotos colligatum* from the same geological horizon, in its little expanded and nonprehensile base, in its nonclaviform fronds, and in the presence of large, oral avicularia. It differs from *Metrarabdotos moniliferum* Milne-Edwards, 1838, in its much less costulated ovicell and in the absence of a spine in the oral sinus.

Occurrence.—Lower Miocene (Bowden marl): Bowden, Jamaica (common). Cotypes.—Cat. No. 68678, U.S.N.M.

METRARABDOTOS AURICULATUM, new species.

Plate 31, figs. 1-9.

Description.—The zoarium is bilamellar and formed of large, long, narrow, bifurcated fronds. The zooecia are distinct, separated by a salient thread, claviform; the frontal is narrow, little concave, surrounded by numerous areolar pores, separated by salient costules. The peristomice bears a rather deep proximal sinus at the base of which there are two lateral condyles limiting the orifice of the compensatrix; the apertura, buried at the base of the peristomice, is elliptical, a little elongated; the peristome bears two lateral avicularia. The ovicell is endozooecial, quite large; its superior wall is convex and costulate; its apertura is large and transverse.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.18 \text{ mm.} \\ la = 0.16 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.70 - 0.90 \text{ mm.} \\ lz = 0.40 \text{ mm.} \end{cases}$

Affinities.—This species differs from Metrarabdotos colligatum and from Metrarabdotos lacrymosum in its much larger zoaria and in its larger zooecial dimensions (Lz>0.70 mm.). It differs from Metrarabdotos moniliferum Milne-Edwards, 1838, in the absence of a pseudolyrule in the peristomice.

The two large oral avicularia occur like two lateral ears; they are rather constant but often are diminished and partially buried by the activity of the calcification. In some rare cases they are replaced by a single supraoral avicularium.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (common). Pliocene (Caloosahatchee marl): Shell Creek, De Soto County (very common), and Monroe County, Florida (very common).

Cotypes.—Cat. Nos. 68679, 68680, U.S.N.M.

Genus HIPPALIOSINA Canu, 1918.

1918. Hippaliosina Canu, Hippaliosina, un nouveau genre de Bryozoaires, Bulletin de la Société géologique de France, ser. 4, vol. 18, p. 88.

The ovicell is endozooecial. The apertura is elongate, elliptical, divided into two parts by two triangular cardelles; the poster is smaller than the anter. The frontal is a granular pleurocyst surmounting an olocyst perforated laterally by areolar pores. There are usually two avicularia on each side of the apertura.

Genotype.—Hippaliosina (Escharella) rostrigera Smitt, 1872.

Range.—Rupelian-Recent.

The family Hippopodinidae is perhaps not a natural one, because the larva is not known. We classify here all the species in which the ovicell is endozooecial. Nevertheless, the genus Hippaliosina is undoubtedly very close to Hippopodina Levinsen, 1909, and Cheilopora Levinsen, 1909, which are characteristic of the family. It differs solely in the function of calcification; the tremocyst is replaced by a granular pleurocyst, accompanied by lateral areolar pores. Metrarabdotos Canu, 1914, presents also lateral areolar pores and a pleurocyst, but the hydrostatic function operates here through a vanna and not a rimule; moreover, the ovicells are totally different.

In addition to two new recent species from the Philippine Islands, the following

species belong to this genus:

Hippaliosina (Lepralia) depressa Busk, 1852. Recent. Hippaliosina (Escharella) rostrigera Smitt, 1872. Recent. Hippaliosina (Lepralia) clavula Manzoni, 1871. Helvetian.

Hippaliosina brevirostris Canu, 1918. Recent.

Hippaliosina (Hemeschara) sandbergeri Reuss, 1869. Rupelian, Aquitanian.

Hippaliosina laxipora Canu, 1918. Miocene.

The genotype Hippaliosina rostrigera has been dredged near the twenty-second parallel a slight distance from the Tropic of Cancer, where it appears to have found its best conditions for existence. We can infer that the fossil species have lived under similar conditions and that their presence indicates likewise the vicinity of the Tropic. In this case their geologic distribution would indicate the displacement of this line through the ages, and consequently the contraction of the equatorial

zone. Employing the known localities we have the following table for the Northern Hemisphere:

Species.	Geological distribution.	Latitude.
H. clavula	Helvetian of Italy	45°
	Pliocene of Rhodes	36°
H. sandbergeri	Aquitanian of Gironde	44°
	Upper Miocene of Virginia	
V	Upper Miocene of North Carolina	
	Lower Pliocene of South Carolina	
	Upper Pliocene of Florida	25°
	Recent of Florida.	

The interpretation of this table indicates in a striking fashion the rapid and continuous descent of the Tropic toward the Equator. No species of the genus exists in the faluns of Touraine (forty-ninth parallel) situated without the equitorial zone. On the contrary, the Helvetian of Italy represented by H. clavula was nearer

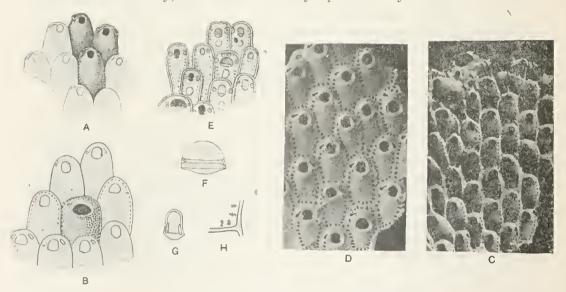


Fig. 30.—Genus Hippaliosina Canu, 1918.

A-C. Hippaliosina rostrigera Smitt, 1872. A. Ordinary zooecia. B. Group with an ovicelled zooecium. (A, B, after Smitt, 1872.) C. Interior of a fossil example. \times 20.

D. $Hippaliosina\ sandbergeri\ Reuss,\ 1869.$ An ovicelled specimen, $\times\ 25$, from the Aquitanian of Leognan, France.

E-H. Hippaliosina brevirostris Canu, 1918. E. Ovicelled specimen, \times 25. F. Operculum of the ovicelled zooccia, \times 85. G. Operculum of ordinary zooccia, \times 85. H. Areolar pores. \times 85. (E-H, after Waters, 1889.)

to it. This latter species does not exist in the more southern European Pliocene. *H. sandbergeri*, not being able to emigrate toward the south probably by the obstacle of a continental barrier, was exterminated in its place. It is probable that it will be discovered in more northern Oligocene deposits.

For the Southern Hemisphere this phenomenon is less evident because of the imperfect geographic conditions. Although the fossil species H. laxipora has been

observed along the thirty-fifth and thirty-sixth parallels, the recent species has been dredged only nearer the Equator from the thirty-fifth to the tenth parallels. The emigration northward toward the Equator appears thus to be a phenomenon as real as in the other hemisphere.

The bryozoa, on account of their method of life and their abundance in the marine currents, are generally rather cosmopolitan. It is remarkable that the genus *Hippaliosina* has been so sensitive to external variations, and this is a new phenomenon which merits recognition.

HIPPALIOSINA ROSTRIGERA Smitt, 1872.

Plate 17, figs. 15-17.

1872. Escharella rostrigera Smrt, Floridan Bryozoa, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol 11, p. 57, pl. 10, figs. 203–205.

1918. Hippaliosina rostrigera Canu, Hippaliosina, un nouveau genre de Bryozoaires, Bulletin de la Société géologique de France, ser. 4, vol. 18, p. 89.

Measurements (fossil specimens):

Apertura of ordi-[ha = 0.15 mm.]nary zooecia _ [la = 0.16 mm.]Cordinary zooecia _ [la = 0.30 mm.]Ordinary zooecia _ [la = 0.30 mm.]Ovicelled zooecia _ [la = 0.30 mm.] [la = 0.16 mm.]Ovicelled zooecia _ [la = 0.38 mm.]

The ovicelled zooecia have an aperture larger and transverse and, moreover, of irregular form. Smitt has noted and has figured an elliptical aperture measuring 0.36 mm. in width. This variation must be rather rare, for we have only observed it on the fossil specimens from the Miocene near Yorktown, Virginia. In the Australian species the small ovicelled zooecia appear to be absent. The ovicelled zooecia should not be confounded with gonoecia. The latter do not contain a polypide, as the ovary alone occupies the interior cavity. The ovicell is little salient, little visible, but always apparent on our fossil specimens. The zooecia which bear them are larger.

This species has been dredged south of Florida in the vicinity of the Tropic of Cancer. The special location gives a clue to the contraction of the equatorial zone

since the Oligocene period.

Geologic distribution.—Miocene (Yorktown formation): Yorktown, Suffolk, and 3 miles southwest of Petersburg, Virginia (common). Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina (common). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare). Pliocene (Caloosahatchee marl): Monroe County, Florida (rare).

Habitat.—South of Florida at a depth of 56 to 59 meters.

Plesiotypes.—Cat. No. 68681-68683, U.S.N.M.

Genus TREMOGASTERINA Canu, 1911.

1911. Tremogasterina Canu, Iconographie des Bryozoaires fossiles de l'Argentine, pt. 2, Anales del Museo Nacional de Buenos Aires, vol. 21 (sec. 3, vol. 14), p. 256.

The ovicell is concealed. The frontal is perforated by a large pore and surrounded by a line of areolar pores. The apertura is semiclliptical. Avicularia between the apertures.

Genotype.—Tremogasterina problematica Canu, 1911.

Range.—Rocanean, Miocene.

In 1911 Canu discovered this remarkable and mysterious genus in the Rocanean strata of Argentina; that is to say, at the base of the Eocene. He noted the nature of the ovicell in his remarks that it opened in the interior of the zooecium. The single described species, Tremogasterina problematica, is insufficiently preserved to serve as a genotype, and as the Miocene species here described are much better preserved, they will give a clearer idea of the genus. The form of the aperture indicates clearly the presence of a compensatrix. Therefore, it is rather probable that the enormous frontal pore is an ascopore.

TREMOGASTERINA HORRIDA, new species. 6

Plate 24, figs. 3, 4.

Description.—The zoarium is free, bilamellar, with small distorted fronds. The zooccia are distinct, very elongate, elliptical, little regular, little convex; the frontal is perforated by a large, median orbicular pore, surrounded by a line of scattered areolar pores. The apertura is very large, elongate, semielliptical; it presents a slight constriction at the lower third, probably at the level of the articulations. The peristome is thick and tuberous. The ovicell is endozooccial, smooth, transverse, little salient; the apertura is of the same size as that of the ordinary zooccia. Between two apertures, on one or the other zooccium, there is an immense triangular avicularium, without pivot with the beak turned toward the top.

 $Measurements. - Apertura \begin{cases} ha = 0.18 - 0.20 \text{ mm.} \\ la = 0.14 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.60 \text{ nm.} \\ lz = 0.30 \text{ mm.} \end{cases}$ Opesium of the $\begin{cases} hav = 0.14 \text{ mm.} \\ lav = 0.10 \text{ mm.} \end{cases}$ Avicularium $\begin{cases} Lav = 0.30 - 0.40 \text{ mm.} \\ lav = 0.24 \text{ mm.} \end{cases}$

Affinities.—This species has a disagreeable aspect and its structure is difficult to understand. We have only found three small specimens and we have not been able to make a good dissection. The ovicell itself was not in condition to be photographed. If the frontal perforation is an ascopore, the apertura ought not to have an operculum moving on a special axis; this is not the case, the proximal border not being straight. This perforation corresponds perhaps to a special axicularium.

Occurrence.—Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (rare).

Holotype.—Cat. No. 68684, U.S.N.M.

CHEILOPORINA, new genus.

The ovicell is endozooccial; the apertura of the ovicelled zooccium is very large. The frontal is a tremocyst. The apertura bears two cardelles and the operculum is narrowed laterally at their level. No separation between the ovicell and the zooccium. Twenty-seven tentacles.

Genotype.—Cheiloporina (Hippoporina) circumcincta Neviani, 1896. Range.—Jacksonian to Recent.

A second new species Tremogasterina truncatorostris is given on p. 244.

Historical.—In our monograph on the Early Tertiary Bryozoa we noted: "The genotype is deprived of cardelles. We think that the species having cardelles and a different operculum must be introduced into a new genus." Our manuscript was written in 1917. In 1918 Waters ⁷ stated that the operculum of Cheilopora circumcincta Neviani, 1896, is quite different from the operculum of the genotype Cheilopora sincera Smitt, 1867, and that these two species can not be maintained in the same genus. Under these conditions we are justified in creating the new genus Cheiloporina.

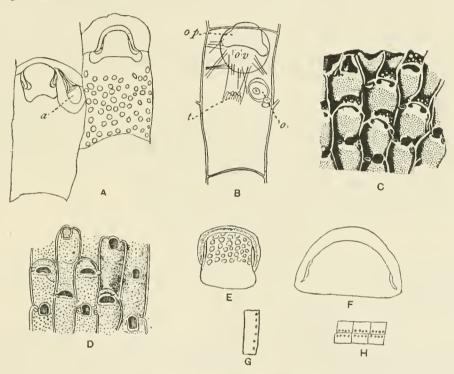


Fig. 31.—Cheiloporina, new genus.

A-H. Chciloporina (Chcilopora) haddoni Harmer, 1902. A. Two zooecia, one ovicelled, showing an avicularium (a) and the two kinds of opercula. B. Basal view of zooecium with trifoliate operculum. (After Harmer, 1902), op. operculum; t, tentacles; o, ovary; ov, ovisac. C. Zoarium with ovicelled zooecia, \times 25. (After Jullien, 1903.) D. Ovicelled zooecia, \times 12. E. Operculum of ordinary zooecium, \times 85. F. Operculum of ovicelled zooecia, \times 85. G. Lateral wall showing septulae. H. Distal wall with septulae. (D-H, after Waters, 1918.)

All the species of Cheilopora described in the Monograph of North American Early Tertiary Bryozoa belong in reality to Cheiloporina. These are as follows: Cheilopora labiosa Ulrich, 1901; Cheilopora orbifera; C. prelucidioides; C. strictocella; C. grandis; C. transversa; C. transversides; C. specula; and C. sulcifera Canu and Bassler.

⁷ On some Mediterranean Bryozoa, Annals and Magazine Natural History, ser. 9, vol. 11, p. 97, pl. 12, figs. 6-10.
12184—23—Bull, 125——12

Family TUBUCELLARIIDAE Busk, 1884.

Genus TUBUCELLARIA D'Orbigny, 1852.

(For description, see Bulletin 106, U. S. National Museum, p. 542.)

TUBUCELLARIA PUNCTULATA Gabb and Horn, 1862.

Plate 40, figs. 1-4.

1862. Entalophora punctulata Gabb and Horn, Monograph Polyzoa Secondary and Tertiary formations of North America, Journal Academy Natural Sciences, Philadelphia, ser. 2, vol. 5, p. 171, pl. 21, fig. 61.

 $\label{eq:Measurements} \textit{Measurements}. \\ - \{ \begin{aligned} \text{Peristome, 0-30 mm.} \\ \text{Peristomice, 0.18-0.20 mm.} \end{aligned} \quad \text{Zooecia} \{ \begin{aligned} Lz &= 1.20-1.30 \text{ mm.} \\ lz &= 0.40-0.50 \text{ mm.} \end{aligned}$

Affinities.—This species is not entirely articulated; the segments are often ramified. The peristome sometimes bears one or two small avicularia. The frontal is a tremocyst with large pores.

The exterior aspect is rather deceiving and the older American authors classified the species in *Entalophora*. The bifurcation of the segments confirmed this reference but the longitudinal section proves their error (fig. 4). The ovicell is buried in the thickness of the peristomial walls; unfortunately we have not yet had the chance to section an ovicelled zoarium.

Occurrence.—Pleistocene: Santa Barbara (very common), Santa Monica (rare), and Dead Mans Island, off San Pedro, California (rare).

Plesiotypes.—Cat. Nos. 68686-68688, U.S.N.M.

TUBUCELLARIA PUNCTULATA, var. MINOR, new variety.

Plate 40, figs. 5, 6.

The micrometric measurements are smaller; the tremopores are smaller and more scattered.

 $\label{eq:lemmatic} \begin{tabular}{ll} \textit{Measurements}. & & & & & & & & \\ \textit{Peristomice}, \ 0.24 \ \text{mm}. & & & & & & \\ \textit{Peristomice}, \ 0.14 \ \text{mm}. & & & & & & \\ \textit{Zooecia} & & & & & \\ \textit{lz} = 1.20 \ \text{mm}. & & & \\ \textit{lz} = 0.40 \ \text{mm}. & & & \\ \end{tabular}$

The constancy of these small micrometric measurements obliges us to separate this variety from the well-known long-ago described species.

Occurrence.—Pleistocene: Santa Barbara (rare) and Santa Monica (Tremochal Canon), California (rare).

Holotype.—Cat. No. 68689, U.S.N.M.

Family PHYLACTELLIDAE Canu and Bassler, 1917.

Genus PHYLACTELLA Hincks, 1880.

(For description see Bulletin 106, U.S. National Museum, p. 573.)

PHYLACTELLA SPINOSISSIMA, var. MAJOR Hincks, 1884.

Plate 39, figs. 8, 9.

1884. Mucronella spinosissima, var. major Hincks, Polyzoa of the Queen Charlotte Islands, Annals and Magazine Natural History, ser. 5, vol. 13, p. 53 (sep. 27); p. 213 (sep. 42).

Affinities.—In Miss Jelly's Synonymic Catalogue of Marine Bryozoa this species is considered a synonym of Mucronella peachi, var. octodentata Hincks, 1880. We believe this arrangement is erroneous because here the ovicell is recumbent and placed

on the collar of the zooecium (necklike of Hincks), which is characteristic of the Phylactellidae. Neither do we believe that it is identical with Mucronella spinosissima Hincks, 1881, from Australia, and Hincks himself has noted some important differences. As we have not the material at hand for comparison we are adopting Hincks name of 1884, as our fossil is identical with the variety figured by him from the Queen Charlotte Islands.

The trace of numerous spines on the peristomice is quite visible on our fossils. We have not observed the tubular system noted by Hincks, but on many of the well-preserved zooccia we have observed the transformation of the tremopores into divergent tubules rather long and very little salient. There are eight spines on the peristome of the ovicelled zooccia.

This species differs from *Phylactella collaris* Norman, 1866, figured by Miss Robertson in 1908, in its ovicell of less width than the zooccium and smaller in its ensemble.

Occurrence.—Pleistocene: Santa Monica (Long Wharf Canyon), California (rare).

Habitat.—Pacific: Queen Charlotte Islands. Plesiotype.—Cat. No. 68690, U.S.N.M.

Genus LAGENIPORA Hincks, 1877.

(For description see Bulletin 106, U. S. National Museum, p. 591.)

LAGENIPORA SPINULOSA Hincks, 1884.

Plate 40, fig. 7.

1884. Lagenipora spinulosa Ilineks, Polyzoa of the Queen Charlotte Islands, Annals and Magazine of Natural History ser. 5, vol. 13, p. 57 (sep. 31), pl. 3, fig. 4; p. 210 (sep. 40), pl. 9, fig. 4.

1908. Lagenipora spinulosa Robertson, The incrusting Cheilostomatons Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 4, No. 5, p. 283, pl. 18, fig. 37.

Our figured specimen, much changed by fossilization, gives only a very poor idea of the beauty of this fragile species. The other specimens observed are equally poorly preserved, but there is no occasion to doubt their identity with this recent species.

Occurrence.—Pleistocene: Dead Mans Island, off San Pedro, California (rare). Habitat.—Pacific: Off California.

Plesiotype.—Cat. No. 68691, U.S.N.M.

LAGENIPORA(?) BREVICOLLIS, new species.

Plate 24, fig. 9.

Description.—The zoarium incrust Cellepores. The zooccia are distinct, separated by a deep furrow, elongated, oriented, elliptical; the frontal is very convex and covered with tremopores. The peristome is incomplete, interrupted distally, little salient, much enlarged in its proximal portion. The apertura hidden at the bottom of the peristome bears cardelles placed very low, and an almost straight proximal border.

Measurements.—Apertura $\begin{cases} ha = 0.10 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{cases}$ Zooecia $\begin{cases} Lz = 0.50 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Variations.—There are sometimes on the peristome two very small avicularia symmetrically placed. We have not observed the ovicell, but the distal interruption of the peristome is a sufficient reason for our generic assignment.

This species differs from Lepralia tenera Reuss, 1867, in the presence of frontal

tremopores.

Occurrence.—Miocene: Kuhns, Carteret County, North Carolina (rare). Holotype.—Cat. No. 68692, U.S.N.M.

Genus MASTIGOPHORA Hincks, 1880.

(For description see Bulletin 106, U. S. National Museum, p. 586.)

MASTIGOPHORA PESANSERIS Smitt, 1873.

Plate 45, fig. 10.

- 1873. Hippothoa pesanseris Smirt, Floridan Bryozoa, pt. 2, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 11, No. 4, p. 43, pl. 7, figs. 159, 160.
- 1880. Mastigophora dutertrei, var. pesanseris Kirkpatrick, Polyzoa of Mauritius, Annals Magazine Natural History, ser. 6, vol. 1, p. 77.
- 1899. Schizoporella pesanseris Waters, Bryozoa from Maderia, Journal Royal Microscopical Society, p. 11, pl. 3, figs. 7, 8.
- 1905. Mastigophora dutertrei, var. pesanseris Thornelly, Report on Polyzoa collected by Professor Herman at Ceylon in 1892, Ceylon Pearl Oyster Fisheries, Rept. Colonial Government, vol. 4, Suppl. Rep. No. 26, p. 117.
- 1909. Schizoporella pesanseris Waters, Marine Biology of Sudanese Red Sea, Journal Linnean Society London, vol. 31, p. 169.
- 1909. Escharina pesanscris Norman, Polyzoa of Maderia and neighboring islands, Journal Linnean Society London, vol. 30, p. 302, pl. 40, fig. 7.
- 1909. Escharina pesanseris Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, p. 326, pl. 18, fig. 1.
- 1914. Escharina pesanseris Osburn, Bryozoa of the Tortugas Islands, Florida, Publication Carnegie Institution of Washngton, No. 182, p. 207.

The generic position of this species is still doubtful. According to the operculum, Waters classified it in the S. cecili group (Arthropoma); according to the form and position of the avicularia, Levinsen classified it in Mastigophora (Escharina). He believed that the ovicell is endozooccial. If this observation be verified it will be necessary to create a new genus for this species. Our specimen is incrusting a coral.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare).

Habitat.—Indian Ocean: Mauritius, Nanaar (55 meters), Siam, Gulf of Suez (Red Sea). Atlantic Ocean: Madeira (90 meters), Tortugas Islands (12-68 meters). Plesiotype.—Cat. No. 68693, U.S.N.M.

MASTIGOPHORA GRANULOSA, new species.

Plate 3, fig. 4.

Description.—The zoarium incrusts shells. The zooecia are distinct, separated by a deep furrow, large, elongated, and swollen; the frontal is very convex, ornamented with numerous small pores and minute granulations. The aperture is small,

somewhat elongated, and bears a wide rimule which is rounded and deep; the peristome is wide, very little salient, and bears spines. The two vibracula are salient and orbicular, the ovicell is small, convex, smooth.

Measurements.—Apertura $\begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases}$ Zooccia $\begin{cases} Lz = 0.60 - 0.75 \text{ mm.} \\ lz = 0.45 - 0.55 \text{ mm.} \end{cases}$

Affinities.—This species differs from Mastigophora pesanseris Smitt, 1872, in its larger micrometric measurements, in its frontal granules larger and more apparent, and in its vibracula placed at the level of the rimule.

Occurrence.—Miocene (Bowden marl): Bowden, Jamaica (very rare).

Holotype.—Cat. No. 68694, U.S.N.M.

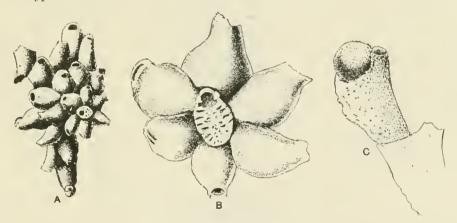


Fig. 32.—Genus Temachia Jullien, 1882.

A-C. Temachia opulenta Jullien, 1882. A. An incrusting specimen, \times 8.5, from the Gulf of Gascony. B. Ancestrula and ancestrular zooecia, \times 29. C. An ovicelled zooecium, \times 29.

Genus TEMACHIA Jullien, 1882.

1882. Temachia Jullien, Dragages du Travailleur, Bulletin de la Societe de France, vol. 7, p. 510.

Zooecia subcrect, dilated at the base and narrowed like the neck of a bottle above; peristome split anteriorly and deprived of spines; ovicell globular with the opening corresponding to the incision in the peristome. Zooecia of origin (ancestrula) with a frontal wall entire and lattice like, with two strong lateral spines at the level of the orifice. (Translation, after Jullien.)

Genotype. - Temachia opulenta Jullien, 1882. Recent.

Genus CREPIDACANTHA Levinsen, 1909.

1909. Crepidacantha Levinsen, Morphologic and Systematic Studies on the Cheilostomatous Bryozoa, p. 266.

The zooccia, whose aperture has strong hinge teeth and a compound, well-chitinized operculum, are in the proximal half provided with 9 to 12 very long marginal spines and 8 to 11 small uniporous pore chambers (dietellae) alternating as a rule with small intermediate chambers, each of which has an uncalcified spot

(a marginal pore) in its roof. Two frontal vibracula without a crossbar. The hyperstomial, almost free ooecia, consist of two calcified layers, of which the ectooecium is provided with a number of pores. (Levinsen.)

Genotype.—Crepidacantha poissoni Savigny-Audouin, 1826. Recent.

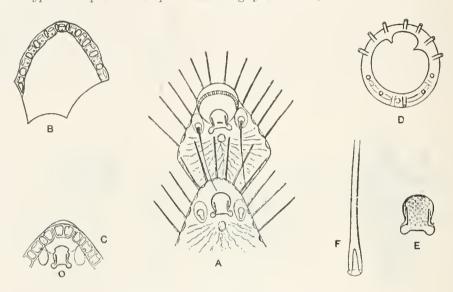


Fig. 33.—Genus Crepidacantha Levinsen, 1909.

A-F. Crepidacantha poissoni, var. crinispina Levinsen, 1909. A. Two zooecia, \times 55. B. A zooecium seen from the basal wall. As in figure C. dietellae alternate with intermediate spaces, \times 55. C. The distal part of a zooecium with ovicell, seen from the basal wall, \times 55. D. An ancestrula of another variety of the same species, \times 175. E. Operculum, \times 140. F. The proximal part of the flagellum, \times 200. (After Levinsen, 1909.)

Family CELLEPORIDAE Busk, 1852.

Genus HOLOPORELLA Waters, 1909.

For description, see Bulletin 106, U. S. National Museum, p. 604.

HOLOPORELLA ALBIROSTRIS Smitt, 1872.

Plate 7, figs. 9-14; plate 32, figs. 6-10.

1872. Discopora albirostris Smitt, Floridan Bryozoa, pt. 2, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 11, no. 4, p. 70, pl. 12, figs. 233–239.

1889. Cellepora albirostris Jelly, A synonymic catalogue of the Recent Miocene Bryozoa, p. 45 (Bibliography).

1914. Holoporella albirostris Osburn, Bryozoa of the Tortugas Islands, Publication 182, Carnegie Institution of Washington, p. 215.

1919. Holoporella albirostris Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa. Publications of the Carnegie Institution of Washington, no. 291, p. 97, pt. 1, fig. 19, pl. 7, figs. 9-14.

Measurements.—Apertura, ha = 0.14 mm.

Zooecia
$$\begin{cases} Lz = 0.40 - 0.60 \text{ mm.} \\ la = 0.30 \text{ mm.} \end{cases}$$
 Avicularium $\begin{cases} La = 0.60 \text{ mm.} \\ la = 0.40 \text{ mm.} \end{cases}$

Variations.—The apertura is semilunar with a concave proximal border; it is indeed only constant in the interior (pl. 32, fig. 10). Exteriorly the peristomice is

quite variable; it presents very frequently an excentric pseudosinus, as in the specimens dredged by the *Challenger* at Heard Island. This is occasioned by the very irregular development of a spiniform mucro. The latter is very fragile and never exists on our fossils; it is scarcely visible, and then only on the deep zooecia. The frontal is smooth and garnished laterally with very small areolar pores (pl. 32, fig. 8) which are rather easily obliterated (pl. 32, fig. 7). The interzooecial avicularium is spatulate; its distribution on the zoarium is very inconsistent.

The zoarium is always attached to algae; it creeps above or indeed completely around them to form masses of 3 centimeters in diameter. The depth at which it is dredged therefore gives no indication of its hydrostatic capacities if it is not living; the disappearance of the algae buries the zoarium in the depths where it did not live. The ancestrular zooecia are oriented (pl. 32, fig. 10) and the ancestrula emits five buds. The tuberosities observed on certain zoaria come from the greater development of certain zooecial groups.

Occurrence.—Oligocene (Emperador limestone): One-third mile north of Empire, Panama Canal Zone (rare). Oligocene (Anguilla formation): Southwest side of Crocus Bay, Anguilla, Leeward Islands (rare). Lower Miocene (Bowden marl): Bowden, Jamaica (very common). Pliocene (Calooshatchee marl): Shell Creek, De Soto County, and Monroe County, Florida (very common).

Geological distribution.—Miocene of Australia; Pliocene of New Zealand.

Habitat.—Atlantic: Off Florida (24–56 meters). Indian Ocean: Heard Island (121 meters). Pacific: Off Australia (to 13 meters).

Plesiotypes.—Cat. Nos. 68695-68697, U.S.N.M.

HOLOPORELLA PARVULA, new species.

Plate 24, figs. 10-13.

Description.—The zoarium is free, subcylindrical, irregularly branched. The zooecia are very small, heaped upon each other, irregular, provided with a small umbo before the apertura and surrounded by some arcolar pores. The apertura is very small, semielliptical, transverse, surrounded by one to four small orbicular avicularia.

Measurements.—Apertura $\begin{bmatrix} ha = 0.08 \text{ mm.} \\ la = 0.10 \text{ mm.} \end{bmatrix}$

Affinities.—In its general aspect this species may easily be confounded with Holoporella maculata Ulrich and Bassler, 1904, and with Holoporella minuta, new species. It differs from both of these in its free and cylindrical zoaria (and not attached to shells), and in the presence of the small avicularia which surround the apertura.

Occurrence.—Miocene (Duplin marl): Cape Fear River, 28 miles northwest of Wilmington, North Carolina (rare).

Holotype.—Cat. No. 68698, U.S.N.M.

HOLOPORELLA ROSTRIFERA, new species.

Plate 24, fig. 14.

Description.—The zoarium incrusts shells and forms small spiny masses. The zooecia are indistinct, erect; the frontal is convex, small, surrounded by a double row of areolar pores and often with costules directed toward the frontal rostrum

placed in front of the apertura. The apertura is somewhat elliptical, elongated; the two small cardelles are placed in the lower third. Around the apertura there are four very salient rostra (0.30 to 0.40 mm.) terminated by a small, round avicularium without pivot. The ovicell is small, convex, not closed by the operculum and presents a very fragile orbicular frontal area.

Measurements.—Apertura
$$\begin{bmatrix} ha = 0.17 \text{ mm.} \\ la = 0.14 \text{ mm.} \end{bmatrix}$$

Affinities.—The present species differs from Holoporella subturrita in its rostra, which are free and not attached to the distal zooccinm; in its frontal, which is not covered with tremopores; and in the presence of cardelles.

Occurrence.—Miocene (Yorktown formation): York River, Virginia (rare). Holotype.—Cat. No. 68699, U.S.N.M.

HOLOPORELLA SUBTURRITA, new species.

Plate 25, fig. 9.

Description.—The zoarium incrusts shells, on which it forms small spinose masses. The zooecia are distinct, little erect, swollen; the frontal is convex and covered with tremopores. The apertura is elliptical, somewhat elongated, without cardelles. It is surrounded by four long rostra terminated by avicularia; the two distal rostra have their peduncles attached to the distal zooecium.

Measurements.—Apertura
$$\begin{cases} ha = 0.16 \text{ mm.} \\ la = 0.14 \text{ mm.} \end{cases}$$
 Zooecia $\begin{cases} Lz = 0.60 - 0.80 \text{ mm.} \\ lz = 0.36 - 0.40 \text{ mm.} \end{cases}$

Affinities.—With its long rostra this species much resembles Holoporella turrita, Smitt, 1872, from which it differs in the two distal rostra attached to the superior zooecium, in its tremopores, and in the absence of small nonpedunculate frontal avicularia.

Occurrence.—Miocene (Duplin marl): 28 miles northwest of Wilmington, North Carolina (rare).

Holotype.—Cat. No. 68700, U.S.N.M.

HOLOPORELLA HEMISPHERICA, new species.

Plate 3, figs. 9, 10.

Description.—The zoarium is free, small, hemispherical, about the size of a pea. The zooecia are large, distinct, heaped upon one another; the frontal is quite convex, smooth, surrounded by some large areolar pores. The apertura is semi-elliptical, transverse with a concave proximal border.

Measurements.—Apertura
$$\begin{bmatrix} ha = 0.17 \text{ mm.} \\ la = 0.15 \text{ mm.} \end{bmatrix}$$

Affinities.—As we have collected only a few specimens of this small species, it is difficult to study its variations or its affinities. It appears very well characterized by the simplicity of its structure and the large areolar pores which surround its broad frontal.

Occurrence.—Lower Miocene (Bowden marl): Bowden, Jamaica (rare). Cotypes.—Cat. No. 68701, U.S.N.M.

HOLOPORELLA MASSALIS Ulrich and Bassler, 1904.

Plate 25, fig. 7.

1904. Cellepora massalis Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 428, pl. 117, fig. 3, 4.

The original description by Ulrich and Bassler is as follows:

Zoarium massive, composed of many layers, often nodose, always rough. Zooecia erect, very irregularly arranged, four or five in 2.0 mm.; orifice circular with a thin raised peristome. Generally the peristome of each zooecium bears upon its inferior side a promineut rostrum containing a large avicularium pointing obliquely upward and outward. Surface of zooecia, excepting the peristome, coarsely punctate. Ovicells not observed.

$$\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.\ 13\text{--}0.\ 15\ \text{mm.} \\ la = 0.\ 17\ \text{mm.} \end{cases}$$

We have little to add to the description of Ulrich and Bassler and are only introducing a new photograph of the type specimen made on the same enlargement as the other species of this volume so as to better allow comparison. In the type we have observed a case of total regeneration which is very rare in the Cellepore bryozoa. This species differs from *Holoporella orbifera* in the very constant presence of a large frontal avicularium and in its smaller apertura.

Occurrence.—Miocene (St. Mary's formation): St. Mary's River, Maryland (rare). Miocene (Choptank formation): Greensboro, Maryland (rare). Miocene (Calvert formation): Chesapeake Beach and Plum Point, Maryland (rare).

Holotype.—Cat. No. 68702, U.S.N.M.

HOLOPORELLA ORBIFERA, new species.

Plate 25, figs. 3-6.

Description.—The zoarium is free, large, very irregular; it forms most often lobate or branched masses 2 centimeters in length. The zooccia are large, little distinct, erect; the frontal is smooth and surrounded by areolar pores. The apertura is very large, orbicular, without cardelles, with a concave proximal border. The oral avicularia are rare and inconstant. The interzooccial avicularia are large elliptical, with neither pivot nor denticles.

$$\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{bmatrix} ha = 0.20 \text{ mm.} \\ la = 0.20 \text{--}0.25 \text{ mm.} \end{bmatrix}$$

Affinities.—This species is very easily recognized by its large oral dimensions. It can be compared only with *Holoporella magnifica* Osburn, 1914, now living in the water off Florida, from which it differs in the absence of the small interzooccial avicularia. Osburn unfortunately has not indicated the enlargement of his figure.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (rare) and Harvey's Mills, Leon County, Florida (rare). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (common).

Cotypes.—Cat. Nos. 68703, 68704, U.S.N.M.

HOLOPORELLA(?) ECHINATA, new species.

Plate 25, figs. 1, 2.

Description.—The zoarium is a globuliform mass, more or less gibbose. The zooeeia are large, salient, convex, smooth. The apertura is subcircular, the prox-

imal border being very concave. There are two oral spines and a small oral very inconstant avicularium. On the deep zooccia the apertura only is visible.

Measurements.—Apertura $\begin{cases} \hbar a = 0.15 \text{ mm.} \\ \ell a = 0.15 \text{ mm.} \end{cases}$

Affinities.—This species differs from Holoporella massalis Ulrich and Bassler, 1904, in the absence of areolar pores. In the presence of two oral spines it approaches Holoporella bicornis, but differs from it in its larger micrometric dimensions and in the absence of areolar pores. It is also quite close to Cellepora tuberosa Smitt, 1872, in the presence of a small oral avicularium and in the nature of the frontal: it differs from it in the presence of two spines and in a very different zoarial form.

Only the figured specimen has been found. More plentiful material will later

on permit a more exact study.

The absence of arcolar pores may cause our generic determination to be doubted; but they are easily obliterated on the fossils.

Occurrence.—Miocene (Yorktown formation): Yorktown, Virginia (very rare). Holotype.—Cat. No. 68705, U.S.N.M.

HOLOPORELLA BICORNIS, new species.

Plate 32, figs. 1-4.

Description.—The zoarium is formed of large shapeless masses attached to algae. The zooecia are erect, salient, very convex; the frontal is smooth, surrounded by some much-scattered arcolar pores. The apertura is semilunar with a concave proximal border; the peristome is wide and bears two large spines. A small, salient avicularium terminates the median umbo. The deep zooecia have only their apertura visible. The interzooecial avicularia are rare and elliptical. Sometimes there is a vibraculum on the zooecia.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{bmatrix} ha = 0.13 \text{ mm.} \\ la = 0.12 - 0.15 \text{ mm.} \end{bmatrix}$

Affinities.—Above many of the apertures a little pore in the form of a lunar crescent is visible; it has the aspect of vibraculum; it is placed on the distal zooecium or indeed it is interzooecial. Its function is unknown.

This species resembles Reptocelleporaria similis Tuomey and Holmes, but the figure of these authors indicates a greater number of areolar pores. It differs from Holoporella albirostris Smitt, 1872, in the presence of two large oral spines and in the massive form of the zoarium. In each zoarium there is always a perforation which is the trace of the alga to which the zoarium was attached. This, moreover, is the most frequent habitat of the Celleporidae.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County. South Carolina (rare).

Cotypes.—Cat. No. 68706, U.S.N.M.

HOLOPORELLA UMBONATA, new species.

Plate 40, figs. 8, 9.

Description.—The zoarium incrusts shells, bryozoa or algae. The zooecia are oriented, distinct, elongate, separated by a deep furrow; the frontal is convex, smooth, surrounded by pores and arcolar costules; a large umbo terminated by an

avicularium hides the apertura. The apertura is semilunar. The ovicell is globular, transverse, smooth; it is widely open in front of the *umbo* and never closed by the operculum.

Measurements.—Zooecia $\begin{bmatrix} Lz = 0.50 - 0.60 \text{ mm.} \\ lz = 0.25 - 0.40 \text{ mm.} \end{bmatrix}$

Affinities.—The interareolar costules are easily attenuated by fossilization and are visible only on the good specimens. The ancestrular zooecia are smaller and raised.

In its exterior aspect this species is close to *Cellepora pumicosa* Linnaeus, 1768; it differs from it in the presence of the interareolar costules and in a different form of the apertura.

Occurrence.—Pleistocene: Santa Barbara (rare) and Santa Monica (Temochal Canyon), California (very rare).

Cotypes.—Cat. No. 68707, U.S.N.M.

HOLOPORELLA TURRITA Smitt, 1873.

Plate 46, fig. 1.

1873. Lepralia turrita Smitt, Floridan Bryozoa, pt. 2, Kongl. Svenska Vetenskaps-Akademiens Handlingar, vol. 11, no. 4, p. 65.

1914. Holoporella turrita Osburn, The Bryozoa of the Tortugas Islands, Florida, Publication Carnegie Institution of Washington, no. 182, p. 217 (bibliography).

Our specimen incrusts a coral. The apertura bears two small cardelles, placed in the lower third.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (very rare).

Habitat.—Waters off Florida (14-71 meters).

Plesiotype.—Cat. No. 68708, U.S.N.M.

HOLOPORELLA AVICULIFERA, new species.

Plate 46, fig. 2.

Description.—The zoarium incrusts corals, forming very thick masses. The zooecia are somewhat erect, convex, and formed of a tremocyst with scattered, spaced pores; the frontal bears some interarcolar costules. The apertura is formed of a large anter and of a concave poster and arranged at the base of a short peristomic. There is a small avicularium in the peristomic. On the frontal some arcolar pores are transformed into small orbicular avicularia.

 $\label{eq:measurements} \textit{Measurements.} - \Delta \text{perturn} [ha = 0.18 \text{ mm.}] \\ la = 0.20 \text{ mm.}$

Affinities.—This species presents somewhat the aspect of Discopora pertusa Smitt, 1872. It differs from it in the presence of the small disseminated avicularia and in the absence of oral mucro.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (rare).

Holotype.—Cat. No. 68709, U.S.N.M.

HOLOPORELLA MUCRONATA, new species.

Plate 46, fig. 7.

Description.—The zoarium incrusts sponges. The zooccia are little erect, surrounded by a line of large, spaced, areolar pores; the little salient, areolar costules meet at an enormous, wide, oral, very salient mucro. Two very small cardelles separate the anter from the concave poster. Some areolar pores are transformed into small orbicular or triangular avicularia.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} [ha = 0.13 \text{ mm.} \\ la = 0.15 \text{ mm.} \\$

Affinities.—The general aspect recalls a little that of Holoporella janthina Smitt, 1872. It differs from it in the presence of areolar pores, much scattered, and in the presence of the large oral mucro. The oral mucro does not exist on the young zooecia.

Occurrence.—Pleistocene: Mount Hope, Panama Canal Zone (very rare).

Holotype.—Cat. No. 68710, U.S.N.M.

Genus SCHISMOPORA MacGillivray, 1888.

(For description, see Bulletin 106, U. S. National Museum, p. 598.)
SCHISMOPORA BREVINCISA, new species.

Plate 32, figs. 11-13.

Description.—The zoarium incrusts shells. The zooecia are erect and heaped at the center and oriented on the margin; the frontal is formed by a granular pleurocyst surrounded by some large areolar pores. The apertura is oblique, semicircular, with a very short, rounded rimule. The interzooecial avicularium is elongate, elliptical; its opesium is very large, and its beak is quite rounded.

Measurements.—Apertura $\begin{bmatrix} ha = 0.15 \text{ mm.} \\ la = 0.16 \text{ mm.} \end{bmatrix}$

Variations.—The adventitions organs are quite irregular. Besides the large interzooceial avicularium without pivot, there are still some much smaller avicularia, almost orbicular or somewhat elongate and provided with a pivot. The medium umbo is always very short when it exists; it limits then an irregular slit which is a false rimule. The marginal zooccia are much elongated and sometimes almost cyclindrical.

Affinities.—This species differs from *Discopora verruculata* Smitt, 1872, in the absence of a transverse avicularium on the frontal.

Occurrence.—Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida (common).

Cotypes.—Cat. No. 68711, U.S.N.M.

SCHISMOPORA ABRUPTA, new species. Plate 40, figs. 10, 11.

Description.—The zoarium incrusts shells. The zooecia are indistinct, erect; the frontal is smooth and terminated by a mucro very little salient. The apertura is semicircular, with a wide rimule short and rounded. The ovicell is deeply embedded between the adjacent zooccia; it is transverse, smooth, convex; it is widely open in a locella limited by the medium umbo, and it can not be closed by the operculum. There are numerous, small, triangular avicularia with pivot disseminated between the apertures.

Measurements.—Apertura ha = 0.15 mm. la = 0.15 mm.

Affinities.—The apertura is rarely visible; it is always deeply embedded at the base of a sort of locella. The small avicularia are sometimes more developed; they elongate, become lanceolate, and lose their pivot. In spite of the lack of precision in its exterior characters this species is not very difficult to determine, thanks to the very special appearance of its avicularia.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Cotypes.—Cat. No. 68712, U.S.N.M.

SCHISMOPORA LANCEOLATA, new species.

Plate 40, figs. 12-15.

Description.—The zoarium formed of small lamellar masses creeps on algae. The zooecia are distinct, erect; the frontal is thick, convex, granular, and bears rarely some areolar pores; the medium umbo is terminated by a little salient avicularium. The apertura is semicircular; it bears a triangular sinus, wide and very short. The ovicell is wide, transverse, convex, smooth; it opens into the locella by a large orifice never closed by the operculum. The incomplete zooecia are very salient. The interzooecial avicularia are large, narrow, lanceolate; the pivot is formed by two denticles. The deep zooecia have their umbo very salient, but their frontal is buried.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.17 \text{ mm.} \end{cases}$

Affinities.—This species is quite easy to recognize by its lanceolate avicularia and its small zoarial lamellae. It differs from Schismopora abrupta in the absence of small triangular avicularia.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (rare). Cotypes.—Cat. No. 68713, U.S.N.M.

Genus COSTAZZIA Neviani, 1895.

For description, see Bulletin 106, U. S. National Museum, p. 603.

COSTAZZIA ROBERTSONIAE, new species.

Plate 39, figs. 10-12.

Description.—The zoarium is formed of small, globular or cylindrical masses attached to algae. The zooecia are erect, very salient; the frontal is finely porous. The apertura is pyriform; it bears a wide rimule of little depth. On the peristome there are two small avicularia. The ovicell is small, much embedded between the adjacent zooecia; it bears a small semicircular area, garnished with some large pores. The interzooecial avicularium is small, oval, with very wide beak; it is traversed by a complete pivot.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.15 \text{ mm.} \\ la = 0.12 \text{ mm.} \end{cases}$

Affinities.—The frontal is almost always smooth. Its small frontal pores are only visible on the perfectly preserved zooecia because they became very easily closed by fossilization. This species differs from Costazzia costazzi Audouin, 1828, figured by Miss Robertson and still living in the Californian waters, in its much smaller ovicell.

We dedicate this species to Miss Alice Robertson, whose work on the Bryozoa of California has been a great addition to the science of bryozoology.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (rare).

*Cotypes.—Cat. No. 68714, U.S.N.M.

Genus TEGMINULA Jullien, 1882.

1882. Tegminula Jullien, Dragages du Travailleur, Bryozoaires, Bulletin Société Zoologique de France, vol. 7, p. 510.

Zooecia urceolate, standing erect irregularly one by the side of another; orifice absolutely circular, surmounted by a tubular peristome partly open in front. (Translation after Jullien.)

Genotype.—Tegminula venusta Jullien, 1882. Recent.

Genus CELLEPORA Linnaeus, 1767.

We retain the ancient name Cellepora for those species, which at present can not be placed more definitely.

CELLEPORA MINUTA, new species.

Plate 25, figs. 10-13.

Description.—The zoarium incrusts in thick masses gastropod mollusks; it sometimes emits irregular and ramified branches. The zooecia are small, little distinct, poorly oriented, little erect; the frontal is little convex, more or less large, surrounded by a line of small areolar pores separated by short costules and formed of a pleurocyst detachable from the subjacent olocyst. The apertura is small, suborbicular; two small cardelles, almost median, separate the anter from the somewhat smaller poster. The incomplete zooecia are rare. A very salient small avicularium is between the apertures.

 $\label{eq:measurements} \textit{Measurements.} - \text{Apertura} \begin{cases} ha = 0.08 \text{ mm.} \\ la = 0.07 \text{ mm.} \end{cases} \quad \text{Zooecia} \begin{cases} Lz = 0.35 \text{ mm.} \\ lz = 0.30 \text{ mm.} \end{cases}$

Affinities.—The phenomenon of symbiosis is characteristic of this species. It apparently can live only on a gastropod; we have not a single specimen fixed on any other substratum. The zoarium envelopes the mollusk and ends by killing it; it is rather regular and frequently presents tuberosities formed of raised zooceia. We always have trouble in understanding the selective faculty of the larvae; the latter can not really choose their substratum of fixation; is it therefore a biochemical reaction which permits them to subsist only on the shells of gastropods?

This species is quite close to *Cellepora maculata* Ulrich and Bassler, 1904, in its zoarium which affects the same phenomenon of symbiosis, in its small dimensions and the absence of deep zooeeia. Confusion of the two is very easy, but the present species differs in the almost general presence of a single row of areolar pores and especially in the very constant occurrence of small interzooecial avicularia, which are tubular and very salient.

A sort of umbo, more or less salient, sometimes partially covers the poster. When we know the ovicell it will perhaps be necessary to place this species in a new genus.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (rare). Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (rare).

Cotypes.—Cat. No. 68715, U.S.N.M.

CELLEPORA MACULATA Ulrich and Bassler, 1904.

Plate 25, figs. 14-20.

1904. Lepralia maculata Ulrich and Bassler, Bryozoa Miocene, Maryland Geological Survey, p. 423, pl. 114, figs, S, 9; pl. 118, fig. 7.

Ulrich and Bassler's original description follows.

Zoarium beginning as a thin sheet on shells of small gastropoda to which other layers are added until masses as much as 2 inches in diameter result. Surface of masses generally exhibiting more or less distinct, usually elevated, clusters of zooecia slightly larger than those occupying the intermediate spaces. Zooecia convex, subovate, not sharply separated nor exhibiting any obvious plan of arrangement; when in rows about six occur in 2.0 mm. Orifice not terminal but situated in the anterior half, rounded and

expanded above, contracted below the middle, where there is a small denticle on each side, and nearly straight or curved slightly outward on the lower side; peristome simple, not elevated. Surface punctate excepting over a space just beneath the orifice that is smooth and elevated into a conical or obtuse umbo. Avicularia of two kinds and sizes, both sets very irregularly distributed. Those of the smaller set are ovate and less than half the size of the zooecial orifice, and situated in one of the corners of a zooecium; those of the larger set occupy each the place of a zooecium, have a triangular or acuminate ovate aperture considerably larger than the zooecial orifice, and have the pointed end or side strongly elevated. Ovicells immersed, somewhat smaller than the zooecia, convex, smooth centrally, punctate marginally, often with an eccentric, smooth oval space distinguished from the rest of the surface by an impressed line.

Further study of the types and other specimens show that the original description is perfectly exact. The aperture measures about 0.12 mm. by 0.08 mm. The frontal is garnished with a double row of areolar pores and with a detachable pleurocyst. Between the apertures there is sometimes a very small, round, non-salient avicularium. The large interzooecial avicularium is very characteristic, but it is rather rare.

The zoarium developes into large nodose masses on gastropod shells. Cellepora minuta shows the same phenomenon of symbiosis and, although its other characters are close, its zooecia are also small and of such a kind that it is very difficult to distingush the two species. The present species differs, however, from Cellepora minuta in the absence of the small, very salient avicularium arranged between the apertures and by the great frequency of the double row of areolar pores. Moreover, the zoarium is larger and more spinous than that of Cellepora minuta. We have not yet discovered the ovicell.

Occurrence.—Miocene (Calvert formation): Plum Point and other localities in Maryland (common). Miocene (Yorktown formation): Yorktown, Virginia (rare). Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (very rare). Miocene (Duplin marl): Harvey's Mills, Leon County, Florida (very rare). Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina, and Muldrows Mills, 5 miles south of Maysville, South Carolina (rare).

Cotypes and Plesiotype.—Cat. Nos. 68716-68720, U.S.N.M.

CELLEPORA CRIBROSA Ulrich and Bassler, 1904.

Plate 25, fig. 8.

1904. Cellepora cribrosa Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 429, pl. 117, figs. 5, 6.

The original description by Ulrich and Bassler is as follows:

Zoarium forming small irregular compressed masses. Zooecia very irregularly disposed, some erect, others prostrate, 0.5 mm. to 0.7 mm. long by 0.4 mm. to 0.6 mm. wide; surface strongly punctate; orifice rounded, the normal form showing a slight constriction a little below the middle, where a small tooth projects into the cavity from each side; peristome thick and more or less elevated, ringlike. Avicularia of moderate size, more or less acuminate ovate, attached to and projecting beyond the plane of the inferior side of the peristome; rarely absent. Ovicells few, known only in the broken condition in which they appear as deep semicircular excavations in front of the zooecial orifices.

Only the type specimen of this species has so far been discovered and we are unable to add anything to the original description or to place the species more accurately generically,

Occurrence.—Miocene (Calvert formation): Reeds, Maryland (very rare). Holotype.—Cat. No. 68721, U.S.N.M.

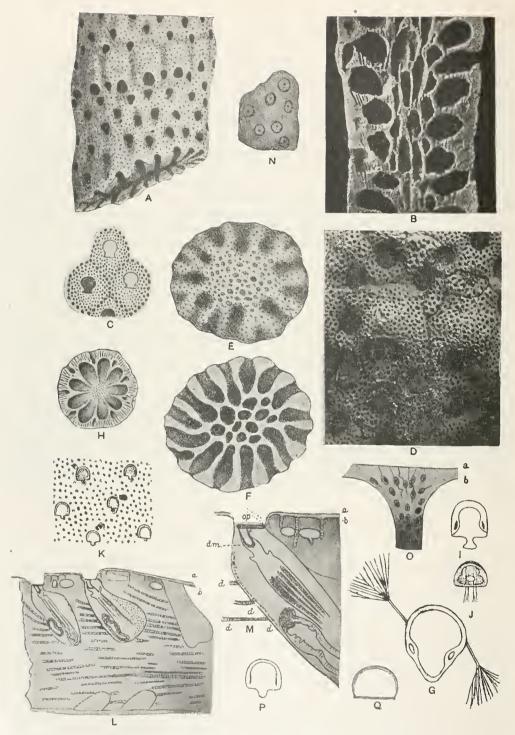


Fig. 34.—Genus Myriozoum Donati, 1750.

A-G. Myriozoum truncatum Pallas, 1766. A. Portion of a fossil zoarium with ovicelled zooecia. (After Manzoni, 1875.) B. Longitudinal section, \times 20, showing ovicelled zooecium in upper left 184

corner. C. Details of the aperture. (After D'Orbigny, 1852.) D. Portion of a recent zoarium, × 20, with several ovicelled zooccia. The operculum closes the aperture. E. Extremity of a zoarium. F. Transverse section. (E, F, after Manzoni, 1877.) G. Operculum, × 85. (After Waters, 1878.) H-J. Myrozoum coarctatum Sans, 1850. H. Transverse section. (After Smitt, 1868.) I. Operculum, × 85. J. Mandible, × 85.

K-O. Myriozoum subgracile D'Orbigny, 1852. K. Portion of a zoarium with avicularia, \times 25. L. Section of tissue, \times 25, showing the polypides in position, the covering integument (a=ectocyst) over the inner one (b); also the long pore tubes which in various places have a disk separating the contents on the two sides of the disk. The oral diaphragm is seen as withdrawn. M. Section through polypide, \times 85, showing: dm, the diaphragm; op, the operculum, with dotted lines to show the position when partly open; d, disks in the pore tubes. N. Covering integument (a), \times 85. O. End of pore tube, \times 500, showing the covering integument (a) and the inner one (b). It contains some cellules of mesenchymatous tissue; P. Operculum \times 85; Q. Mandible, \times 50. (I-2 after Waters, 1900.)

Family MYRIOZOIDAE Smitt, 1868.

The frontal is thick and bears a tremocyst with tubules. Uniporous septulae or dietellae are present. The avicularia are adventitious and bear a pivot. The ovicell is hyperstomial not adjacent to the zooecium and lodged in a nichelike depression of the distal zooecium.

According to Levinsen the genera of this family are as follows: Haswellia Busk, 1884 (referred to the Galeopsidae by us). Gephyrophora Busk, 1884 (Galeopsidae). Muriozoella Levinsen, 1909.

Myriozoum Donati, 1750.

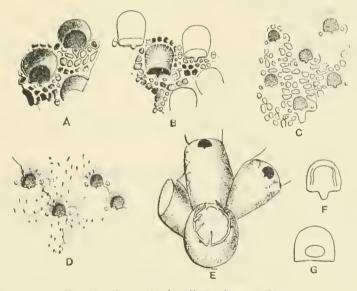


Fig. 35.—Genus Myriozoella Levinsen, 1909.

A-G. Myriozoella crustacea Smitt, 1868. A. Zooecia with peristomes and with cancellated frontal B. Zooecia showing the ovicell. C. Zooecia without peristome, \times 26.5. (A-C, after Smitt, 1868.) D. Zooecia, \times 26.5, much calcified, in which the frontal is not cancellated. E. Ancestrula and ancestrular zooecia, \times 50. F. Operculum, \times 85. G. Mandible, \times 250. (E-G, after Waters, 1900.)

12184-23-Bull, 125-13

So far as known this family is not represented in the American Tertiary, but its species, on the contrary, are of common occurrence in the Miocene and Pliocene of Europe. The larva in all of these genera is unknown and their classification necessarily remains doubtful.

Genus MYRIOZOUM Donati, 1750.

1750. Myriozoum Donatt, Stagie della storia naturale dell' Adriatico.

The zoarium is free, cylindrical, and arborescent. Septulae are present. Sixteen tentacles.

Genotype.—Myriozoum (Millepora) truncatum Pallas, 1766.

Range.—Helvetian-Recent.

Genus MYRIOZOELLA Levinsen, 1909.

1909. Myriozoella Levinsen, Morphological and Systematic Studies on the Cheilostomatous Bryozoa, p. 297.

The zoarium is incrusting. Dietellae are present. Fifteen tentacles. Genotype.—Myriozoella (Myriozoum) crustacea Smitt, 1868. Recent.

ORBITULIPORIDAE, new family.

The zooecia are regularly arranged vertically; the gemmation is lateral. The apertura is terminal. The ovicell is hyperstomial and forms a tube placed in a zooecium larger than usual.

The genera of this family are as follows:

Orbitulipora Stoliczka, 1861.

Stichoporina Stoliczka, 1861.

Batopora Reuss, 1867.

Mamillopora Smitt, 1872.

Sphaerophora Haswell, 1880.

Fedora Jullien, 1882.

Schizorthosecos Canu and Bassler, 1917.

? Diplotaxis Reuss, 1867.

Affinities.—This new family differs from the Conescharellinidae Levinsen, 1909, in the constant presence of an ovicell. It differs from the Myriozoidae in having the ovicell adjacent to a zooecium.

Historical.—In 1917 we included the genera of this new family with the Conescharcllinidae Levinsen, 1909, but after a careful study of this latter family based upon specimens from the Philippine Islands we believe that the analogy between them is purely zoarial and that they must be separated. Their system of incubation is totally different, indicating that their larval system is also very different.

In 1885 Koschinsky discovered in the Bavarian Lutetian a series of forms which he classed in *Stichoporina* Stoliczka, 1881. This error has been repeated by Waters, Kirkpatrick, Neviani, and Canu. Calvet alone, in 1907, compared *Stichoporina* of authors with *Mamillopora* Smitt, 1872. In 1919 Waters established the truth of this observation by a study of some excellent specimens and classified the principal genera as follows:

- A. With a pit: Batopora, Orbitulipora, Sphaerophora, Stichoporina.
- B. Without a pit: Mamillopora, Conescharellina.

We have made only the few following changes in this classification. First, Conescharellina, provided with a peristomial ovicell, belongs to another family; second, we add the American genus Schizorthosecos as a valid genus in the first group; third, we believe it best to separate the group Fedora from Mamillopora until the type of the latter genus is better known.

The hydrostatic function of the central "pit" noted by Waters is unknown. On the inner side of *Schizorthosecos* and of *Mamillopora* there are hydrostatic cavities of different functions, for they are surmounted by ordinary but much

smaller zooccia and there are many of them to a zoarium.

In this group the ancestrula engenders six zooccia and not five as in most other bryozoans. As a result of the studies of Waters in 1919 we have modified some of the generic diagnoses given in our monograph on the early Tertiary Bryozoa. The reader will find studies on the Concscharellinidae in our forthcoming monograph of the recent bryozoa from the Philippine Islands.

Genus BATOPORA Reuss, 1867.

1867. Batopora Reuss, Ueber einige Bryozoen aus dem deutschen Unteroligocän, Sitzungsberichte der K. Akademie der Wissenschaften, Wien, vol. 55, Abth. 1, p. 8.

With a pit toward which the zooecia are directed. Oral aperture small (0.09 mm.), nearly round, but examination shows straight lower edge. Bilaminate. Primary zooecia hidden (Waters). The frontal is a granular olocyst. The zoarium is conical, never hollow and formed of two superposed lamellae.

Genotype.—Batopora stoliczkai Reuss 1867 (probably young of B. multiradiata

Reuss, 1869). Stampian, Tortonian.

The known species of the genus are:

Batopora rosula Reuss, 1847. Tortonian.

Batopora scrobiculata Koschinski, 1885. Lutetian.

Batopora conica Sequenza, 1880. Tongrian.

Batopora stoliczkai Reuss, 1867. Priabonian.

Batopora multiradiata Reuss, 1869. Priabonian.

Genus ORBITULIPORA Stoliczka, 1861.

1861. Orbitulipora Stoliczka, Oligocane Bryozoen von Latdorf in Bernberg, Sitzungsberichte der k. Akademie der Wissenschaften, Wien, vol. 65, Abth. 1, p. 90.

With a pit toward which the zooccia are directed. Oral aperture large, with straight lower edge. Pit on the side bimultilayered (Waters). The frontal is a tremocyst. The zoarium is orbicular and formed of two layers placed back to back.

Genotype.—Orbitulipora haidingeri Stoliczka, 1861.

Range.—Bartonian, Tortonian.

The other known species of this genus are:

Orbitulipora (Cellepora) petiolus Lonsdale, 1850. Bartonian.

Orbitulipora excentrica Sequenza, 1879.

Orbitulipora lenticularis Reuss, 1869. Tongrian.

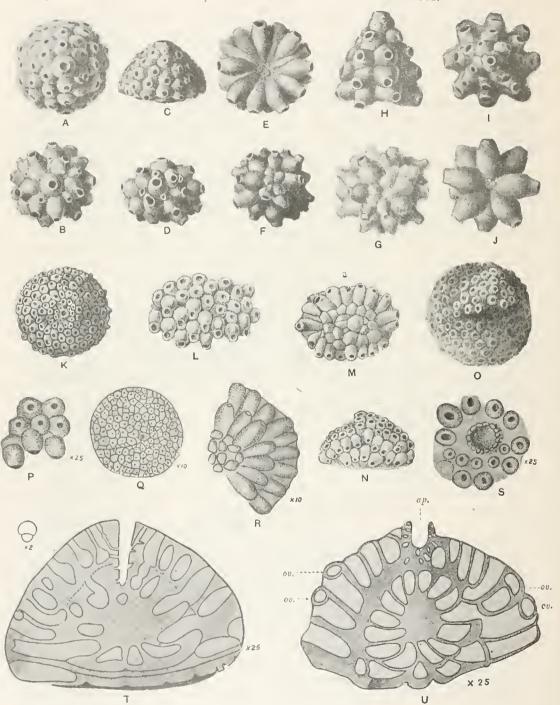


Fig. 36.—Genus Batopora Reuss, 1867.

B, D, F, G. Batopora stoliczkai Reuss, 1867. Frontal, lateral, and basal views of the zoarium. A, C, E, H, I, J. Batopora rosula Reuss, 1847. Frontal, lateral, and basal views of two specimens from the Priabonian. (A-J, after Reuss, 1867.)

K-U. Batopora multiradiata Reuss, 1869. K. Superior face of a colony showing the central pit. L, N. Groups of ovicelled zooecia. M. Inferior face of the colony K. O. Specimen showing a second layer. P. Group of ovicells, × 25. Priabonian at Montecchio-Maggiore, Italy. Q. Specimen from Val di Lonti, Italy (Priabonian), showing a second layer from the neighborhood of the pit, × 10. R. Base showing two circles of zooecia, × 10. S. Smmit of a colony showing the pit with small zooecia around it as well as ordinary zooecia. T. Section showing the pit and the zooecia in a second layer around the first. From near Novezzina, Italy (Priabonian), × 25; a, zoarium showing a cap formed by a second layer of zooecia, from Montecchio-Maggiore, Italy (Priabonian). U. Meridian section showing the apical pit, the second layer and the ovicells. (After Waters, 1891.) ap, aperture; ov, ovicell.

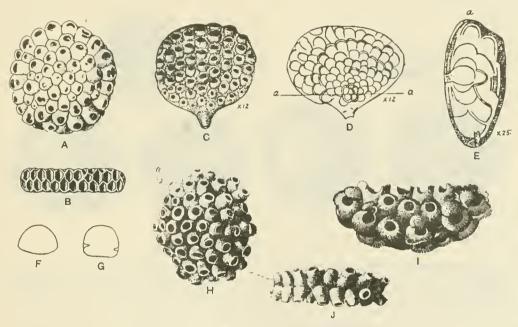


Fig. 37.—Genus Orbitulipora Stoliczka, 1861.

A, B, H-J. Orbitulipora petiolus Lonsdale, 1850. A. An entire zoarium from Latdorf. B. Transverse section showing the two lamellae placed back to back (A, B, after Stoliczka, 1851.) H. An entire bilamellar zoarium. I. Group of ovicelled zooecia. J. Lateral view of the same zoarium as H, showing the two lamellae. (H-J, after Reuss, 1867.)

C-E. Orbitulipora excentrica Seguenza, 1878. C. An entire zoarium showing a lateral pit, \times 12. D. Meridian section showing the ancestrula surrounded by 5 zooecia, \times 12. E. Transverse section following the line a-a at right angles to D, \times 25.

F, G. Orbitulipora lenticularis Reuss, 1869. Oral apertures, × 85. (C-G, after Waters, 1919.)

Genus STICHOPORINA Stoliczka, 1861.

1861. Stichoporina Stoliczka, Oligocäne Bryozoen von Latdorf in Bernberg, Sitzungsberichte der k. Akademie der Wissenschaften, Wien, vol. 65, Abth. 1, p. 92.

"With a pit towards which the zooecia are directed. Oral aperture small. Unilaminate to bilaminate? Pit central." (Waters.)

Genotype.—Stichoporina reussi Stoliczka, 1861.

Genus SPHAEROPHORA Haswell, 1881.

1881. Sphaerophora Haswell, On some Polyzoa from the Queensland coast, Proceedings Linnean Society New South Wales, vol. 5, p. 42.

"With a pit towards which the zooccia are directed. Oral aperture (0.12 mm.) with straight lower edge. Grows in all directions from the early zooccia. Pit central. Multilaminate." (Waters.)

Genotype.—Sphaerophora fossa Haswell, 1881. Range.—Miocene, Recent.

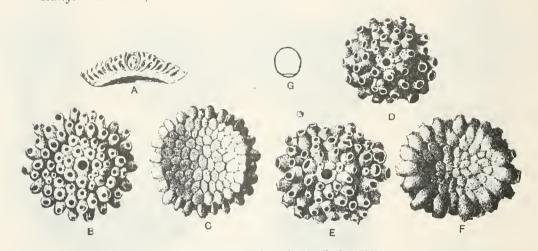


Fig. 38.—Genus Stichoporina Stoliczka, 1861.

A-G. Stichoporina rcussi Stoliczka, 1861. A. Meridian section. B. Exterior side showing the central pit. C. Interior side of the same zoarium. (A-C, after Stoliczka, 1861.) D, E. Exterior side of colonies in which the central pit is surrounded by small zooecia. F. Interior side of E. (D-F, after Reuss, 1867.) G. Form of the orifice, × 85. (After Waters, 1919.)

Genus SCHIZORTHOSECOS Canu and Bassler, 1917.

1917. Schizorthosecos Canu and Bassler, A synopsis of American Early Tertiary Cheilostome Bryozoa, Bull. 106 U. S. National Museum, p. 74.

Completed definition.—With a pit closed externally by a perforated lamella. The zoarium is cupuliform. The apertura is oval with a rounded proximal rimule. There are numerous interzooccial zoocciules which may be transformed into avicularia, into radicular zoocciules or into compensation zoocciules.

Genotype.—Schizorthosecos (Orbitolites) interstitia Lea, 1833. Range.—Claibornian, Jacksonian.

Genus FEDORA Jullien, 1882.

1882. Fedora Jullien, Dragages du Travailleur, Bulletin Société Zoologique de France, vol. 7, p. 17.

Without a pit. The ovicell is hyperstomial, not closed by the operculum, arranged between two zooccia. The apertura presents an anter and a poster separated by two salient cardelles; it is elongated on the ordinary zooccia but transverse and larger on the ovicelled zooccia. The frontal is a granular pleurocyst bordered

by very small areolar pores. There are often one or two avicularia in the vicinity of the apertura. On the inner side the zooccia are hexagonal or fusiform, terminated by a small orbicular pore or covered with large pores.

Genotype.—Fedora edwardsi Jullien, 1882.

Range.-Lutetian to Recent.

The known species of this genus are as follows:

Fedora cdwardsi Jullien, 1882. Recent (Atlantic).

Fedora (Stichoporina) simplex Kirkpatrick, 1890 (not Koschinsky, 1885). Recent (Indian Ocean).

Fedora (Mamillopora) smithi Calvet, 1907 (possibly F. persimplex Neviani, 1895). Recent (Atlantic).

Fedora (Discoflustrellaria) dactylus D'Orbigny, 1852. Lutetian.

Fedora (Stichoporina) simplex Koschinsky, 1885 (=Stichoporina reussi Canu, 1907). Lutetian.

Fedora (Stichoporina) protecta Koschinsky, 1885. Lutetian, Jacksonian.

Fedora (Kionidella) excelsa Koschinsky, 1885. Lutetian, Priabonian.

Fedora (Stichoporina) crassilabris Koschinsky, 1885. Lutetian.

Fedora (Cupularia) bidentata Reuss, 1869. (See Waters, 1919.) Priabonian.

Fedora (Kionidella) obliqueseriata Koschinsky, 1885. Lutetian.

Fedora (Stichoporina) persimplex Neviani, 1895. Plaisancian.

Fedora (Lepralia) minutissima Sequenza, 1880. Helvetian.

Genus MAMILLOPORA Smitt, 1873.

1872. *Mamillopora* Smitt, Floridan Bryozoa, pt. 1, Kongl. Svenska Vetenskaps Akademiens Handlingar, vol. 10, no. 11, p. 33.

Oral aperture large (0.12 mm.), contracted at each side. Primary zooecium erect, surrounded by six similar zooecia. Only unilaminate, showing the position of the zooecia on the under surface. (Waters.) The zoarium is cupuliform. The apertura is elliptical with two submedian cardelles. Avicularia are present. The ovicelled zooecia are much larger; their apertura is not transverse. The zooecia and their inner side are covered with tuberosities.

Genotype. — Mamillopora cupula Smitt, 1872.

The genotype of the genus is incompletely known.

We have had the good fortune to secure some recent specimens of Mamillopora dredged in the Gulf of Mexico. The species appears to be new, but it shows some interesting facts on the structure of the genus. It differs from Mamillopora cupula Smitt, 1872, in having a much smaller and nonbilobed ovicell, in the presence of a peristomial avicularium instead of an interzooccial one, and in having analogous apertures on the ovicelled zooccia. Our specimens were unfortunately insufficient for a complete study, but nevertheless we have observed the following points:

1. The ovicell is closed by the operculum. The operculum of the ovicelled zooecia is identical with the operculum of the other zooecia but often somewhat higher.

2. On the inner, noncellular (superior) side each tuberosity is a pyriform avicularium in which the opesium, which is elliptical and without pivot, bears a small semielliptical mandible. The mandibles are quite variable in dimensions, as some are small and others are large.

- 3. On the inner face there are large cavities which, as they are closed by the ectocyst, are indeed hydrostatic. We have not been able to make zoarial sections of such a nature that we can see by what mechanism the sea water enters or departs.
- 4. The mandibles of the oral avicularia are identical with those of the avicularia of the inner face.
- 5. The ovicells observed were marginal and not inserted between the other zooecia, but our specimens are insufficient to note if this is a constant character.
- 6. This species is very close in its dimensions to *Mamillopora tuberosa* Canu and Bassler, 1919, and differs only in the position of its ovicells and in the constancy of its oral avicularia.

We do not know the inner side of Mamillopora smithi Calvet, 1907. That of Mamillopora cupula Smitt, 1872, has not been figured, but Smitt speaks of "a thin layer of bladders (aborted avicularia) on the back of the zooecia," a description which appears to accord with our observation. Smitt's specimens were deprived of the ectocyst and he was not able to determine the true nature of the avicularia.

MAMILLOPORA TUBEROSA Canu and Bassler, 1919.

Plate 6, figs. 16-19; plate 7, figs. 1-8.

1919. Stichoporina tuberosa CANU and BASSLER, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, no. 291, p. 98, pl. 1, figs. 20-23; pl. 6, figs. 16-19; pl. 7, figs. 1-8.

Description.—The zoarium is free, conical, hollow, with very thick walls. The peristome is salient, ornamented with small tuberosities; it bears one or two small, elliptical avicularia with bar or denticles. The apertura is elliptical, elongated, hidden at the base of a short peristomie; it is formed of a large, semielliptical anter and of a small, concave poster, separated by two small, salient cardelles. The ovicell is large, somewhat salient, convex; it is hyperstomial and always closed by the operculum. A salient, elliptical avicularium, with two denticles serving as pivot, is placed at the base of each zooccium; it deforms the adjacent peristomes. The inner side is tuberose and bears very large pores arranged in quincunx. On the lower face there are large pores surrounded by very small ones.

 $\begin{array}{c} \textit{Measurements.} - \text{Apertura} \left\{ \begin{array}{l} ha = 0.14 \text{ mm.} \\ la = 0.10 \text{ mm.} \\ \end{array} \right. \\ \text{Opesium of} \left\{ \begin{array}{l} ho = 0.10 \text{ mm.} \\ lo = 0.10 \text{ mm.} \\ \end{array} \right. \\ \text{Avicularium} \left\{ \begin{array}{l} Lz = 0.30 \text{ mm.} \\ lz = 0.30 \text{ mm.} \\ \end{array} \right. \\ \text{Avicularium} \left\{ \begin{array}{l} Lav = 0.20 \text{ mm.} \\ lav = 0.12 \text{ mm.} \\ \end{array} \right. \\ \end{array}$

This is a very elegant species characterized by its peristomial tuberosities. The ancestrula is visible only in the interior of the zoarium; it is covered exteriorly by the first zooecia. All the zooecia are separated from each other by small canals which appear to end in the large, inferior pores. The oral tuberosities are hollow. The pores of the internal cavity are avicularia, of which the pivot is formed by two denticles; they are analogous with those of the external face (inferior). The internal face (and upper) bears also large cavities which we believe to be hydrostatic cavities; but it must be proved that the ectocyst is resistant enough to confine an equal amount of water. We must suppose also that these cavities are intended to counterbalance the irregularities of calcification and to assure the perfect equilibrium of the zoarial system.

This species must not be confounded with Mamillopora cupula Smitt, 1872. It differs from it in its ovicell, which is not bilobate, and in its ovarian zooecia which are not larger than the others.

Occurrence.—Lower Miocene (Gatun formation): Banana River, Costa Rica (common). Lower Miocene (Bowden horizon): Rio Cana, Rio Gurabo, and Cercado de Mao, Santo Domingo; Bowden, Jamaica (common).

Cotypes.—Cat. Nos. 68722-68725, U.S.N.M.

Order CYCLOSTOMATA Busk.

Family HETEROPORIDAE Pergens and Meunier, 1886.

Genus CERIOPORA Goldfuss, 1827.

(For description, see Bulletin 106, U.S. National Museum, p. 678.)

CERIOPORA VIRGINIANA, new species.

Plate 26, figs. 1-3.

Description.—The zoarium is an irregular globular mass. The tubes are polygonal; their walls are very thin. No peristomes.

Affinities.—This species may be compared only with the European species from the faluns of Touraine not yet published. Only the figured specimen has been found.

Occurrence.-Miocene (Yorktown formation): 1 mile northeast of Suffolk, Virginia (very rare).

Holotype.—Cat. No. 68726, U.S.N.M.

Family DIASTOPORIDAE Gregory, 1899.

Forma PROBOSCINA Audouin, 1826.

PROBOSCINA MESLERI, new species.

Plate 26, fig. 7.

Description.—The zoarium incrusts shells and forms long triserial branches dividing almost at a right angle. The tubes are long, convex, distinct, porous. The peristomie is perpendicular to the zooecial axis. The apertura is orbicular and the peristome is thin and sharp. The apertures are arranged in quincunx or trigeminal.

Measurements .-

Diameter of orifice..... 0.10 mm. Diameter of aperture 0.40-0.50 nm. Zoarial width..... 0.40-0.45 mm.

This species is named in honor of Mr. Rector D. Mesler, of the United States Geological Survey, who has made important collections of Miocene fossils for our study.

Occurrence.—Miocene (Duplin marl): Wilmington, North Carolina (rare).

Holotype.—Cat. No. 68727, U.S.N.M.

Forma BERENICEA Lamouroux, 1821.

BERENICEA FLABELLUM? Reuss, 1847.

Plate 27, fig. 17.

1847. Diastopora flabellum Reuss, Die fossilen Polyparien des Wiener Tertiarbeckens. Haidingers naturwissenschaftliche Abhandlungen, vol. 2, pl. 7, fig. 9.

The specimens from the Miocene of Virginia which we have referred doubtfully as above have the general zoarial aspect of Reuss's species, but without direct comparisons we are unable to assert their identity positively. The measurements of the American specimens are as follows:

Measurements .-

Peristome 0.24 mm.

Diameter of the orifice 0.18 mm.

Distance between the peristomes 0.80-0.90 mm.

Separation of the peristomes 0.80 mm.

Occurrence.—Miocene (Yorktown formation): Weavers Pond, Gloucester County, and 3 miles southwest of Petersburg, Virginia (rare).

Geological distribution.—Miocene of the Vienna Basin.

Plesiotype. - Cat. No. 68728, U.S.N.M.

ATELESOPORA, new genus.

Greek; Ateles, incomplete; in allusion to the lack of peristome.

The tubes are expanded and have no peristome.

This form of tubes was rather common in the Paleozoic era and during the Cretaceous, and it has been noted in many families. It is very rare in the Tertiary formations and in the recent seas, but its study has never been undertaken seriously.

Our generic definition is incomplete, since we are ignorant of the ovicell. In giving a generic name to this group, our main purpose is to call the attention of the zoologists to these singular remains of ancient periods.

ATELESOPORA REPTANS, new species.

Plate 26, figs. 4-6,

Description.—The zoarium creeps over shells, in salient masses, more or less flabelliform. The tubes are adjacent, irregular, polygonal. The zone of growth is thick and irregular.

Occurrence.—Miocene (Duplin marl): Muldrows Mills, 5 miles south of Mayville, Sumter County, South Carolina (rare); and Natural Well, 2 miles southwest Magnolia, Duplin County, North Carolina (rare). Miocene (Yorktown formation): 3 miles southwest Petersburg, 1 mile northeast, and 1 mile west of Suffolk, 1 mile west of Fort Nonsense, Gloucester County, and Beulahland, King and Queen County, Virginia (rare).

Cotypes. Cat. Nos. 68729-68731, U.S.N.M.

Family MECYNOECIIDAE Canu, 1918.

Forma ENTALOPHORA Lamouroux, 1821.

(For remarks see Bulletin 106, U.S. National Museum, p. 734.)

ENTALOPHORA FASCICULIFERA, new species.

Plate 41, figs. 1-5.

Description.—The zoarium is free, cylindrical, with the Entalophora form of growth. The tubes are long, distinct, separated by a furrow, convex, and slightly striated transversely; at their extremity they are bent almost perpendicularly to the zoarium and are prolonged in a long, free peristomie; they are sometimes bigeminate. The peristomes are thin and widened.

Measurements:

Diameter of the little salient peristomes	0.14 mm.
Length of the peristomie	0.50 mm.
Diameter of the peristome at the extremity of the long	
peristomies	0.20 mm.
Diameter of the tubes on the zoarium.	0.30 mm.
Distance between the peristomes	1.00 mm.
Diameter of the branches	0.80 mm.

Affinities.—The characteristic of this species is furnished by the presence of small bundles formed of two tubes partially joined by their peristomie. The species differs from Mecynoecia proboscidea Milne-Edwards, 1838, in its smaller micrometric dimensions and in the frequent presence of bigeninate zooecia.

Occurrence.—Pleistocene: Santa Monica (Tremochal Canyon) (rare), and Dead Mans Island, off San Pedro, California (very rare).

Cotypes.—Cat. No. 68732, U.S.N.M.

Family ONCOUSOECHDAE Canu, 1918.

Forma FILISPARSA D'Orbigny, 1853.

FILISPARSA CLARKI, new species.

Plate 41, figs. 11-19.

Description.—The zoarium is free, compressed, bifurcated; the zooecia are placed on the anterior face; the dorsal is convex, striated longitudinally and transversally. The tubes are visible, convex, curved outward at their extremity and terminated by a rather long peristomie; the peristomes are thin, orbicular, arranged in quincunx or in oblique rows.

Measurements:

a concordo.	
Diameter of the orifice	0.14 mm.
Diameter of the peristome	0.18-0.20 mm.
Zooecial diameter (on the zoarium)	0.24 mm.
Distance between the peristomes	1.00 mm.
Separation of the peristomes	0.76-0.80 mm.

Affinities.—In its general aspect this species is close to Oncousoccia varians Reuss, 1869; it differs from it in its smaller orifice (0.14 mm. and not 0.20-0.28 mm.) and in the arrangement of its tubes, which are much less regular. The number of longitudinal series of tubes is from three to four. Very rarely a dorsal apophysis is developed. Although we figure the ovicell we have not been able to place this species in its natural genus because the oeciopore has not been observed.

The specific name is in honor of Dr. F. C. Clark, of Los Angeles, California,

who has collected many interesting species for our study.

Occurrence.—Pleistocene: Santa Barbara (very common), and Dead Mans Island, off San Pedro, California (rare).

Cotype.—Cat. No. 68733, U.S.N.M.

FILISPARSA CLARKI, var. PARVULA, new variety.

Plate 41, figs. 20-22.

Description.—The dimensions are smaller than in the typical form. The number of longitudinal series of tubes is five.

Measurements:

Diameter of the orifice	0.14 mm.
Diameter of the peristome	0.16-0.18 mm.
Zooecial diameter (on the zoarium)	0.20 (0.24) mm.
Distance between the peristomes	0.60-0.80 mm.
Separation of the peristomes	0.64-0.90 mm

Variations.—The tubes are visible on the dorsal; their diameter there is 0.04 mm. Through fossilization they disappear and the dorsal appears more or less smooth; accidentally the latter may be transversally wrinkled.

Occurrence.—Pleistocene: Santa Barbara, California (rare). Cotype.—Cat. No. 68734, U.S.N.M.

Family CRISIIDAE Johnston, 1847.

Genus CRISIA Lamouroux, 1816.

(For description see Bulletin 106, U. S. National Museum, p. 703.)

CRISIA SERRATA Gabb and Horn, 1862.

Plate 42, figs. 1-7.

- 1862. Crisia serrata Gabb and Horn, Monograph Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy Natural Sciences Philadelphia, ser. 2, vol. 5, p. 174, pl. 21. fig. 66.
- 1910. Crisia pacifica Robertson, Cyclostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 6, p. 242, pl. 20, figs. 16, 17.

Measurements.—

Diameter of the peristome	0.08-0.10 mm.
Distance between the peristomes	0.36-0.42 mm.
Zooecial diameter	0.12 mm.
Width of a segment.	0.46 mm.
Diameter of the basis ramae	0.14-0.20 mm.

Variations.—This species, which has been known for a long time, is very common in the post-Pliocene formations of California. When the illumination is lateral, the segments present a longitudinal and median keel very characteristic. The distance between the tubes is smaller than the zooecial width. This character is always important to eonsider in the determination of species of *Crisia*.

For reasons of equilibrium there are usually two bases ramae to every segment, one on each side. However, it was not rare to find short segments having only one

basis ramae.

The number of tubes per segment varies from 11 to 16.

Affinities.—There is a rather great difference between our photographs and the drawings given by Miss Robertson of Crisia pacifica. This difference of aspect is due to fossilization; the fossils lose their translucency and the tubes are then scarcely visible.

This species has the general aspect of *Crisia denticulata*, but differs from it in its larger micrometric dimensions, the distance between the peristomes being 0.40 mm. and not 0.30 mm.

Species of *Crisia* are attached to floating algae. The depth at which they are dredged has no bathymetric significance.

Occurrence.—Pleistocene: Santa Barbara (very common), and Santa Monica

(very common), California.

Habitat.—Pacific: Off California (24–48 meters). Plesiotypes.—Cat. Nos. 68735, 68736, U.S.N.M.

Family TUBULIPORIDAE Johnston, 1838.

Genus TUBULIPORA Lamarck, 1816.

(For description see Bulletin 106, U. S. National Museum, p. 753.)

TUBULIPORA FASCICULIFERA Hincks, 1884.

Plate 42, figs. 9-17.

1884. Tubulipora fasciculifera Hincks, Report on the Polyzoa of Queen Charlotte Islands, Annals and Magazine of Natural History, ser. 5, vol. 13, p. 206, pl. 9, fig. 6.

Measurements.—Diameter of the peristome, 0.12 mm.

Variations.—The fascicles are monoserial or biserial, very short, and composed of two to six tubes or more. The zoarium is generally flabelliform, more or less elongate; it creeps over algae. The ovicell is small, ramified between some fascicles only. The oeciostome is wide and little salient.

Affinities.—This species differs from Tubulipora tuba Gabb and Horn, 1862, in its very short fascicles, never composed of more than six tubes, and in its wide and

little salient oeciostome.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Habitat.—Pacific: Off Queen Charlotte Islands.

Plesiotypes.—Cat. No. 68737, U.S.N.M.

TUBULIPORA TUBA Gabb and Horn, 1862.

Plate 42, figs. 18-23.

1862. Semitubigera tuba Gabb and Horn, Monograph Polyzoa of the Secondary and Tertiary formations of North America, Journal Academy Natural Sciences Philadelphia, ser. 2, vol. 5, p. 169, pl. 21, fig. 57.

1910. Tubulipora occidentalis Robertson, Cyclostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 6, p. 249, pl. 22, figs. 29-31.

Measurements.—Diameter of the peristome, 0.12 mm.

Variations.—The zoarium creeps on algae, with generally flabelliform fronds; it sometimes entirely surrounds the delicate radicells and thus becomes tubular. We have observed some specimens on other bryozoa.

The fascicles are monoscrial or biserial; they have a large number of tubes, from 6 to 20; they are often arranged on each side of the zoarial axis in rather irregular order.

The ovicell is large and is inserted between six or eight fascicles. The oeciostome is a very small tube, quite salient, adjacent to the first tube of a fascicle.

Affinities.—This species differs from Tubulipora fasciculifera Hincks, 1884, in its long fascicles composed of more than six tubes and in its long capillary oeciostome.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), (rare), Dead Mans' Island, off San Pedro (rare), and Santa Barbara (rare), California.

Habitat.—Pacific: Off California, and the Queen Charlotte Islands.

Plesiotype.—Cat. Nos. 68738, 68739, U.S.N.M.

Genus IDMONEA Lamouroux, 1821.

(For description, see Bulletin 106, U. S. National Museum, p. 770.)

IDMONEA DISPAR, new species.

Plate 41, figs. 6-10.

Description.—The zoarium is free, bifurcated, with subcylindrical section; the branches are undulated and at a bifurcation; they are not equal. The fascicles are alternate or almost opposite and diverge from each side of the median crest. The tubes are visible, convex, and three or four in number to a fascicle; the peristome is thin and orbicular.

Measurements.—Width of salient fascicles	0.14 mm.
Width of little salient fascicles	0.20 mm.
Diameter of first zooeeium (on the zoarium)	0.18 mm.
Distance between the fascicles	0.40-0.60 mm.
Zoarial diameter	0.80 mm.

Affinities.—This species is quite well characterized by the inequality of the branches at the bifurcations; there is always one a little smaller than the other.

Occurrence.—Pleistocene: Santa Monica (Tremochal Canyon), California (common).

Cotypes.—Cat. No. 68740, U.S.N.M.

IDMONEA PLANULA, new species.

Plate 26, figs. 13-15.

Description.—The zoarium is free, wide, with semielliptical section; the dorsal is flat and striated transversally. The fascicles are salient, opposite, quite close but diverging from the median crest. The tubes are visible, separated by a little salient thread, and number three or four to a fascicle; the peristomes are thin and rectangular.

i eo tangular.	
Measurements.—Width of fascicles	0.20 mm.
Distance between the fascicles	0.40-0.60 mm.
Diameter (on the zoarium) of the first tube	0.24 mm.
Number of the tubes.	3-4.
Width of the zoarium.	1.20 mm.
Tracer of the second	1 70 1 1

Affinities.—This species has the general aspect of *Idmonea petri* D'Archiac, 1846. It differs from it in a greater distance between the fascicles (more then 0.40 nm.) and in the larger zoarial dimensions.

Occurrence.—Miocene (Choctawhatchee marl): Jacksons Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida (very rare).

Holotype.—Cat. No. 68741, U.S.N.M.

IDMONEA CALIFORNICA D'Orbigny, 1852.

Plate 43, figs. 1-9.

1852. Idmonea californica D'Orbigny, Paleontologie francaise, Terrains crétacés, vol. 5, Bryozo-aires, p. 732.

1855. Idmonea californica Conrad, Notes on Miocene and Post-Placene deposits of California, Proceedings Academy Natural Sciences Philadelphia, vol. 7, p. 441.

1862. Idmonea californica Gabb and Horn, Monograph Polyzoa, Secondary and Tertiary formations
North America, Journal Academy Natural Sciences Philadelphia, ser. 2, vol. 5, p. 168,

1910. Idmonca californica Robertson, Cyclostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 6, p. 253, pl. 23, figs. 39-41 (bibliography).

Measurements.—Diameter of the orifice	0.20 mm.
Diameter of the peristome	0.28 mm.
Width of the fascicles	0.28 mm.
Distance between the fascicles	0.80 mm.

Variations.—This giant species seems to be restricted to the American shores of the Pacific.

The fascicles are almost opposite. On the median axis there is a longitudinal row of isolated tubes, often closed by a calcareous diaphragm.

In tangential section the walls are perforated, but the perforations are no larger than in other species of *Idmonea* with small dimensions. In longitudinal section the zooccial walls appear very thick. In transverse section the tubes are rectangular. The ovicell is large, convex, placed on the median crest and finely porous. It surrounds the isolated zooccia of the median axis, which gives it the aspect of *Diaperoecia*, but it does not surround the fascicles and its interfascicular lobes are very short.

Occurrence.—Pleistocene: Santa Monica (rare), Dead Man's Island, off San Pedro (very common), and Santa Barbara, California (very common).

Plesiotypes.—Cat. Nos. 68742, 68743, U.S.N.M.

IDMONEA(?) EXPANSA Ulrich and Bassler, 1904.

Plate 26, fig. S.

1904. Idmonea (?) expansa Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 404, pl. 109, figs. 6-8.

The original description was as follows:

Zoarium adnate, beginning with a single zooecium to which others are added rapidly until an irregular flabellate expansion is produced that with further growth becomes more or less lobate. In the older examples the lobes are seen to be due to the development of the zooecia in systems composed of two pinnate series of transverse rows springing alternately from the opposite sides of a zigzag or wavy median line. In the rows the zooecial apertures, which are rounded quadrate in shape and elevated, are in contact, with four to six in each row and this greater number in about 0.8 mm. The furrows between the rows of apertures are often irregular, and where this in the case the rows themselves are not continuous. When the arrangement is normal the average width of the furrows is a little less than that of the rows of apertures, allowing about four of the latter to come within the space of 1.0 mm. The growing margins of the expansions are occupied by numerous crowded angular cells, decreasing in size toward the extreme edge. Zooecial walls minutely porous.

Occurrence.—Miocene (St. Mary's formation): Cove Point, Maryland (rare). Cotypes.—Cat. No. 68744, U.S.N.M.

Genus CRISINA D'Orbigny, 1852.

CRISINA? STRIATOPORA Ulrich and Bassler, 1904.

Plate 27, figs. 1-4.

1904. Crisina striatopora Ulrich and Bassler, Bryozoa, Miecene, Maryland Geological Survey, p. 406, pl. 118, figs. 1-4.

Ulrich and Bassler's original description was as follows:

Zoarium erect, ramose, probably not exceeding 1 cm. in height, dividing dichotomously at intervals of about 1.5 mm.; branches subovate in cross section, thickest, uniformly convex and traversed longitudinally by from sixteen to twenty punctate striae on the reverse side, narrower and carrying alternating series of zooccial apertures on the obverse side. Zooccial apertures rarely three usually four in each series, in contact laterally, the inner one of each series largest, most prominent, and subcircular the outer one smallest, drawn ont distally and apparently grading into the pores lying between the longitudinal ridges of the reverse side. Series of zooccia curving first forward then slightly backward, separated by a deep interspace averaging about 0.2 mm. in width; about five rows in 2.0 mm. Over the basal part of the zoarium the zooccial apertures are covered one after the other by the growth of the striato-punctate dorsal integument.

In order to properly classify this species thin sections and the nature of the ovicell are necessary. Additional specimens are needed before further studies upon it are made.

Occurrence.—Miocene (Choptank formation): Jones Wharf, Maryland (rare). Holotype.—Cat. No. 68745, U.S.N.M.

Family THEONOIDAE Busk, 1859.

Genus THEONOA Lamouroux, 1821.

THEONOA GLOMERATA Ulrlch and flassler, 1901.

Plate 26, figs. 9-12.

1904. Theonoa glomerata Ulrich and Bassler, Maryland Geological Survey, Miocene, p. 406. pl. 109, figs. 4, 5.

The original description of this species was as follows:

Zoarium cake shaped when young and growing irregular with age, the under side covered with a concentrically wrinkled epitheca, the upper side with short or broken irregularly arranged celluliferous ridges separated by deep interspaces. Ridges abruptly elevated, their flattened summits usually exhibiting a double row of subangular zooecial apertures. Here and there, probably through confluence of two or more ridges, considerable clusters of apertures occur, while other groups may not contain more than three or four cells. Occasionally an irregular radial arrangement of the ridges is apparent. About four zooecial apertures in 1.0 mm.

Occurrence.—Miocene (St. Mary's formation); St. Marys River, Maryland (rare). Cotupes.—Cat. No. 68746, U.S.N.M.

Family DIAPEROECIIDAE Canu, 1918.

Genus STATHMEPORA Canu and Bassler, 1922.

Greek: stathme=line or cord. In allusion to the rectilinear form of the fascicles.

The ovicell is a vesicle traversed by the tubes of which the peristomes are much scattered. The tubes are cylindrical and grouped in linear, uniserial fascicles. Gemmation is triparietal.

Genotype.—Stathmepora flabellata, new species. Pleistocene.

STATHMEPORA FLABELLATA, new species.

Plate 43, figs. 10-17.

Description.—The zoarium is bushy and formed of bilamellar, flabellate fronds. The fascicles are uniserial, little salient, arranged perpendicularly to the zoarial margins. The tubes are visible only when they are isolated. The ovicell is an irregular vesicle pierced by fascicles whose tubes are then more adjacent.

Measurements .-

 Diameter of the peristome
 0. 12 mm.

 Zooecial diameter
 0. 18 mm.

 Distance between the peristomes
 1. 00 mm.

 Separation of the peristomes
 0. 56-0. 80 mm.

Variations.—The fascicles are not exactly analogous to each other. They form, moreover, some lines with tubes adjacent like true fascicles. The basal lamella is large and the zone of growth is thick.

The ovicell is formed after the consolidation of the adjacent tubes, for the fascicles are not disarranged. If the peristomes are scattered the peristomies are long and never adjacent. It is probable that fossilization caused the long non-adjacent peristomies to disappear from the species, of which the bases alone are visible.

12184-23-Bull. 125--14

Affinities. -- This species differs from Mesenteripora meandrina Robertson, 1910, in the ovicell not placed near the zoarial margin, and in its tubes grouped in fascicles.

In spite of exterior appearances the tubes are not arranged as in *Reticulipora*, the axis of the apertura being placed in the longitudinal axis of the zooecia.

Occurrence.—Pleistocene: Santa Barbara (common), and Santa Monica (Rustic Canyon), California (rare).

Cotypes.—Cat. Nos. 68747, 68748, U.S.N.M.

Genus DIAPEROECIA Canu, 1918.

(For description, see Bulletin 106, U.S. National Museum, p. 740.)

DIAPEROECIA FLABELLATA, new species.

Plate 43, figs. 18, 19.

Description.—The zoarium is free, bilamellar and formed of flabellate fronds, and irregularly twisted. The tubes are little visible, widened at their extremity; the peristome is thin, salient, elliptical, horizontal. The ovicell is convex, smooth, traversed by 6–10 tubes; the oeciostome is crescentric and joined to an ordinary peristome.

Measurements .-

Diameter of the peristome. 0. 15 mm.
Distance between the peristomes 0. 42-0. 50 mm.
Separation of the peristomes 0. 50-0. 55 mm.

Occurrence.—Pleistocene: Santa Monica (Tremochal Canyon) (rare), and Dead Mans Island, off San Pedro, California (very rare).

Holotype.—Cat. No. 68749, U.S.N.M.

DIAPEROECIA MILNEANA D'Orbigny, 1839.

Plate 6, figs. 20, 21.

- 1839. Idmonea milneana D'Orbigny, Voyage dans L'Amérique Méridionale, vol. 5, pt. 4, p. 20, pl. 9, figs. 17-21.
- 1919. Idmonea milneana Canu and Bassler, Geology and Paleontology of the West Indies, Bryozoa, Publications of the Carnegie Institution of Washington, No. 291, p. 99, pl. 6, figs. 20, 21.
- 1920. Idmonea milneana Canu and Bassler, North American Early Tertiary Bryozoa, Bulletin 106, U. S. National Museum, p. 773, pl. 136, figs. 1-12.

This recent species has been identified in a number of Tertiary formations of Europe and in the Jacksonian and Vicksburgian of North America. So far the only discovery of it in the post-Oligocene strata of North America is in the Lower Miocene of Santo Domingo, where the single specimen here illustrated has been found.

The discovery of numerous ovicelled specimens of this species in dredgings from the Gulf of Mexico cause us to refer it generically as above.

Occurrence.—Lower Miocene (Bowden horizon): Cereado de Mao, Santo Domingo (rare).

Plesiotype.—Cat. No. 68750, U.S.N.M.

Family LICHENOPORIDAE Smitt, 1866.

Genus LICHENOPORA Defrance, 1823.

(For description, see Bulletin 106, U. S. National Museum. p. 812.)

LICHENOPORA CALIFORNICA Conrad, 1855.

Plate 44, figs. 4-7.

?1852. Unicavea californica D'Orbigny, Paleontologie francaise, Terrains crétacés, vol. 5, Bryo-zoaires, p. 972.

1855. Lichenopora californica Conrad, Note on Miocene and Postpliocene deposits of California, Proceedings Academy Natural Sciences Philadelphia, vol. 7, p. 441.

1910. Luchenopora californica ROBERTSON, Cyclostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 6, p. 261, pl. 25, figs. 48, 49 (bibliography).

Historical.—Unicavea californica is thus defined by D'Orbigny, 1852: "Espèce très convexe endessus, ayant le centre excavé, et pourvue de pores intermédiaires énormes. Madelaine, Basse Californie." Its classification in the genus Unicavea indicates moreover that the colony is discoidal, creeping, incrusting, provided with a single series of cellules in the rows.

Waters, 1905, examined D'Orbigny's type preserved in the Museum of Paris. He notes: "The zooecia are uniserial, slightly raised; zooecia and cancelli about the same size. This does not seem to be the *Discoporella californica* of Busk."

What Conrad, Gabb and Horn, Busk, and Robertson have named *Lichenopora* californica does not appear therefore to be the species of D'Orbigny, as the zoarium is free and the fascicles are biserial. Under these conditions we believe it necessary to substitute the name of Conrad, 1855, for that of D'Orbigny. It is useless to change the specific name, as the specimen of D'Orbigny has not been figured.

Structure.—The structure of this species is quite constant. The zoarium is free and very convex. The fascicles are regular and biserial. The cancelli are very large and often wider than the tubes. The ovicell is placed in the center of the zoarium and hidden by the cancelli. The oeciostome is large, orbicular, salient, placed excentrically in the vicinity of the fascicles.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Plesiotypes.—Cat. No. 68751, U.S.N.M.

LICHENOPORA HISPIDA Fleming, 1828.

Plate 41, figs. 1-3.

1828. Discopora hispida Fleming, History of British Animals, p. 530.

1884. Lichenopora hispida Hincks. Polyzoa of Queen Charlotte Islands, Annals and Magazine Natural History, ser. 5, vol. 13, p. 207.

1889. Lichenopora hispida Jelly, A synonymic catalogue of marine Bryozoa, p. 134 (general bibliography).

1901. Licheno pora hispida Whiteaves, Catalogue of the marine invertebrates of eastern Canada, Geological Survey of Canada, p. 112.

1905. Lichenopora hispida Neviani, Briozoi fossili de Carrubare, Calabria, Bollettino della Societa geologica italiana, vol. 23, p. 554 (52).

Notes on some Recent Bryo. oa in D'Orbigny's collection, Annals and Magazine of Natural History, ser. 7, vol. 15, p. 15.

1907. Lichenopora hispida CALVET, Expéditions scientifiques du Travailleur et du Talisman, Bryozoaires, p. 466 (complementary bibliography).

1907. Luchenopora hispida Nordgaard, Bryozoen von dem norwegischen-Fischereidampfer "Michael Sars," Bergens Museum Aarbog, p. 17.

1908. Lichenopora hispida Canu, Les Bryozoaires du Sud-Ouest de la France, Bulletin societe geologique France, ser. 4, vol. 8, p. 388, pl. 7, fig. 18.

1912. Luchenopora hispida Guérin-Ganivet, Contributions a l'etude des Bryozoaires des côtes armoricaines, III, Bryozoaires de la région de Concarnean et de l'Archipel de Glenan, Travaux scientifiques du Laboratorie de Zoologie de Concarneau, vol. 4, p. 21.

1912. Lichenopora hispida Barroso, Briozoos de la Estaceon de Zoologia maritima de Santauder, Trabajos del Museo de ciencias naturales, no. 5, p. 59.

1913. Lichenopora hispida Guérin-Ganivet, Bryozoaires de la mission arctique du "Jacques Cartier," Société d'oceanographie du Golfe de Gascogne, fasc. 7, p. 42.

1916. Lichenopora hispida F. Canu, Bryozoaires fossiles des Terrains du Sud-Ouest de la France, Bulletin Société géologique de France, ser. 4, vol. 15, p. 333.

1918. Lichenopora hispida Nordoaard, Bryozoa from the Arctic regions, Tromso Museums Aarschefter, vol. 40 (1917), no. 1, p. 21.

Variations.—This species appears to us as poorly defined. Generally authors have classed under this name all specimens in which the fascicles do not reach the zoarial margin, which is therefore garnished with tubes in quincunx. But the ovicells on these various specimens are not exactly similar. The fossils have lost their visor, which renders their determination still more difficult and less certain.

All the fossil species of *Lichenopora* resemble each other, and our determination is perhaps not good because in addition only the two figured specimens have been found and we have been unable to make any useful sections.

The ovicell does not bear cancelli in Fleming's species; and we have not observed the occiostome.

Occurrence.—Pleistocene: Santa Barbara, California (rare).

Geological distribution.—Aquitanian of France (Canu); Helvetian of Italy (Seguenza), of France (Canu collection), Tortonian of Austria Hungary (Reuss); Zanclean of Italy (Seguenza); Plaisancian of England (Busk), of Italy (Manzoni); Astian of Italy (Seguenza); Sicilian of Italy (Seguenza, Neviani); Quaternary of Italy (Seguenza, Neviani).

Habitat.—Arctic Ocean: Greenland, Scandinavia (20–420 meters), White Sea-Finmark. Eastern Atlantic: North Sea, England, English Channel (240–300 meters), Gulf of Gascony. Western Atlantic: Canada. Mediterranean: At Cette (15–90 meters), Corse (40–60 meters), Marseilles, Naples, Oran (75 meters). Pacific: Australia, New Zealand, Queen Charlotte Islands.

In the great depths of the Arctic regions (420 meters) the temperature observed, according to Nordgaard, has been $+5^{\circ}$ C.

Plesiotype.—Cat. No. 68752, U.S.N.M.

LICHENOPORA RADIATA Savigny-Audouin, 1826.

Plate 44, fig. 10.

1826. Melobesia radiata Audoutn, Explication sommaires de planches de polypes de Savigny Description de l'Égypte, vol. 1, p. 235, pl. 6, fig. 3.

1889. Lichenopora radiata Jelly, A synonymic catalogue of marine Byrozoa, p. 137 (general bibliography).

1905. Lichenopora radiata Neviani, Briozoi fossili di Carrubare (Calabria), Bollettino della Società geologica italiana, vol. 23, p. 554.

1910. Lichenopora radiata Waters, Reports on the marine biology of the Sudanese Red sea, The Bryozoa, Journal Linnean Society, London, vol. 31, p. 237 (complementary bibliography).

1910. Lichenopora radiata Robertson, Cyclostomatons Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 6, p. 262, pl. 24, figs. 46, 47.

1912. Lichenopora radiata Barroso, Bryozoos de la estación biológica marítima de Santander.

Trabajos del Museo de ciencias naturales, no. 5, p. 60.

1915. Lichenopora radiata Barroso, Contribución al conocimiento de los Briozoos marinos de España, Boletín de la real sociedad española de Historia natural, vol. 15, p. 419.

Our specimens agree exactly with the typical form of this widespread recent species. According to Waters, there are nine tentacles.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (rare).

Geological distribution.—Miocene of Australia and New Zealand (Waters). Zanclean of Italy (Seguenza); Astian of Italy (Neviani); Sicilian of Italy (Neviani, Seguenza), of Rhodes (Pergens); Quaternary of Italy (Neviani, Waters, Seguenza).

Habitat.—Atlantic: British Channel, Gulf of Gascony (135 meters), Madeira. Mediterranean: Corse, Naples, Adriatic (32–89 meters), Majorca. Pacific: Australia, Japan, and California.

Plesiotype.—Cat. No. 68753, U.S.N.M.

LICHENOPORA VERRUCARIA Fabricius, 1780.

Plate 44, figs. 8, 9.

1899. Lichenopora verrucaria Jelly, A synonymic catalogue of marine Bryozoa, p. 138 (bibliography).

1900. Lichenopora verrucaria Robertson, Bryozoa, Harriman Alaskan Expedition, Proceedings Washington Academy Sciences, vol. 2, p. 329.

1900. Lichenopora verrucaria Nordgaard, Polyzoa, Norske Nordhavs Expedition, Zoology, no. 27 vol. 27, p. 20.

1901. Lichenopora verrucaria Whiteaves, Catalogue of the marine Invertebrata, Canadian Geological Survey, p. 113.

1905. Lichenopora verrucaria Nordgaard, Hydrographical and biological investigations in Norwegian fiords, Bergen Museum, p. 173.

1906. Lichenopora verrucaria Nordgaard, Bryozoa from the Second Fram Expedition, The Society of Arts and Sciences of Kristiania, no. 8, p. 37.

1910. Lichenopora verrucaria Robertson, Cyclostomatous Bryozoa of the west coast of North America, University of California Publications, Zoology, vol. 6, p. 263, pl. 25, fig. 50.

1912. Lichenopora verrucaria Osburn, Bryozoa of the Woods Hole region, Bulletin Bureau of Fisheries, vol. 30, p. 219, pl. 18, fig. 13.

1912. Lichenopora verrucaria Nordoaard, Duc d'Orléans. Campagne arctique de 1907, p. 25.

1912. Lichenopora verrucaria Guérin-Ganivet, Contributions à l'étude des Bryozoaires des côtes armoricaines, III, Bryozoaires de la région de Concarneau et de l'Archipel de Glenan, Travaux scientifiques du laboratoire de Zoologie de Concarneau, vol. 4, p. 22.

1918. Lichenopora verrucaria Nordoaard, Bryozoa from the Arctic region, Tromso Museums Aarschefter, vol. 40, (1917), p. 21.

Variations.—The occiostome of our ovicelled specimen was well preserved in form, very large, salient, and auricular. Unfortunately it was broken in cleaning the specimen. It was larger than the occiostomes figured by Smitt and by Miss Robertson, and it corresponded more to Osburn's figure.

This is a polar species, as it does not pass beyond the fortieth parallel in the Atlantic and the thirtieth parallel in the Pacific. This is the first time that it has been found fossil.

Occurrence.—Pleistocene: Santa Monica (Rustic Canyon), California (very rare).

Habitat.—Arctic Ocean: Finmark, Spitzberg, Kara Sea (166-216 meters), Greenland, Jean Mayen (160-180 meters), Sea of Barents, Iceland (162 meters). Atlantic: Scandinavia (6-24 meters), Denmark (13-18 meters), England, Labrador (24 meters), Canadian coast (11-97 meters), United States to the fortieth parallel. Pacific: Alaska and California.

Verrill has observed this species at 56 meters with a temperature of 6.1° C. *Plesiotype.*—Cat. No. 68754, U.S.N.M.

Family TRETOCYCLOECHDAE Canu, 1918.

Genus TRETOCYCLOECIA Canu, 1919.

(For description, see Bulletin 106, U. S. National Museum, p. 826.)

TRETOCYCLOECIA TORTILIS Lonsdale, 1845.

Plate 28, figs, 1-12.

1845. Heteropora tortilis Lonsdale, Report on the Corals from the Tertiary formation of North America, Quarterly Journal Geological Society London, vol. 1, p. 500, text figure.

1857. Heteropora tortilis Tuomey and Holmes, Pleiocene Fossils from South Carolina, p. 16, pl. 4, figs. 15, 16.

1862. Multicrescis tortilis GABB and HORN, Monograph Polyzoa Secondary and Tertiary formations North America, Journal Academy Natural Science Philadelphia, ser. 2, vol. 5, p. 178.

Structure.—The zoarium is rarely globular but is almost always irregularly cylindrical, branched, attached to the radicells of algae. The ovicell is suborbicular and regularly perforated by the tubes. It is rather deep and of variable dimensions. At the exterior surface the walls of the tubes are thin or thick. The mesopores are rare on certain branches, abundant in groups on others. This irregularity must correspond to conditions of equilibrium which we still do not comprehend very well.

In longitudinal sections the tubes are cylindrical, bifurcating at all heights; their walls are vesicular. The mesopores are irregularly distributed between the orifices.

In transverse sections the tubes are polygonal at the center of the zoarium. The mesopores are always rather short.

In tangential sections the zooccial walls are thick. The interior walls are strongly calcified and appear as thin black lines separated by large clearer spaces corresponding to a less dense calcification.

This is one of the most showy species of the American Miocene.

Occurrence.—Miocene: Yorktown, Williamsburg, and 3 miles southwest of Petersburg, Virginia (common); Smith's Goose Creek, South Carolina.

Plesiotypes.—Cat. No. 68755, U.S.N.M.

TRETOCYCLOECIA AVELLANA, new species.

Plate 27, figs, 5-11.

Description.—The zoarium is free, spherical, born on a flat surface, and of the size of a hazel nut, simple or lobed. The tubes are cylindrical, branched at all

heights, separated by a thick continuous cuticle; the orifice is polygonal with salient peristome. The ovicell is large, irregular, surrounding a score of tubes.

Variations.—The form of the zoarium is quite variable, but it is always a small globular mass simple or mammillated. It grows either on shells or on algae. The small pores observed on the surface of the zoarium are not mesopores but are young tubes in process of formation. The diameter of the normal apertures varies from 0.12 to 0.15 mm.

In longitudinal section the zooccial walls are not vesicular; they show a long, minute canal corresponding to the disappearance of the cuticle which surrounded the tubes on a living specimen.

Affinities.—This species differs from Tretocycloecia tortilis Lonsdale, 1845, in its nonarborescent zoarial form, in the smaller zooecial diameter, and in its larger ovicell.

Occurrence.—Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina (very rare). Miocene (Duplin marl): One-half mile above Edenhouse Point, Chowan River, and 10 miles south of Greenville, North Carolina (rare). Cotypes.—Cat. Nos. 68756-68757, U.S.N.M.

Genus PSILOSOLEN Canu and Bassler, 1922.

1922. Psilosolen Canu and Bassler, Studies on Cyclostomatous Bryozoa, Proc. U. S. National Museum, vol. 61, p. 112.

There are no adventitious tubes. The tubes are cylindrical with peripheral gemmation.

Genotype.—Psilosolen capitiferax, new species.

Range.—Pleistocene, Recent.

The ovicell is a swelling perforated by the tubes as in the Diaperoeciidae, but it is not inserted in the tubes themselves. On the contrary, the ovicell is perpendicular to the tubes and surrounds only the peristomes as in the family Tretocycloeciidae, where this new genus may be naturally classed. The ovicell is little convex and very different from the elongated and very salient sac of the Ascosoeciidae.

It is remarkable to note again that through the geological ages it is the simplest form of the family that has persisted. The Cretaceous and Tertiary genera of this family are provided with adventitious tubes.

PSILOSOLEN CAPITIFERAX, Canu and Bassler, 1922.

Plate 44, figs. 11-21.

1922. Psilosolen capitiferax Canu and Bassler, Studies on Cyclostomatous Bryozoa, Proc. U. S. National Museum, vol. 61, p. 112, pl. 13, fig. 8.

Description.—The zoarium is free, with the form of Entalophora, more or less compressed, dichotomous; the extremity of the branches is enlarged, flattened, and bears the ovicell. The tubes are visible, separated by a furrow, convex, wrinkled transversely, somewhat widened at the summit; the peristome is thin, salient, elliptical or suborbicular. The ovicell is a swelling covering the extremity of a branch; it is perforated by a dozen tubes, some of which are closed by a finely porous diaphragm.

Measurements .--

Structure.—We have made longitudinal and meridianal sections of a number of examples, all of which show the same structure. The zoarial form is a simple compression without any relationship to the structure of the species. In longitudinal section the tubes are cylindrical with peripheral gemmation, reproducing at all heights. The exterior walls of the zoarium are thick.

The transverse section is elliptical in consequence of the compression of the zoarium. The tubes here are of equal size as characteristic of cylindrical tubes. The very small tubes scattered among the large ones are the tubes newly formed by peripheral generation which have not reached their normal diameter.

Variations.—The peristomes are not regularly placed on the zoarium; they are grouped in irregular zones as in *Peripora*. They are little salient, but the irregularity of the dimensions indicates the peristome was prolonged by a very long peristomic broken by fossilization.

The tube measures its greatest width in the portion where it bends away from the zoarium. The tubes which appear exteriorly to arise from an immediately inferior tube are the shorter. Those which appear to arise from before the last row and which slip between the proximal peristomes are the longer.

The ovicell located at the end of the branch is not very salient. It is hollowed out of the zoarium itself, as is easy to verify in the sections.

Affinities.—Our species is almost identical with Entalophora capitata Robertson, of whose variations we are ignorant. It differs from it in its ovicell, which completely covers the end of the branch, and in the smaller micrometric dimensions (if the enlargement indicated on Miss Robertson's figures is exact).

Occurrence.—Pleistocene: Santa Barbara (very common), and Dead Mans Island, off San Pedro, California (rare).

Cotypes .- Cat. No. 68758, U.S.N.M.

BIBLIOGRAPHY OF LITERATURE ON BRYOZOA SINCE 1899.

The following list, arranged chronologically and alphabetically under each year, gives the title of papers upon both fossil and recent bryozoa issued since the publication of the Synopsis of American Fossil Bryozoa in 1900,⁹ in which most of the papers issued before 1899 are eited. On account of the great difficulty of securing literature during the war and since then, it is feared that the lists for the last few years are quite inadequate.

1898.

Canu, F. Etude sur les ovicelles des Bryozoaires du Bathonien D'Occaignes. Bulletin de la Société Géologique de France, ser. 3, vol. 26, pp. 259-285, 20 figs.

1899.

- ALLEN, E. J. On the fauna and bottom deposits near the thirty-fathom line from the Eddystone Grounds to Start Point. Journ. Mar. Biol. Assn., vol. 5, pp. 365-542; Polyzoa, pp. 511, 512, and 534, chart 16.
- Gregory, J. W. Catalogue Fossil Bryozoa in Department Geology, British Museum. Cretaceous Bryozoa, vol. 1, 457 pp. 17 pls.
- Lo Bianco, S. Notizie biologiche riguardanti specialmente il periode di maturita sessnale degli animali del golfo di Napoli. Mt. Stat. Neapel, vol. 13, pp. 448-573; Bryozoa, pp. 493-495.
- Marion, A. F. Notes sur la Faune des Dardanelles et du Bosphore. Annales du Musée de Marseille, ser. 11, vol. 1, 1898, pp. 163–182.
- MATZDORFF, CARL. Jahresbericht über die Bryozoen für 1894, 1895, und 1896. Archiv für Naturgeschichte, vol. 60, pt. 2, pp. 89–126, 1894; issued July, 1899.
- Monroe, C. E., and Teller, E. E. The fauna of the Devonian formation at Milwaukee, Wisconsin. Journal of Geology, vol. 7, pp. 272-283, lists.
- Neviani, A. Briozoi, Idroidi e Foraminiferi di Ciprio, Giaffa ed Alessandria d'Egitto. Comunicazioni preventiva. Bolletino della Societa Zoologica Italiana, Rome, vof. 8, p. 66.
- Opell, W. S. Notes on fresh-water Polyzoa. Ottawa Naturalist, vol. 13, pp. 107-113.
- Petry, S. L. Some Polyzoa, etc., from Walney and Bardsea, North Lancashire. Naturalist, 1899, pp. 59 and 60.
- Pfeffer, G. Über die gegenseitigen Beziehungen der arktischen und antarktischen Fauna. Verhandlungen der deutschen Zoologischen Gesellschaft, 1899, pp. 266–287.
- Richard, Jules. Essai sur les parasites et les commensaux des Crustacés. Archives de parasitologie, vol. 2, pp. 548-595. List of commensal Bryozoa, pp. 575-576.
- Scherren, H. Cristatella mucedo. Nature, vol. 59, p. 150. (Near London.)
- Schultze, L. S. Das Verhältnis der Regeneration und Knospung zur Keimblätterlehre. Jenaische Zeitschrift für Wissenschaft, vol. 33, Bryozoa, p. 293.
- THOMPSON, D'ARCY W. On a supposed resemblance between the Marine Faunas of the Arctic and Antarctic Regions. Proceedings of the Royal Society of Edinburgh, vol. 22, pp. 311-349.
- WHITFIELD, R. P., assisted by Hovey, E. O. Catalogue of the types and figured specimens in the palaeontological collection of the Geological Department, American Museum of Natural History. Bulletin of the American Museum of Natural History, vol. 11, pt. 11.
- Wolleman, A., and Kloos, J. H. Ueber das Vorkommen von Bryozoen und Korallen. Jahresber, Vereins für Naturwissenshaft Braunschweig, vol. 11, p. 200.

BIDENKAP, OLAF. Die Bryozoen, I. Teil. Die Bryozoen von Spitzbergen und Koenig-Karls-Land, Fauna Arctica, vol. 1, Lief 3, pp. 503-540, pls. 9-10.

Boeggild, O. B. The deposits of the sea bottom. Danish Ingolf-Expedition, vol. 1, no. 2, 89 pp., 7 maps, 6 figs.

Calvet, Louis. Contribution à l'histoire naturelle des Bryozoaires ectoproctes marins. Travaux Institute Zoologie de l'Université Montpellier, new ser., Memoire no. 8, 488 pp., 13 pls.

CANU, F. Note preliminaire sur les bryozoaires de Tours. Comptes Rendus Association francaise Avancement Science, Sess. 2me, pp. 406-411.

Canu, F. Contribution a la géologie de Romorantin Paleontologie. Bulletin Société Géologique France, ser. 3, vol. 28, pp. 96-104, 7 figs.

CANU, F. Revision des Bryozoaires du Crétacé figurés par D'Orbigny, 2me Pt. Cheilostomata. Bulletin Société Géologique France, ser. 3, vol. 28, pp. 334-463, 4 pls., 71 figs.

CAR, L. Plumatella (Alcyonella) fungosa Pallas. (Glasnik hrvats. narav. Drustva. God., vol. 12, pp. 142-143.)

CLARKE, JOHN M. The Oriskany fauna of Becraft Mountain, Columbia County, New York. 53d Memoir New York State Museum, no. 3, vol. 3, Annual Report New York State Museum, 1899, vol. 2, pp. 1–128, 9 pls., 1 map.

DAVENPORT, C. B. On the variation of the Statoblasts of *Pectinatella magnifica* from Lake Michigan. American Naturalist, vol. 34, pp. 959-968, 9 figs.

Dendy, Arthur. Cryptopolyzoon an emendation in Nomenclature. Zoologischer Anzeiger, vol. 23, no. 620, pp. 391–392.

Gilson, G. Exploration de la Mer sur les côtes de la Belgique en 1889. Memoires Museum Histoire naturelle Belgique, pp. 1–81, 3 pls. and 10 figs. (List of Bryozoa, p. 34.)

HARMER, SIDNEY F. On the Structure and Classification of the Cheilostomatous Polyzoa. Proceedings Cambridge Philosophical Society, vol. 11, pt. 1, pp. 11-17.

HARMER, SIDNEY F. A revision of the genus Steganoporella. Quarterly Journal Microscopical Science, new ser., vol. 43, pp. 225-297, 2 pls.

Hempel, E. Cristatella mucedo Cuv. aus der Gegend von Chemnitz und gesammelte Statoblasten desselben. Bericht der naturwissenschaftlichen Gesellschaft zu Chemnitz, vol. 14, pp. lxvi-lxviii.

Ladewie, Franz. Ueber die Knospung der ektoprokten Bryozoen. Zeitschrift für wissenshaften; Zoologie, vol. 67, p. 323-339, 1 pl.

Mapplestone, C. W. Further descriptions of the Tertiary Polyzoa of Victoria, Pt. 3. Proceedings Royal Society, Victoria, new ser., vol. 12, pp. 162-169, 2 pls.; vol. 13, pp. 1-9, 12 pls.

NICKERSON, W. S. Double Loxosomae. American Naturalist, vol. 34, pp. 891-895, 6 figs.

Nickles, J. M., and Bassler, R. S. A synopsis of American fossil Bryozoa, including bibliography and synonymy. Bulletin 173, U. S. Geological Survey, 663 pp.

NEVIANI, A. Bryozoi neozoici di alcuna localita d'Italia, pt. 6. Bollettino della Societa Zoologica Italiana, ser. 2, vol. 1, pp. 58-68.

Neviani, A. Briozoi Terziari i Posterziari della Toscana. Bollettino della Société geologica Italiana, vol. 19, pp. 349-375, 6 figs. in text.

Neviani, A. Monografia del Genere Idmonea (Bryozoa Ciclostomata). Pt. 1, Bibliografia generale. Roma, 52 pp.; pt. 2, Cap. 1° Stora del Genere Roma, pp. 53-66.

Neviani, A. Revision Generale dei Briozoi Fossile Italia. Bollettino della Sociéta Geologica Italiana, vol. 19, p. 10–25.

NORDGAARD, O. Polyzoa. Den Norske Nordhavs. Expedition, Zool. no. 27. The Norwegian North Atlantic Expedition (1876-1878), vol. 27, 30 pp., 1 pl., 1 map.

ORTMANN, A. E. Synopsis of the collections of invertebrate fossils made by the Princeton Expedition to Southern Patagonia. American Journal Science, ser. 4, vol. 10, pp. 368-381. (Bryozoa, p. 370.)

Philipps, E. G. Report on the Polyzoa collected by Dr. Willey from the Loyalty Isles, New Guinea and New Britain. Zoological Results, A. Willey, pp. 439-450, pls. 42-43.

Reeker, H. Ueber die Biologie der Susswasserbryozoeu. Jahrbücher des nassauischen Vereins für Naturkunde, Wiesbaden, 27 m. Proc. Verb., pp. 22–26.

Remes, M. Bermerkungen über Süsswasserbryozoen Nordmährens, Verhandlungen der k. k. Zoologisch Botanischen Gesellschaft in Wien, vol. 50, pp. 1-5.

Robertson, Alice. Papers from the Harriman Alaska Expedition, 6. The Bryozea. Proceedings Washington Academy Science, vol. 2, pp. 315-340, 3 pls.

Robertson, Alice. Studies in Pacific Coast Entoprocta. Proceedings California Academy Science, ser. 3, vol. 2, no. 4, pp. 323-348, pl. 16.

ROGERS, A. F. Occurrence of the bryozoan genus *Rhabdomeson* in America. Kansas University Quarterly, vol. 9, pp. 173–174.

Rogers, A. F. New Bryozoans from the Coal Measures of Kansas and Missouri. Kansas University Quarterly, vol. 9, pp. 1-12, pls. 1-4.

Schepotieff, A. Die Pterobranchier des Indischen Ozeans Zeologische Jahrbuch vol. 28, Systematik, pp. 429–448, 2 pls.

TAYLOR, T. H. Investigations made at the marine Biological Laboratory, Plymouth. The Embryology of the Polyzoa. Report 69th Meeting British Association, Dover, p. 437.

Tretiakow, D. Bryozoa und Mollusca des Wolgathals im Government Jaroslav. Transactions Société St. Petersbourg, Comptes Rendus, no. 2, pp. 111-117.

Verrill, A. E. Additions to the Tunicata and Molluscoidea of the Bermudas. Transactions Connecticut Academy, vol. 10, pt. 2, pp. 588-594, pl. 70, 2 text figs.

WATERS, A. W. Bryozoa from Franz-Josef Land, collected by the Jackson-Harmsworth Expedition in 1896-1899. Chilostomata. Journal Linnean Society London, Zoology, vol. 28, pp. 43-105, 6 pls., 3 figs.

ZSCHOKKE, F. Die Tierwelt der Hochgebirgsseen. Denkschr. Schweiz. Gesellshaft, 400 pp., 8 pls. 4 maps. (Bryozoa, pp. 115-119.)

1901.

ALLEN, E. J., and Todd, R. A. The fauna of the Exe Estuary. Journal Marine Biological Association, vol. 6, pp. 295-335. Bryozoa, p. 330.

ALLEN, E. J., and Todd, R. A. The fauna of the Saleombe Estuary. Journal Marine Biological Association, new ser., vol. 6, pp. 151-210. Polyzoa, pp. 204-205.

Arnold, A. F. The sea beach at ebb tide, a guide to the study of the seaweeds and the lower forms of animal life found between tide marks. New York, Century Company, vol. 10, 490 pp., 600 figs. Review, American Naturalist vol. 35, p. 937.

Beede, J. W., and Rogers, A. F. Coal Measures faunal studies. Kansas Quart., vol. 9, pp. 233–254. Blockenhorn, M. Neues zur Geologie und Paläontologie Aegyptens. Zeitschr. Deutsch. geol. ges., vol. 53, pp. 52–132, pls.

CLARKE, J. M. Limestones of central and western New York interbedded with bituminous shales of the Marcellus stage, with notes on the nature and origin of their faunas. Bull. N. Y. Museum, vol. 49, pt. 2, pp. 115-138, pl. 8. (Bryozoa, pp. 133 and 134.)

CORI, C. J., and STEUER, A. Beobachtungen über das Plankton des Triester golfes in den Jahren, 1899 und 1900. Zool. anz., vol. 24, pp. 111–116, with table. (Bryozoa, p. 115.)

CUMINGS, E. R. A section of the upper Ordovician at Vevay, Indiana. American Geologist, vol. 28, pp. 361–380, 2 pls.

Delheid, E. Bryzzaires poederliens determines par le Dr. Pergens provenant du bassin America (Anstruwell-Anvers). Bulletin Société Géologique Belge, vol. 14, p. 332.

Elliot, G. F. Scott; Laurie, M.; and Murdoch, J. B. Fauna of Clyde area. (Marine Polyzoa by A. Gray, Freshwater Polyzoa by M. Laurie.)

Ficheur, E. Note sur le terrain carboniferien de la region d'Igli (Sahara-Oranais). Bulletin Société Géologique France, ser. 3, vol. 28, pp. 915–926. (Bryozoa, pp. 921–922.)

Frič, A. Die thierischen Reste der Perucer Schichten. Archives Landesdf. Bohmen, vol. 11, pp. 163-180.

Gadeau de Kerville, H. Recherches sur les faunes marine et maritime de la Normandie, 3° voyage. Bulletin Société Rouen, vol. 2, pp. 145–224. (Bryozoa, pp. 206–208; list of spp. without descriptions locality, and depth at which found.)

Gradau, A. W. Geology and Paleontology of Eighteen Mile Creek and the Lako Shore sections of Eric County, N. Y. Bull. Buffalo Soc. Nat. Sci., vol. 6. (Bryozoa, pp. 161-176, figs.)

Grabau, A. W. Guide to the Geology and Paleontology of Niagara Falls and Vicinity. Bulletin Buffalo Society Natural Science, vol. 7, no. 1. (Bulletin New York State Museum, No. 45, vol. 9.)

Gray, A. Marine Polyzoa. Fauna Clyde Area. Handbook Natural History Glasgow, pp. 209-214.
 HARMER, S. F. President's address. Bryozoa in Britain. Transactions Norfolk and Norwich Naturalists Society, vol. 7, pp. 115-137.

HERDMAN, W. A. Dredging Expeditions. Transactions Liverpool Biological Society, vol. 15. (Bryozoa, pp. 24, 30, and 31.)

Kukenthal, W. Leitfaden für das zoologische Praktikum. (Bryozoa, pp. 92-96, 2 text figs.)

Maplestone, C. M. Further descriptions of the Tertiary Polyzoa of Victoria. Pt. 5. Procedings Royal Society Victoria, new ser., vol. 13, pp. 183-190, 2 pls., pt. 6, pp. 204-213, 2 pls.

MAPLESTONE, C. M. On a new name Vittaticella for the polyzoan genus Caloporella McG. Proceedings Royal Society Victoria, new ser., vol. 13, pp. 201–203.

MAYER, P. Bryozoa and Brachiopoda. Zoologische Jahresberr, 2 pp. Summary Zoologischer Anzeiger vol. 25, p. 575.

McIntosh, W. C. The coloration of marine animals. Annals Magazine Natural History, ser. 7, vol. 7, pp. 221-240. (Coloration of Bryozoa, p. 230.)

MUDGE, C. P. A textbook of zoology. London, Ed. Arnold, viii, and 416 pp.

Neviani, A. Monografia de genere Idmonea. (Briozoo ciclostomato.) Pt. 2, Cap. 2. Storia delle specie, prima centuria Roma, pp. 67–98, with figs.

Neviani, A. Briozoi neogenici delle Calabrie. Paleontographia Italica, vol. 6, pp. 115–266, 4 pls. Neviani, A. Note bibliografiche su lavori concernenti lo studio di alcuni Briozoari. Bollettino Società Zoologica Italiana, ser. 2, vol. 11, pp. 87–92.

Neviani, Antonio. Nuovi generi e sottogeneri di Radiolari e Briozoi fossili italiana. Bollettino Società Zoologica italiana, ser. 2, vol. 2, pp. 41–43,

Nickerson, W. G. On Loxosoma davenporti, n. sp. An Endoproct from the New England coast. Journal of Morphology, vol. 17, pp. 351-380, pls. 32, 33.

Oppenheim, P. Die Priabonaschieten und ihre fauna. Paleontographica, vol. 47, pp. 137-348, pls. 13-21. (Bryozoa, pp. 260-276.)

Petty, S. L. Some Polyzoa, etc., of the Cumberland Coast, Naturalist, pp. 13-15.

Sardeson, F. W. Problem of the Monticuliporoidea. Journal of Geology, vol. 9, pp. 1-27, pl. A; pp. 149-173, pl.

STENROOS, K. E. Das Thierleben in Nurmijarvi See. Eine faunistischbiologische Studie Acto Soc. Faun. Fenn., vol. 16, no. 1, pp. 38-50, 98-197, etc.

STURANY, R. Bryozoen. Botanik und Zoologie in Österrich in den Jahren 1850–1900. Festschrift, k. k. zool.-bot. Gessellschaft Wien, pp. 267–268.

RUEDEMANN, RUDOLF. Trenton Conglomerate of Rysedorph Hill, Rensselaer County, New York, and its fauna. Bulletin New York State Museum, no. 49. (Bryozoa, pp. 12-14, pls. 1, 2, 3.)

Schulz, Karl. Untersuchungen über den Ban der Bryozoen mit besonderer Berücksichtigung der Exkretionsorgane. Archives Naturges-Jahresbücher, vol. 67, pt. 1, pp. 115-144, pls. 6, 7, 3 figs.

Shipley, A. E. The abysmal fauna of the Antarctic region. Antarctic Manual, chap. 18, pp. 241-275. (Bryozoa, p. 256.)

Shipley, A. E., and McBride, E. W. Zoology: an elementary textbook. Cambridge Natural Science Manuals. Biological Series. Cambridge. (Bryozoa, pp. 282–286.)

SMITH, E. A.; Bell, F. J.; and Kirkpatrick, R. A guide to the shell and starfish galleries (Mollusca, Polyzoa, Brachiopoda, Tunicata, Echinoderma, and Worms), in the British Museum (Natural History), London, printed for the Trustees, v and 130 pp., figs.

WHITEAVES, J. F. Catalogue of the Marine Invertebrata of Eastern Canada. Report Geological Survey Canada. (Separate publication), 272 pp. Bryozoa, pp. 91-114.

Young, John. The Carboniferous Polyzoa of the Clyde Drainage Area. Elliot and others, Fauna, Flora, and Geology of Clyde Area, pp. 486, 489.

Ulmer, George. Lophopus cristallinus Pall. bei Hamburg. Verhandlungen vereins naturwissenschaftlichen Unterh. Hamburg, vol. 11, pp. 195–198.

Ulrich, E. O. Maryland Geological Survey, Eocene, Bryozoa, pp. 205-222, pls. 59, 60.

- Andersson, K. A. Bryozoen während der Schwedischen Expeditionen (ins Nordliche Eismeer) 1898 und 1899 unter Leitung von Professor A. G. Nathorst und 1900 unter Leitung von Conservator G. Kolthoff gesammelt. Zoologischer Jahrbüch Abt. Systematik, vol. 15, pp. 537-560, 1 pl.
- Calvet, L. Bryozoaires marins de la region de Cette. Travaux Institut Zoologie Université Montpellier, ser. 2, mém. no. 11, 103 pp., 3 pl.
- Calvet, L. Bryozoaires marins des côtes de Corse. Travaux Institut de Zoologie Université Montpellier, ser. 2, mem. no. 12, 52 pp., 2 pls.
- Canu, F. Contributions á étude des Bryozoaires fossiles. I, Collection Campiche (Néocomien); II, Collection Dutemple (Sénonien supérieur). Bulletin Société Géologique France, ser. 4, vol. 2, pp. 10-19.
- CONDRA, G. E. New Bryozoa from the Coal Measures of Nebraska. American Geologist, vol. 30, pp. 337-358, pls. 18-25.
- CONTE, A., and VANEY, C. Contributions a l'étude anatomique du Rhabdopleura normani Allman. Comptes Rendus Academie Science, Paris, vol. 135, pp. 63-65.
- Cuminos, E. R. A revision of the bryozoan genera Dekayia, Dekayella, and Heterotrypa of the Cincinnati group, American Geologist, vol. 29, pp. 197-217, 4 pls.
- Forel, F. A. Le Léman, monographie limnologique, vol. 3, Lausanne. (Bryozoa, pp. 113-115, fig. 189.)
- Gardiner, C. J., and Reynolds, S. H. The fossiliferous Silurian beds and associated igneous rocks of the Clogher Head district (County Kerry). Quarterly Journal Geological Society, vol. 58, pp. 226–266 and map.
- GOETTE, A. Lehrbuch der Zoologie. Leipzig. Engelmann, 504 pp., 512 figs.
- HARMER, S. F. Polyzoa. Encyclopedia Brittanica, vol. 31, pp. 826-839, 6 figs.
- HARMER, S. F. On the morphology of the Cheilostomata. Quarterly Journal of Microscopical Science, new ser., vol. 46, pp. 263-350, 4 pls.
- HERDMAN, W. A. Guide to the Port Erin Aquarium. (Bryozoa, pp. 85 and 86, fig.)
- HERTWIG, R. A manual of zoology. Translated from the 5th German edition by Kingsley, New York, Holt & Co., 704 pp.
- Kirkpatrick, R. Polyzoa. Report Collections Natural History Southern Cross, pp. 286-289.
- Labre, A. Review of L. Calvet's Contribution à l'histoire naturelle des Bryozoaires ectoproctes marins. Thése, Paris, 484 pp., 13 pls., 1900, Annee biol., vol. 6, p. 99.
- LANDACRE, F. L. Spouges and Bryozoans of Sandusky Bay. Ohio Naturalist, vol. 1, p. 96.
- Levander, K. M. Förekomsten af Bryozoo-kolonier i Nyländska skärgarden. Meddelanden Societas Fauna et Fennica, vol. 26, p. 6. (Summary, Zoologischer Anzeiger, vol. 25, p. 576.)
- Levinsen, G. M. R. Studies on Bryozoa. Videnskabelige Meddelelser fra den Naturalistorisk Forening Kjobehavn, pp. 1-31. (Summary, Zoologischer Anzeiger, vol. 25, p. 358.)
- MAPLESTONE, C. M. Further descriptions of the Tertiary Polyzoa of Victoria. Pt. 7, Proceedings Royal Society Victoria, vol. 15, new ser., pp. 65-74, pls. 6-8; pt. 8, vol. 15, pp. 17-27, 2 pls.
- MATZDORFF, C. Jahresbericht über die Bryozoen für 1897 und 1898. Archives für naturgeschichte, vol. 61, pp. 173-198.
- MEISSNER, M. Liste der von Herrn Professor Semon bei Amboina und Thursday Island gesammelten Bryozoan. Semon, Zool. Forschg. Malay Archip., vol. 5, pp. 727-731 (=Denk. Ges. Jena) (n. g. Radulina).
- Munthe, H. Stratigrafiska studier öfver Gotlands silurlager. Geologiska Foreningens Stockholm förhandlingar, vol. 24, pp. 221–273. (Bryozoa mentioned in the lists of spp.)
- Neviani, A. Briozoi Ctenostomi fossili. Bollettino della Societa Geologica italiana, vol. 21, pp. 216-220
- Neviani, A. I. Bryozoi pliocenici e miocenici di Pianosa raccolti dal Prof. V. Simonelli e Studiate dal Dott. G. Gioli. Bollettino della Societa Geologica Italiana, vol. 21, pp. 329-343.
- NEVIANI, A. Rhyncopora incurvata n. sp. Bollettino della Societa Geologica Italiana, vol. 21, fasc. 11, pp. 260-262, 3 figs.
- Neviani, A. Sulla Terebripora manzonii, nella Protulophila gestroi Roverto. Bollettino della Societa Geologica Italiana, vol. 21, pp. 41-49, figs.

NEWTON, R. B., and HOLLAND, R. On some fossils from the island of Formosa and Riu-Kiu (Loo Choo), Journal of the College of Science, Japan, vol. 17, art. 6, pp. 1-23, 4 pls.

NICKLES, J. M. Descriptions of New Bryozoan Homotrypa bassleri, n. sp., from the Warren beds of the Lorraine group. Journal Cincinnati Society Natural History, vol. 20, no. 2, pp. 103-105, 5 fige. NICKLES, J. M. The Geology of Cincinnati. Journal Cincinnati Society Natural History, vol. 20,

pp. 49-100.

Pearson, K. Statoblasts of Pectinatella magnifica. Biometrika, vol. 1, p. 128.

PRATT, H. S. A course in invertebrate zoology. A guide to the dissection and comparative study of invertebrate animals. Boston, Ginn & Co.

ORTMANN, A. E. Report Princeton University Expedition to Patagonia. Vol. 4, Paleontology, pt. 2, Tertiary Invertebrates.

Remes, M. Nachträge zur Fauna von Stramberg. I. Die Fauna des rothen Kalksteins (Nesselsdorfer Schiehten). Beitrag Paleontolographica und Geologie Oesterreich Ungarns, vol. 24. (Bryozoa,

RITTER, W. E. A summer's dredging on the coast of southern California. Science, vol. 15. (Bryozoa, p. 62.)

ROBERTSON, ALICE. Some observations on Ascorhiza occidentalis Fewkes and related Alcyonidia. Proceedings California Academy Science, ser. 3, vol. 3, Zoology, pp. 99-108, I pl.

ROVERTO, G. Briozoi, Anellidi e Spugne perforanti del neogene ligure. Paleontographia italiana, vol. 8, 1901, pp. 219-234, pl. 28.

SCHARFF, R. F. (Polyzoa) Report British Association, p. 229.

Schneider, K. C. Lehrbuch der vergleichenden Histologie der Tiere. Jena. (Bryozoa, pp. 223-226, fig. 278.)

THORNELEY, Laura R. Polyzoa from Ballycastle and Rathlin Island. Irish Naturalist, vol. 11, p. I61-I62.

Tolmacev, I. P. Une espece nouvelle des Bryozoaires du calcaire carbonifere inferieure du district de l'Altai. St. Petersburg, Travaux Societe Naturelle, vol. 33, 1.

Ulmer, Georg. Etwas über deutsche Moostierchen. Nerthus Jahrg. 4, pp. 525-528, 541-544, II figs. Vaney, C., and Conte, A. Recherches sur le bourgeonnement de Rhabdopleura normanii All. Comptes Rendus Academie Science Paris, vol. 135, pp. 748-750.

Wanner, Johannes. Die Faunen der obersten weissen Kreide in des libyschen Wüste. Paleonto-

graphica, vol. 30, pp. 91-151.

Wiman, Carl. Uber die Borkholmer Schicht im Mittelbaltischen Silurgebiet. Bulletin Geological Institute, University Upsala, vol. 5, pt. 2, no. 10, pp. 149-222, pls. 5-8.

Zykoff, W. Bemerkung zur Kenntnis der geographischen verbreitung der Süsswasser-Bryozoengat tung Plumatella. Zoologischer Anzeiger, vol. 25, p. 181.

1903.

Adams, G. G.; Girty, G. H.; and White, D. Stratigraphy and paleontology of the upper Carboniferous rocks of the Kansas section. Bull. U. S. Geol. Surv., no. 211, 123 pp.

Andersson, K. A. Eine Wiederentdeckung von Cephalodiscus (McIntosh). Zoologischer Anzeiger, vol. 26, pp. 368-369.

Barnes, J. On a fossil Polyzoan from the Mountain limestone, Castleton (Evactinopora Castletoniensis n. s.). Transactions Manchester Geological Society, vol. 28, pt. 9, pp. 243-245, 1 fig.

Bassler, R. S. The structural features of the bryozoan genus Homotrypa, with descriptions of species from the Cincinnatian group. Proceedings U. S. National Museum, vol. 26, pp. 565-591, pls.

Beede, J. W. Fauna of the Shawnee formation (Haworth), the Wabaunsee formation (Prosser), the Cottonwood Limestone. Coal Measures Faunal Studies II, J. W. Beede and Austin J. Rogers. Kansas Quarterly, vol. I, pp. 163-181.

Bresson, A. Etudes sur les formations anciennes des Hautes et Basses-Pyrénées (Haute Chaine). Bulletin Carte géologique France, vol. 14, no. 93, pp. 1-273.

CALVET, L. Description d'une nouvelle espece de Bryozonire Cténostome du genre Alcyonidium Lamouroux (A. brucei). Bulletin Société Zoologique France, vol. 28, pp. 33-36, 4 figs.

Canu, F. Essal sur une échelle de Bryozoaires pour l'établissment des synchronismes à grande distance. Bulletin Société Géologique France, ser. 4, vol. 3, pp. 115-117.

Canu, F. Note sur la constance de la faune de la Craie de Villedieu. Bulletin Société Gélogique France, ser. 4, vol. 3, pp. 265–268.

Chapman, Frederick. New or little known Victorian fossils in the National Museum. II. Some Silurian Molluscoidea. Proceedings Royal Society Victoria, vol. 16, new ser., pp. 60-82, pls. 10-12.

CONDRA, G. E. The Coal Measures Bryozoa of Nebraska. Nebraska Geological Survey, vol. 2, pt. 1. 168 pp., 21 pls.

CONDRA, G. E. On Rhombopora lepidodendroides Meek. American Geologist, vol. 31, pp. 22-24, 1 pl.

DAVENPORT, C. B. The animal ecology of the Cold Spring Sand Spit, with remarks on the theory of adaptation. Decennial Publications. Chicago, vol. 10, pp. 157-176, 7 figs. (Faunistic.)

Destinez, P. Faune du petit-granite (T₂b) de Belgique. Annales Société géologique Belgique, vol. 30, pt. 1, 27, pp. 71-74.

ETHERIDGE, R. A monograph of the Cretaceous invertebrate fauna of New South Wales. Memoir Geological Survey New South Wales, no. 11, vol. 12, 98 pp., 11 pls.

FAIRE, J. Sur les fossiles crétacés du district de Slavianosserbsk du Government d'Ekatérinoslav. Charikov, Trd. Ohsc. isput, prir., 38, 2, pp. 91-173, 4 pls.

GINESTE, C. L'organogénèse et l'histrogènése au point de vue phylogénique. Trans. Lab. Aracachon, vol. 7, pp. 87-161.

GIRTY, G. H. The Carboniferous formations and faunas of Colorado. Professional Papers U. S. Geological Survey, no. 16, 546 pp.

(Girty, G. H.) Tabulated list of invertebrate fossils from the Carboniferous section of Kansas. Bul letin U. S. Geological Survey, no. 211, pp. 73-83.

Hall, T. S. The possibility of detailed correlation of Australian formations with those of the Northern Hemisphere. Presidential address. Melbourne. (Publication not named. Recorded from separate copy.) Bryozoa, pp. 180-182.

Hamilton, A. On the occurrence of Paludicella in New Zealand. Transactions Proceedings New Zealand Institute, vol. 35, p. 262-264.

HAYES, C. W., and Ulrich, E. O. Illustration sheet, U. S. Geological Survey Geological Atlas, folio 95. HARMER, S. F. On new localities for Cephalodiscus. Zoologischer Anzeiger, vol. 26, pp. 593-594.

Henscher, J. Untersuchungen über die biologischen und Fischereiverhältnisse des Klöntalersees. Pfäffikon-Zürich, Zwingli, ser. 8, 50 pp., 1 pl., 4 figs.

Herdman, W. A. Report of Southport meeting of the British Association. Encrusting Bryozoa forming calcareous masses on the sea bottom in the Gulf of Manaar. Science, vol. 48, p. 614.

HICKSON, S. J. (President's address to the Zoological Section.) Report British Association, 1903. (Manchester mains choked by fresh-water Polyzoa sp.)

Hind, W. Life zones in the British Carboniferous rocks. Report of the Committee. Report 71st British Association, Glasgow, pp. 288-296.

Jacon, C. Sur la signification du gisement cénomanien à Ichtyosarcolithes ét à faune dù Maine de Saint Laurent près Vachères (Basses-Alpes). Comptes Rendus Academie Science, vol. 136, pp. 703-705.

JORDAN, A. Die organischen Reste in den Bohrproben von der Tiefbokrung auf dem Schlachthofe. Abhandlungen naturwissenschafteichen verein zur Bremen, vol. 17, pp. 523-541.

JULLIEN, JULES, and CALVET, LOUIS. Bryozoaires provenant des campagnes de l'Hirondelle (1886-88).

Resultats des Campagnes scientifiques du Prince de Monaco. Fasc. 23, 188 pp., pls. 1-18, pp. 1-120, pls. 1-15 by Jullien; pp. 120-188, pls. 16-18 by Calvet.

KATZER, F. Grundzüge der Geologie des unteren Amazonasgebietes (des Staates Para in Brasilien). Leipsiz. Weber, 8 vol., 296 pp., 1 pl., 261 figs.

KNUDSEN, ——. (Plankton of Northern Europe.) Bulletin des resultats acquis pendant les coursee périodiques publié par le bureau du conseil avec l'assistance de M. Knudsen. Copenhagen, 1902-1903; no. 2, pp. 85-111; no. 3, pp. 147-170; no. 4, pp. 223-309; 1903-1904, no. 1, 62 pp.

Korschelt, E., and Heider, K. Lehrbuch der vergleichenden Entwicklungsgeschiehte der wirbellosen Thiere. Allgemeiner Theil. I, II. Jena, G. Fischer.

Kulczycki, W. Résultats des dernières explorations sur les animanx sous-marins des ocèans atlantique et indien. Kosmos, pp. 453-468.

LANG, W. D. On a fossiliferous bed in the Selbornian of Charmouth. Geological Magazine, ser. 4, vol. 10, pp. 388-392, 1 fig.

LORIE, J. Sondages en Zélande et en Brabant. Bulletin Société Belge geologie, vol. 17, pp. 203-258.

MAPLESTONE, C. M. Further descriptions of the Tertiary Polyzoa of Vietoria, pt. 9. Proceedings Royal Society Victoria, new ser., vol. 16, pt. 1, pp. 140-147, pl. 16, 17.

Nelli, B. Fossili miocenici del Macigno di Porretta. Bollittino Sociétá Géologica Italiana vol. 22, pp. 181-252.

Nordoaard, O. Die Bryozoen des westlichen norwegens. Meeresfauna von Bergen, pp. 75-107, 2 pls. Norman, A. M. Notes on the natural history of East Finnmark. Annals Magazine Natural History, ser. 7, vol. 11, Polyzoa, pp. 567-598, pl. 13; vol. 12, pp. 87-128, pls. 8, 9.

Ostroumoff, A. Sur le développement du cryptocyste et de la chambre de compensation. Zoologischer Auseiger, vol. 27, pp. 96-97.

RIGGENBACH, E. Die Selbstverstummelung der Tiere. Ergebnissne der Anatomie und Entwick lungsgeschichte, 12 (Bryozoa, pp. 828–829, Abth. 2.)

ROBERTSON, ALICE. Embryology and embryonic fission in the genus Crisia. University of California Publications Zoology, No. 3, pp. 115-156, pls. 12-15.

Rowe, A. W. The zones of the white chalk of the English coast, 3, Devon, pp. 1-51, pls. 1-13; 4, Yorkshire, pp. 193-296, pls. 17-21. Publication Geological Association, vol. 18.

TAYLOR, T. H. Investigations made at the Marine Biological Laboratory, Plymouth. The embryology of the Polyzoa. Report 69th British Association, Dover, p. 437.

Todd, R. A. Notes on the invertebrate fauna and fish-food of the bays between the start and Exmouth. Journal Marine Biological Association, vol. 6, no. 4, pp. 541-561.

Van Beneden, E. On demande de nouvelles recherches sur l'organisation et le developpement d'un Phoronis, en vue d'elucider les rapports existant entre les animaux de ce genre, les genres Rhabdopleura et Cephalodiscus, et le groupe des Enteropneustes. Bulletin Academie Belgique, pp. 1216-1232.

1904.

ALLEN, E. J.; TODD, R. A.; PACE, S.; and others. Marine Biological Association. Plymouth marine invertebrate fauna, being notes of the local distribution of species occurring in the neighborhood. Compiled from the records of the laboratory of the Marine Biological Association. Journal Marine Biological Association, vol. 7, pt. 2, pp. 155-298, 1 chart.

AMI, II. M. Preliminary lists of fossil organic remains from the Potsdam, Beekmantown (Calciferous), Chazy, Black River, Trenton, Utica, and Pleistoeene formations comprised within the Perth Sheet (no. 119) in Eastern Ontario. Report Geological Survey Canada, Ottawa, vol. 14, pp. 80-90.

Anonymous Classification. Canadian Naturalist, vol. 31, pp. 210-218. (Bryozoa, p. 212, 2 figs.) (Anonymous.) Polyzoa as food of fishes. Pennsylvania State Department of Agriculture, Monthly Bulletin, vol. 2, no. 8, p. 243.

Anonymous. An expanded polypide of Alcyonella stagnorum. Report Hastings Natural History Society, vol. 11, pl. opp. p. 51.

BOOOJAWLENSKI, N. W. Zur Frage über die Vermehrung von Zoobotryon pellucidus Ehrbg. Mt. Ges. Nat. Moskau, vol. 98. Review by Adelung, Zoologische-Centralblatt, vol. 11, pp. 250-251.

Bolton, H. The paleontology of the Lancashire Coal Measures. Transactions Manchester Geological Society, vol. 28, pt. 1, pp. 378-420; pt. 2, pp. 578-650; pt. 3, pp. 668-689.

Bonarelli, G. Appunti sulla costituzione geologica dell'isola di Creta. Atti Accademia pontifica dei Nuovi Lincei, ser. 5, vol. 3, pp. 518-548, 1 pl. 4 figs. (Bryozoa, pp. 536 and 537 by Neviaui.)

BOVERI, T. Ergebnisse über die Konstitution der chromatischen Substanz des Zellkerns. Jena, 130 pp., 75 figs. Review by Fick, Zoologisches Centralblatt, vol. 11, pp. 97-99.

Calver, Louis. Bryozoeu. Ergebnisse der Hamberger Magalhaensiche Sammelreise, 1892-93, vol. 3, 45 pp. 3 pls.

Calver, Louis. Diagnoses de quelques especes de Bryozoaires nouvelles ou incompletement decrites de la region sub-antarctique de l'ocean Atlantique. Bulletin Société zoologique France, vol. 29, p. 50-59.

Calver, Louis. La distribution géographique des Bryozoaires marius et la theorie de la bipolarite. Comptes Rendus Academie Science, vol. 138, pp. 348-387.

CANU F. Bryozoa. Pal. Univ. Pustulopora semiclausa, 6 figs., with diagnosis.

Canu, F. Bryozoaires fossiles d'Egypte, I. Bulletin institut Egypte, ser. 4, vol. 4, pp. 223-229, 2 pls., 5 figs.

Canu, F. Les Bryozoaires du Patagonien échelle des Bryozoaires pour les terrains tertiares. Memoires Société géologique France, Paleontologie, vol. 12, no. 33, 30 pp., 5 pls., 6 figs.

Canu, F. Étude des Bryozoaires tertiares recuellis en 1885 et 1886 par M. Ph. Thomas dans la region sud de la Tunisie. Exploration scientifique de la Tunisie, pp. 1–37, 3 pls.

GANU, F. Contributions a l'étude des Bryozoaires tertiaires, 111. Description de quelques Membranipores de Tunisie. Bulletin Société Géologique France, ser. 4, vol. 3, pp. 659-660, 1 pl.

CHIRICA, C. Notes sur les Bryozoaires de Roumanie. Annales Scientifique de Université de Jassy, vol. 3, pp. 4-14.

Claus and Grobben. Lehrbuch der Zoologie, begrüdet von C. Claus neu bearbeitet von Karl Grobben. Marburg, 1904, 480 pp., 507 figs. Review by Schuberg, Zoologishes Centralblatt, vol. 11, pp. 233–238.

CLARKE, J. M. Stratigraphic and paleontologic map of Canandaigua and Naples quadrangles. Report New York Museum, vol. 2, App. 4, and Bulletin 63, pp. 1-76. (Bryozea, pp. 46 and 53.)

Couffon, O. Étude critique sur les faluns du Haguineau. Bulletin Société Angers, vol. 33, pp. 35-85. Cyclostomata, p. 44; Cheilostomata, p. 48; list of 36 spp.

CUMINGS, EDGAR R. Development of some Paleozoic Bryozoa. American Journal Science, ser. 4, vol. 17, pp. 49-78, 83 figs.

DAVENFORT, C. B. Report on the fresh-water Bryozoa of the United States, Proceedings U. S. National Museum, vol. 27, pp. 211-221, pl. 17.

Diener, C. Himalayan Fossils. Permian fossils of the Central Himalayas. Memoirs Geological Society of India, Paleontologia Indica, vol. 15, 204 pp., pls. 1–10. (Bryozoa, p. 199.)

Drevermann, F. Die Fauna der Siegener Schichten von Seifen unweit Dierdorf (Westerwald). Palaeontographia 1, 6, pp. 229–282.

ENTZ, G. Az édesvizek élete. Termes. Kozl. Magyar Tars., pp. 616-636, 9 text f.g.

ETHERIDGE, R. Determinations of palaeozoic and mesozoic fossils. Record Geological Survey, Victoria, vol. 1, pt. 1, pp. 10-12.

FOERSTE, A. F. Silurian and Devonian limestones of Termessee and Kentucky. Bulletin Geological Society America, XI, pp. 395–445, 8 figs.

FOERSTE, A. F. The Ordovician-Silurian contact in the Ripley Island area of southern Indiana, with notes on the age of the Cincinnati geanticline. American Journal Science, ser. 4, vol. 18, pp. 323-342, 1 pl.

FOWLER, G. H. Notes on Rhabdopleura normani Allman. Quarterly Journal Microscopical Society, n. s., vol. 48, pp. 23-31, 1 pl.

Fox-Strangways, C. The geology of the Oolitic and Cretaceous rocks south of Scarborough. Memoir Geological Survey England and Wales, Nos. 54 and 55, pp. 1-112, 11 pls., 12 text f.gs. Lists of spp. found.

GARRARD, M. A. Fresh water Polyzoa. Journal Northamptonshire Natural History Society Field Club, vol. 12, p. 158-164.

Gilson, G. Explorations de la mer sur les côtes de la Belgique en 1899. Memoires Museum Belgique, vol. 1, 81 pp.

Grabau, A. W. Paleozoic coral reefs. Bulletin Geological Society America, vol. 14, pp. 337-352, pls. 47 and 48. Bryozoan reefs, p. 348.

Gräntz, — Pflanzengeographisches und Floristisches von Chemnitz. Bericht der naturwissenschaftlichen Gesellschaft zu Chemnitz, vol. 15, p. 1. (Abhandlungen). (Alcyonella füngosa.)

Hall, T. S. Remarks on the deposits. Appendix to Maplestone's tabulated list. Proceedings Society Victoria, vol. 17, pt. 1, pp. 218-219.

12184—23—Bull, 125——15

HAYDEN, H. H. Memoirs of the Geological Survey of India. Memoir Geological Survey, India, No. 36, pp. 1-129.

Hind, W. On the homotaxial equivalents of the Lower Culm of North Devonshire. Geological Magazine, vol. 1, pt. 8, pp. 397-403.

Hinde, G. J. On the zones of Marsupites in the chalk at Beddington, near Croydon, Surrey. Geological Magazine, vol. 1, pt. 10, pp. 482-487.

Holm, G. Paleontologiska notiser. Sveriges Geologiska Undersökning Serien, afh. No. 179, 104 pp., 14 pls.

HOLMES, W. MURTON. List of fossils collected (Surrey), P. Croydon Club, 1903 and 1904, pp. 45 and 46. HUDLESTON, W. H. On the origin of the marine (halolimnic) fauna of Lake Tanganyika. Geological Magazine, vol. 7, pp. 337–382, 2 pls.

HURRELL, H. E. Polyzoa. Transactions Norfolk Society, vol. 7, p. 755.

Jukes-Browne, A. J. The Cretaceous rocks of Britain. The upper chalk of England. Memoir Geological Survey of the United Kingdom, 33, 566 pp. Important list of Bryozoan species, pp. 484-493.

Karaskasch, N. I. Note sur la faune contenue dans les galets de Bolschezemelskaia toundra. Protok St. Peterb. Obshch., vol. 35, No. 3, pp. 130-146. (French summary, p. 162.)

Kishinouye, K. Notes on the natural history of corals. Journal Imperial Fisheries Bureau Tokyo, vol. 14, pt. 1, 32 pp., pls. 1-9.

Kitson, A. E. Report on the Bryozoan limestone at Flinders. Record Geological Survey, Victoria, vol. 1, pt. 1, pp. 44-51. List of species found.

Kitti, E. Geologie der Umgebung von Sarajevo. Jahrbuch geologischen Reichasant. Wien, vol. 53, pp. 515-748, 3 pls. 47 figs.

Knipowitsch, N. Explorations zoologiques sur le bateau casse-glace "Ermak" en été de 1901. Annales Museum Zoologique Academie St. Petersburg. vol. 6, pp. 1–20, 1 pl.

Koford, C. A. Biological survey of the waters of southern California at San Diego. Science, vol. 19, pp. 505-508.

LAMPLUGH, S. W.; KILROE, J. R.; McHENRY, A., SEYMOUR, H. J.; WRIGHT, W. B.; and MUFF, H. B. The geology of the county of Belfast. Memoir Geological Survey Ireland, 166 pp.

Lang, W. D. The Jurassic forms of the "genera" Stomatopora and Proboscina. Geological Magazine, dec. 5, vol. 1, no. 7, pp. 315-322, text figs.

LAUTERBORN, R. Beiträge zür Fauna und Flora des Oberrheins und seiner Umgebung. Mittheilungen der Pollichia eines naturwissenschaft Vereins der Rheinplatz Durkheim, Jahrbuch 60, no. 19, pp. 42–130.

Lebedinsky, G. Die Embryonale Entwicklung der Pedicellina. Travaux Société Naturelle St. Petersbourg, Comptes Rendus, vol. 35, livr. 1, p. 468-471.

LHOMME. Coquilles fossiles trouvées en 1903 dans les sables de Saint-Gobain (Yprésien). Feuille de jeunes naturalists, ser. 4, vol. 34, pp. 103-106. List of spp. collected.

I.INDER, C. Étude de la faune pélagique du Lac de Bret. Revue Suisse Zool, vol. 12, pp. 144-158, pl. 4.

Lindinger, L. Verzeichnis der in und um Erlangen beobachteten Mollusken. Anhang: Ein neurer Fundort von Cristatella mucedo Cuv. Abh. nat. Ges. Nürnberg, vol. 15, pp. 65–84.

Lonmann, H. Eier und sogennante Cysten der Plankton-Expedition. Anhang: Cyphonautes. Ergebn Plankton-Expedition, vol. 4, pp. 1-61, pls. 1-7.

Lomas, J. On Polyzon as Rock-cementing organisms. Report 73d Meeting British Association Advancement Science, pp. 663–664.

Longchamps, M. de Selys. Développement postembryonnaire et affinities des *Phoronis*. Memoire Academie Belgique, vol. 1, 150 pp., 7 pls. Affinities with Bryozoa, pp. 129 and 130.

Loppens, K. Sur une variété de Membranipora membranacea L., et sur quelques animeaux marins vivant dans l'eau saumatre. Annales Société Zoologique Belgique, vol. 38, Bulletin, pp. 152 and 153.

LOPPENS, K. Bryozoaires et Cnidozoaires nouveaux pour la faune Belge trouves par la plupart pendant l'année 1903. Bulletin Société Zoologique Belgique, vol. 39, pp. 65-67.

Luttier, A. Plankyologiska och hydrofaunistiska studier i Lojo sjö under sommaren 1901. Meddelanen Société Fauna Fennica, vol. 28, A, pp. 52-55.

MAEHRENTHAL, F. C. v. Entwurf von Regeln der Zoologischen nomenclatur. Als Grundlage für eine Neubearbeitung der internationalen Regeln der internationalen nomenclatur. Zoologischer Anzeiger, vol. 1, pt. 2, pp. 89-138.

MAPLESTONE, C. M. Notes on the Victorian fossil Selenariidae, and descriptions of some new species (Recent and Fossil). Proceedings Royal Society Victoria, new. ser., vol. 16, pp. 207-216, 2 pls.

MAPLESTONE, C. M. Tabulated list of the fossil Cheilostomatous Polyzoa in the Victorian Tertiary deposits. Proceedings Royal Society Victoria, new ser., vol. 17, pp. 182-217.

Marchal, P. Recherches sur la biologie et le développement des Hyménoptères parasites, 1. La polyembryonie specifique ou germinogonie. Archives zool. exp., vol. 32, no. 3, pp. 257-335, pls. 9-13. Relations of germinogony to ontogeny of Bryozoa, pp. 322-324.

Matzdorff, C. Jahresbericht über die Bryozoen für 1899–1902. Archives Naturges., 1897, vol. 2, pt. 3, pp. 97–146.

MEYER, E. (Affinities of Bryozoa.) Zoologische Jahrbücher Anat., vol. 21, pt. 2, pp. 21-34.

MICHAELSEN, W. Revision der composition Styeliden oder Polyzoinen. Mittheilungen Naturhistorischen Museum Hamburg, vol. 21, pp. 1–124, 2 pls. Affinities of Bryozoa discussed.

Munthe, H. Stratigrafiska studier öfver Gotlands Silurlager. Sveriges Geologiska Undersökning, no. 192, 55 pp.

Neviani, Antonio. Appunti sui Briozoi del Mediterraneo, II. Bollettino Societiá Zoologica Italiana, anno 13, p. 1-3.

Neviani, A. Schizotheca serratimargo Hincks sp. Bollettino Societá Geologica Italiana, vo 1.23, fasc. 11, p. 270–276.

Nobre, A. Subsidios para o estudo da fauna marinha do norte de Portugal. Annales Sciencias Naturales, Porto, vol. 8, pp. 37-94. (Bryozoa, pp. 78-86.)

Nobre, A. Subsidos para o estudo da fauna marinha do sul de Portugal. Annales Sciencias Naturaes, Porto, pp. 153-160. (Bryozoa, p. 159, 18 spp.)

Parks, W. A. Devonian fauna of Kwataboahegan River (Ontario). Bureau of Mines, Report 1904, pt. 1, Bryozoa, p. 185.

Parks, W. A. A remarkable parasite from the Devonian rocks of the Hudson Bay slope. American Journal Science, ser. 4, vol. 18, pp. 135-144.

PARKINSON, J. H. The zoning of the culm in South Germany. Geological Magazine, ser. 5, vol. 1, pp. 272-276.

Polénoff, B. Description géologique de partie nord-ouest de la 15me feuille du VIII zone et de la partie sud-ouest de la 15me feuille du VII zone de la carte générale du gouvernement Tomsk. Travaux géologique St. Petersburg, vol. 3, pp. 133-339.

Ркосна́жа, V. J. Das ostböhmische Miocaen. Arch. nat. Bohmen, vol. 10, pt. 2, 173 pp., 72 fig.

RANGE, P. Das Diluvialgebiet von Lubeck und seine Dryastone nebst einer vergleichenden Besprechung der Glazialpflanzen führenden Ablagerungen überhaupt. Zeitschrift Naturwissenschaften, vol. 76, pp. 161–273, 1 pl. (Bryozoa, p. 238.)

RANSOME, F. L. The geology and ore deposits of the Bisbee Quadrangle, Arizona. Professional Papers U. S. Geological Survey, 21, 112 pp. (Lists of species, pp. 36-38.)

Retzius, G. Zur Kenntnis der Spermien der Evertebraten. Biol. Untersuchen, vol. 11, pp. 1-32, pls. 1-13.

Riche, A. Étude stratigraphique et paléontologique sur la zone à *Lioceras concavum* du Mont d'Or Lyonnais. Annales Université Lyon, fasc. 14, 252 pp., 8 pls. (Bryozoa, p. 207, note of occurrence.) ROSENFELD, G. Studien über das Fett der Meeresorganismen. Wissen-Meeresuntersuch, vol. 5, pl. 21 pp. 57-85.

ROUSSEL, M. Tableau stratigraphique des Pyrénées. Bulletin Carte géologique France, vol. 15, no. 97, pp. 1-116, 3 pls., 66 figs.

ROUSSELET, CHARLES F. On a new fresh-water Polyzoon from Rhodesia, Lophodella thomasi gen. et sp. nov. Journal Quekett Microscopical Club, ser. 2, vol. 9, p. 45-46, 1 pl.

Schepotieff, A. Zur organisation von Rhabdopleura. Bergens Museums Aarbok, no. 2, 21 pp., 3 pls. (Faunistic.)

Seelingero, O. Ueber die Larven und Verwandtschaftbeziehungen der Bryozoen. Archives Ver. Freunde Nat. Mecklenburg, Jahrg. 58, pp. 30-37.

STIASNY, Y. Beitrag zur Kenntniss des Exkretionsapparates der Entoprocta. Arbeiten zoologischen Instituten Wien, vol. 15, pp. 183-196, 1 pl.

STUCKENBERG, A. Coraux et bryozoaires recueillis par N. Sibirtzen dans le government de Vladimir. Bulletin Comité Geologique, vol. 23, pp. 497-502.

STUCKENBERO, A. Anthozoen und Bryozoen des unteren Koklenkalkes von Central Russland. Memoires du Comite Geologique, new ser., vol. 54, pp. 68-199, 9 tables.

Toula, Franz. Geologische Beobdachtuergen auf einer Reise in die Gegend von Silistra und in die Dobrudscha im Jahre 1892. Jahrbuch geologischen Reichs-Anstalt, vol. 54, pp. 1-46, 3 pls.

Ulrich, E. O., and Bassler, R. S. Maryland Geological Survey, Miocene, Bryozoa, pp. 404-429, pl. 109-118.

ULRICH, E. O., and BASSLER, R. S. A revision of the Paleozoic Bryozoa. 1. On genera and species the Ctenostomata. Smithsonian Miscellaneous Collections, vol. 45, pp. 256-294, 4 pls., 2 figs.

Ulrich, E. O., and Bassler, R. S. A revision of the Paleozoic Bryozoa. Pt. 2. On genera and species of Trepostomata. Smithsonian Miscellaneous Collection, vol. 47, pp. 15-55, 9 pls.

Ussing, N. V. Danmarks Geologi i almenfatteligt Omrids. Danmarks geologiske undersogelse, vol. 3, pt. 2, pp. 1-359, 3 pls.

Vanhöffen, E. Die Tierwelt des Südpolargebiets. Zeitschrift Gesellshaft Erdkunde zu Berlin, pp. 362-370, 11 figs.

Verrill, A. E. Additions to the *Tunicata* and *Molluscoidea* of the Bermudas. Transactions Connecticut Λ cademy, vol. 10, pt. 2, pp. 588-594, 4 figs.

Verrill, A. E. Additions to the fauna of the Bermudas from the Yale Expedition of 1901, with notes on other species. Transactions Connecticut Academy, vol. 11, pp. 15-62, pls. 1-11. (Bryozoa, p. 54, list of species only.)

Verrill, A. E. The Bermuda Islands. Transactions Connecticut Academy, vol. 11, pl. 2, pp. 413-913
 Waters, A. W. Bryozoa from Franz-Josef Land, collected by the Jackson-Harmsworth Expedition 1896-97. Pt. 2, Cyclostomata, Ctenostomata, and Endoprocta. Journal Linnean Society London, Zoology, vol. 29, pp. 161-184, 3 pls.

Waters, A. W. Résultats Voyage Belgica. Zoologie, Bryozoa, 114 pp., 9 pls., 3 figs.

Weigelt, C. L'assainissement et le repeuplement des rivières. Memoir Cour. Academie Belgique, no. 64, (Bryozoa, p. 516, fig. 83).

Wesenberg-Lund, C. Studien over Danske Soers Plankton. Specielle Del. (English summary) 223 pp. Copenhagen, 1904. (Review by Zschokke, Zoologischer Centralblatt, vol. 11, pp. 680-687.

Whiteaves, J. F. Preliminary list of fossils from the Silurian (Upper Silurian) rocks of the Ekwan River and Sutton Mill Lakes, Keewatin, collected by D. B. Dowling in 1901, with descriptions of such species as appear to be new. Report Geological Survey, Canada, vol. 14, pt. F, pp. 38-59.

Whitfield, R. P. Notice of a new genus and species of Lower Carboniferous Bryozoan. American Museum of Natural History, Bulletin, vol. 20, p. 469, pl. 11.

Wollemann, A. Ein Aufschluss im Mukronatensenon bei Rotenkamp, nordwestlich von Königslutter, vol. 13. Jahresbericht Vereins für Naturwissenschaften, Braunschweig, pp. 40–42.

Wolterstrorff, W. Beitrage zur Fanna der Tucheler Heide. Bericht über eine Zoologische Bereisung der Kreise Tuchel und Schwetz in Jahre, 1900. Schr. Ges. Danzig, vol. 11, pts. 1 and 2, pp. 140-234, 1 pl.

Wood, Elvira. Marcellus (Stafford) limestones of Lancaster, Erie County, New York. Report New York State Museum vol. 55, pp. 139–181. (Bryozoa, p. 156. List of species found.)

1905.

AMEGHINO, F. L'Age des Formations sédimentaires de Patagonie. Critique de Hatcher. Annales Societá Argentina, vol. 51, (nos. 3 and 4) p. 65. (Bryozoa, p. 88.)

Anonymous. The geology of the country round Cork and Cork Harbour. Memoirs Geological Survey, 1reland. (Geology of Cork, pp. 1-126; Bryozoa, p. 27.)

Bidenkap, Olaf. Fortegnelse over de arktiske bryozoa. Bergens Museum Aarbok, no. 9, 79 pp. Brehm, V. Zur Kenntniss der Mikrofauna des Franzensbader Torimoordistriktes Bryozoa. Archives Hydrobiologie und Planktonkunde, vol. 1, pp. 211–233.

Brown, E. T. Note on the pelagic fauna of the Firth of Clyde. Proceedings Royal Society of Ediuburgh, vol. 25, pp. 779-791.

Buen, Odon De. La région méditerranéenne des Baléares. Bulletin Société Zoologique France, vol., 30, no. 5 (Bryozoa, p. 104).

Burrows, A. W. Note on a bryozoan attached to Neptunea found in one of the Mekran nodules. Geological Magazine, ser. 2, dec. 5, vol. 2, no. 7, pp. 303-305, fig. 1.

Calvet, L. La station zoologique de Cette. Travaux Institut Montpellier, ser. 2, mem. no 15 (Bryozoa, list of, pp. 61-63).

Calver, L. Liste de Bryozonires marins de collections du musée Royal d'Historie naturelle de Bruxelles. Annales Société malacologique Belgique, vol. 39, pp. 1–8.

CAREZ, L. La géologie des Pyrénées Francaises. Feuilles de Tarbes et Luz. Groupe primaire. Système Permien. Geol. Pyrénées. Memoires Carte Géologique France, Fasc. II (Bryozoa, pp. 797 and 798).

CHAPMAN, F., and McCoy. Notes on fossil easts in Tertiary ironstone from Stawell. Victorian Naturalist, vol. 21, pp. 178-180.

CLARKE, J. M. Type specimens of paleozoic fossils in New York State Museum. Report New York Museum, vol. 56 (I-IV), and Bull. 65, pp. 1-847. (Bryozoa, pp. 94, 187, 786, 816, 817, 821, 823, 829, 833, 839, 846.)

CLARKE, E. Fossils of Waitemata and Papakura series. Transactions New Zealand Institute, vol. 37, p. 413. (Bryozoa, p. 415.)

COLGAN, N. Notes on the invertebrate fauna of Skerries, County Dublin. Irish Naturalist, vol. 14, pp. 205-213. (Bryozoa, p. 209.)

Couffon, O. Bryozonires. Étude critique sur les Faluns de Chalonnes. Bulletin Société Angers, vol. 34, pp. 165-214.

Couffon, O. Les faluns de l'Anjon et de la Lonraine dans le Saumurois. Bulletin Société Angers, new ser, vol. 33, pp. 216-223. (Bryozoa, p. 218.)

CRAGIN, F. W. Paleontology of the Malone Jurassic formation of Texas. Bulletin U. S. Geological Survey, no. 266, pp. 172, 3 pls.

CUMINGS, E. R. Development of Fenestella. American Geologist, vol. 35, pp. 50 and 51.

CUMINGS, EDGAR R. Development of Fenestella. American Journal Science, ser. 4, vol. 20, pp. 169-177, 3 pls.

Dacqué, E. Beiträge zur Geologie des Somalilandes. 1 Teil, Untere Kreide. Beiträge Paleontologie und Geologie Osterreich-Ungarns und des Orient, vol. 17, Heft 1 and 2, pp. 7-20 (2 and 3); 2 Teil, Oberer Jura, Heft 3 and 4, pp. 119-160 (with pls. 14 and 18).

DERYNTGHINZH, K. Otochezh po osorudovananiyn Murmanskoi Biologhicheskoi Stanjn i lyetniya rasotzhina nei vzh 1904. Protok St. Petersberg Obshch, vol. 36, pt. 1 (2, 3), pp. 78-114. (Bryozoa, p. 99.)

Desmazières, O. Notes paléontologiques sur l'Arrondissement de Segré. Bulletin Société Angers, vol. 36, pp. 123-130.

Destinez, P. Complément de la Fanne des psammites du Condroz. Annales Société géologique Belgique, vol. 32, pt. 2, pp. 123–127. (Bryozoa, p. 127.)

Douglas, Earl. Some notes on the geology of sonthwest Montana. (Bryozoa in Carboniferous formation.) Annales Carnegie Museum, p. 419.

Dublin, L. The history of the germ cells in *Pedicellina americana*. (American Society Zoology) Science, p. 381, new ser., vol. 21.

Dublin, L. The history of the germ cells in *Pedicellina americana* (Leidy). Annals New York Academy Science, vol. 5, pp. 1-55, 3 pls., 2 figs.

Dunlin, L. On the Nucleoli in the Somatic and germ cells of *Pedicellina americana*. Biological Bulletin, vol. 8, pp. 345-364, 14 figs.

Fearnsides, W. G. On the geology of Arenig Fawe and Moel Llynfnant. Quarterly Journal Geological Society, vol. 61, pp. 608-637, 1 pl., 1 map, 2 figs.

Fox-Strangeways, C. Carboniferous limestone fossils from South Derbyshire. Memoir Geological Survey United Kingdom, no. 141 (Bryozoa, p. 15).

Fox, Howard. Further notes on the Devonian rocks and fossils in the parish of St. Minver. (With notes by F. A. Bather, G. C. Crick, W. A. E. Ussher, and H. Woodward). Penzance, Transactions Royal Geological Society Cornwall, vol. 13, pp. 33-87.

Hallez, Paul. Rhéotropisme de quelques Hydroides monosiphonés et des Bugula. Comptes Rendus Academy Science Paris, vol 141, p. 840-843.

HALLEZ, PAUL. Notes Fauniques. Bryozoaires, Archives Zoologue experimentale et generale Zool. Exp., ser. 4, vol. 3, p. 49.

Harbort, E. Die Fauna der Schaumburg-Lippeschen Kreidemulde Berlin, Abhandlungen der kgl. preussischen geologischen Landesanstalt, vol. 45, pp. 1-112.

HARMER, S. F. The Pterobranchia of the Siboga Expedition, with an account of other species. Siboga Expedition Monograph, no. 26, 132 pp., 14 pls., 2 figs.

*Hennig, Anders. Gotlands Silur-Bryozoer, I. Arkiv för Zoologi K. Svenska Vetenskapsakademiens i Storkholm, vol. 2, no. 10, 37 pp., 2 pls., 35 figs.

Hudson, George H. Contributions to the fauna of the Chazy limestone on Valcour Island, Lake Champlain, New York State Museum, Bulletin 80, pp. 270-295.

JOHNSON, CH. W. Annotated list of the types of invertebrate Cretaceous fossils in the collection of the Academy of Natural Sciences, Philadelphia. Proceedings Academy Philadelphia, vol. 57, pp. 4-28. (Bryozoa, pp. 5 and 6.)

KAZARSKII, P. Materialen zur Kenntniss d. Devon-Fauna d. Urals. Trudui Kazan Univ., vol. 34, no. 2, pp. 1-51. (Bryozoa, p. 29.)

Kemp, Helen P. Bryozoa. (Bibliography for 1905.) Zoological Record, London, vol. 42, p. 10. Klaer, H. Dyfelivet i Drøbaksund. Nyt magazin Naturvidenskaberne, pp. 61-89, pls. 2 and 3.

KNUDSEN, M. Résultats acquis pendant les courses périodiques Plankton (Norwegian, English, Belgian German, Danish, and Swedish waters). Bulletin, Conseil Permanent International pour l'exploration de la mer 1904-1905. (Cyphonautes, no. 1, d, pp. 2, 8, 10, 14, 18, 27, 30, 35, 42; no. 2, d, pp. 64, 66, 72, 76, 79, 81, 83, 85, 86, 92; no. 3, d, pp. 108, 110, 114, 115, 117, 124; no. 4, d, pp. 137, 142, 151, 152, 160, 176, 180.)

LAMANSKI, V. Die aeltesten silurischen Schichten Russlands (Etage B). St. Petersbourg, Memoires comite geologique, new ser., vol. 20.

LANO, W. D. On Stomatopora antiqua Haime and its related Liassic forms. Geological Magazine, dec. 5, vol. 2, no. 6, pp. 258-268, table, pl. 14, text fig.

LANKESTER, E. R. On a new species of Cephalodiscus from the Antarctic Ocean. Proceedings Royal Society London, vol. 76 B, p. 400-402.

Lebedinsky, ——. Die Embryonal entwicklung der *Pedicellina echinata* Sars. Biologisches Centralblatt, vol. 25, p. 536-548, 2 figs.

LOPPENS, K. Bryozoaires. Bulletin Société zoologique malacologique Belgique, vol. 39, p. 14.

Maplestone, C. M. Lord Howe Island Polyzoa. Proceedings Royal Society Victoria, vol. 17, pp. 386-390, 2 pls.

McIntosh, W. C. Budding in animals. Zoologist, January 1905, pp. 1-21.

Meissner, Wallerian. Ueber die Winterlauna im Kaban-See. Trudui Kazanskom Universitetye, vol. 39, ser. 3, 1904, pp. 1-118, 1 pl.

MERRILL, G. P. (and others). Catalogue of the type and figured specimens of fossils, etc. Bulletin U. S. National Museum, no. 53, pt. 1, 1905.

Monroe, Cfr. E. List of fossils from Bethany, New York (Hamilton beds). Bulletin Society Wisconsin, vol. 2, pt. 1, p. 63.

Muntue, Henr. Om den submoräna Hernögyttijan och dess Alder. Sveriges geologiska undersökning Afh. Ser C. Co., vol. 196, pp. 1–32. (Bryozoa, p. 13.)

NAPSAL, Dr. Seznam zkamenčlin v billem vápenci na Stramberku naizenych. Vestnik. Česke Ak., vol. 13, pp. 288–295. (Bryozoa, p. 291.)

Napsal, Dr. Seznam zkamenělin z cerveného vápence kopřvoníckého. Vestnik Česke Ak., vol. 13, pp. 360-379. (Bryozoa, p. 361.)

NEVIANI, A. Sulla Schizothcae serratimargo Hincks. Napoli. Aunuario del Museo zoologico, Universita di Napoli, new ser. 2, no. 1, pp. 1-6.

Neviani, A. Materiali per una bibliografia degli studi sui Bryozoi viventi e fossili dal 1800 al 1900. Bollettino Naturalista Siena, Anno 20, pp. 104-109, 122-125; Anno 21, pp. 4-8, 29-33, 47-50, 66-67, 102,105, I29-113; Anno 22, pp. 24-28, 44-47, 53-55, 71-73, 96-98; Anno 23, pp. 11-15, 31-34, 46-50, 59-62, 75-76, 90-91, 101-102, 113, Anno 24, pp. 1-10; Anno 25, pp. 21-22.

NEVIANI, ANTONIO. Briozoi fossili di Carrubare (Calabria). Bollettino Società Geologica Italiana,

vol. 23 (fasc. 3.), p. 507-555, 21 figs.

NEVIANI, ANTONIO. Di alcuni Briozoari eocenici di Villatorta (Spagna). Bullettino della Societià Geologica Italiana, vol. 24, p. 158-163.

NICKLES, J. M. The Upper Ordovician rocks of Kentucky and their Bryozoa. Kentucky Geological Survey, Bulletin no. 5, 64 pp., 3 pls.

Nobre, A. Fauna Portugesa. Ectoprocta. Annuar Ac. Porto, 1903-4, p. 115.

NORDGAARD, O. Hydrographical and biological investigations in Norwegian fiords. Bergen Museum, 254 pp., 21 pls. Bryozoa, pp. 164-174, pls. 3-5.

NORMAN, A. M. Notes on the natural history of East Finmark. Annals and Magazine of Natural History, ser. 7, vol. 15, pp. 341-360, pl. 27.

PEETZ, H. DE. Description géologique de la 13 feuille (X zone) de la carte generale du governement. Tomsk (feuilles; Zmélnogorsk, Bieloglasowo, Loktewsky, Sawod et Kabania). Travaux Sect, Geol. Cab. S. M. 1. St. Petersburg, vol. 6, pp. 1-273.

Retzius, G. Zur Kenntniss der Spermien der Evertebraten. Biol. Untersuch., vol. 13, pp. 79-102, 8 pls. (Bryozoa, pp. 89-90.)

Retzius, G. Das sensible Nerven system der Bryozoen. Biol. Unters., new. ser., vol. 12, p. 49-54, 1 pl.

RICHET, ——. Génération. Dict. de Physiologie, 1905 (Bryozoa, p. 70).

ROBERTSON, ALICE. Nonincrusting Chilostomatous Bryozoa of the west coast of North America. University of California Publications, Zoology, vol. 2, no. 5, pp. 235-322, pls. 4-16.

RÖMER, F. Bericht der Senkenbergischen Naturforschenden Gesellsch. in Frankf. am. Main, 1905. Museumsbericht, No. 1. Zool. Sammlung, p. 186. Bryozoen an d. Norwegischen Küste.

SCHARDT u. DUBOIS. Descriptions Géologiques de la région des Jura Neufchätelois. Bulletin Society Neuchatel, vol. 30 (1902), p. 195. (Bryozoa, pp. 268-275, 284.)

SMYČKA, F. Novější nálezy v čelechovském devonu. Vestnik Klub. Prostejove, vol. 7, pp. 53-72. (Bryozoa, p. 61.)

THEVENIN, ARMAND. Note sur des Fossiles du Carbonifère infr. du Djebel Bechar (Sud Oranais). Bulletin Société geologique France, vol. 4, no. 6 (Bryozoa, p. 820). THOMAS, H. J. Neue Beiträge zur Kenntniss der devonischen Fauna Argentiniens. Zeitschr. Deutsch.

geol. Ges., vol. 57, no. 11 (Bryozoa, p. 287).

THORNLY, LAURA R. Report on the Polyzoa collected by Professor Herman at Ceylon in 1902. Ceylon pearl oyster fisheries, Report to Col. Gov., pt. 4, suppl. Report no. 26, pp. 107-130, 1 pl.

ULRICH, E. O. Geology and general relations. In Ulrich, E. O., and Smith, W. S. T. Lead, zinc, and fluorspar deposits of western Kentucky. Professional Paper 36, U. S. Geological Survey, pp. 7-105, with plates.

WATERS, A. W. Bryozoa from near Cape Horn. Journal Linnean Society London, Zool., vol. 29, pp. 230-251, pls. 28, 19.

WATERS, A. W. Notes on some Recent Bryozoa in d'Orbigny's Collection. Annals and Magazine Natural History, ser. 7, vol. 15, pp. 1-16, 1 pl.

WILSON, J. HOWARD. The pleistocene formations of Sankaty Head, Nantucket. Journal Geology, Chicago, Illinois, vol. 13, pp. 713-33.

Young, G. W. The chalk area of N. E. Surrey. Proceedings Geological Association, vol. 19, pt. 4, pp. 188-219. (Bryozoa, p. 217.)

ZAHÁLKA, ČENĚK. Pásmo X, kridového utvaru v Pojizeri (Zone X of the Cretaceous of the Iser District of Bohemia). Prag, Sitz-Bereichte Böhmen Gesellschaft Wissenschafte, no. 17, pp. 1-184.

ZIMMER, C. Pectinatella magnifica (Leidy) in der Oder. Zoologischer Anzeiger, vol. 29, pp. 427-428. ZYKOFF, W. Ueber das Plankton des Saisan-Sees. Bryozoa. Zoologischer Anzeiger, vol. 29, no. 15, pp. 477-482, 2 figs.

Annandale, N. Notes on the fresh-water fanna of India, no. 11. Affinities of Hislopia. Journal Proceedings Asiatic Society Bengal, vol. 2, pp. 59-63, 1 fig.

Arstein, C. Plankton in Nord- und Ostsee auf den deutschen Terminfahrten. 1 Teil. (Volumina 1903.) Wissenschaftliche meeresuntersuchungen, Kiel, new ser., vol. 8, Abt. Kiel, pp. I-26 and LIX.

Bassler, R. S. The bryozoan fanna of the Rochester Shale. U. S. Geological Survey, Bulletin 292, 137 pp., 31 pls.

Bassler, R. S. A study of the James types of Ordovician and Silurian Bryozoa (with bibliography). Proceedings U. S. National Museum, vol. 30, pp. 1-66, pls. 1-7.

Bonnevie, Kristine. Physiologische Polyspermie. Archiv Mathematik oy Naturvidenskab, Kristiania, vol. 27, no. 13, 15 pp., 1 pl.

Boule, Marcellin, and Thévenin, Armand. Types du Prodome de paléontologie stratigraphique universelle de D'Orbigny. Annales Paleontologie, Paris, vol. 1, pp. 97-100, 165-72, 193-6.

BRYDONE, R. M. Further notes on the stratigraphy and fauna of the Trimmingham chalk. Geological Magazine, new ser., vol. 3, pp. 13-22, 72-78, 124-131, 289-300.

Bullen, R. A. 11. Notes on some Microzoa and Mollusca from East Crete. *Geological Magazine, dec. 5, vol. 3, no. 8, pp. 345-358, pls. 18, 19.

Calvet, Louis. Note préliminaire sur les Bryozoaires recueillis par les expeditions du *Travailleur* (1881-82) et du *Talisman* (1883). Bulletin Museum Ilistoire Naturelle Paris, pp. 154-166.

Calvet, L. Deuzieme note préliminaire sur les Bryozoaires récoltés par les expéditions du *Travailleur* (1881-82) et du *Talisman* (1883). Bulletin Museum Histoire naturelle, Paris, no. 12, p. 215-223.

Clarke, J. M. Percé, a brief sketch of its geology. Report New York State Museum, vol. 57, Bulletin 80, pp. 134-I71.

Cossman, M. Rectifications de nomenclature (Bryocryptella n. nom for Cryptella Jullein). Revue critique de paleozoologie, vol. 10, pp. 78-131.

Cumings, E. R. The fauna of the Salem limestone of Indiana. 30th Annual Report Indiana Department of Geological and Natural Resources, pp. 1274-1296, pls. 27-40.

DARTON, N. H. Geology of the Bighorn Mountains. U. S. Geological Survey, Professional Paper 51, pp. 1-129.

DEGRANGE-TOUZIN. Le Falun de Saint-Denis. 1le D'Oléron (Charente-Inferieure), Actes Société Linneene de Bordeaux. 61, pp. 17-22.

EVANS, DAVID CLEDLYN. The Ordovician rocks of Western Carmarthenshire. Quarterly Journal Geological Society, London, vol. 62, pp. 597-643.

Foerste, August F. Silurian, Devonian, and Irvine formations of East central Kentucky. Kentucky Geological Survey, Bulletin no. 7, 369 pp., 33 pl.

Gough, Lewis Henry. Plankton collected at Irish light stations in 1904. Fish. Ireland, Scientific Investigations, 1904 (1906), pp. 3-79.

Grabau, A. W. Guide to the geology and paleontology of the Schoharie Valley in eastern New York. New York State Museum, Bulletin No. 92.

Hennig, Anders. Gotlands Silur-Bryozoer 2. Arkiv. för Zoologie. K. Svenska Vetenskapsakademien, Stockholm, vol. 3, no. 10, pp. 1-62, 7 pls.

JOHNSEN, A. Bryozoen aus dem karnischen Fusulinenkalk. Neues Jahrbuch Min. Geol. Pal., vol. 2, Heft 3, pp. 135-160, pls. 10, 11.

Kluge, Hermann. Zoologische Ergebnisse einer Untersuchungsfahrt des deutschen Seefischerei-Vereins nach der Bäreninsel und Westspitzbergen ausgeführt im Sommer 1898 auf S. M. S. Olga. Bearbeitet nach Sammlungen von Dr. Hartlaub... Wissenschaftliche Meeresuntersuchungen, new ser., vol. 8, Abt. Helgoland, p. 31-55, 10 figs.

Köhler, Walther. Süsswasser-bryozoen in geheizten Aquarien. Blätter Aquarienkunde, Magdeburg, vol. 17, p. 312.

Kraepelin, K. Eine Süsswasserbryozoë (Plumatella) aus Java. Mittheilungen naturhistorischen Museum Hamberg, Jahrg. 23, p. 143–146, 3 figs.

Kupelwieser, H. Untersuchungen über den feineren Bau und die Metamorphose des Cyphonautes. Zoologie, vol. 19, Heft 47, 50 pp., 5 pls., 8 figs.

Lang. W. D. A key to the published figures of the Cretaceous forms of the Polyzoan genus Entalophora. Geological Magazine, dec. 5, vol. 3, no. 10, pp. 462-467.

Lang, W. D. The Reptant Eleid Polyzoa. Geological Magazine, dec. 5, vol. 3, no. 11, pp. 60-69,

12 text figs.

Lang, W. D. Bryozoa in Woods, Henry. The Cretaceous fauna of Pondoland, Cape Town. Annals South African Museum, vol. 4, pt. 7, pp. 282-286, pl. 33.

Lemoine, Paul. Études géologiques dans le nord de Madagascar. Annals Hébert, vol. 3.

Lissajous. Toarcien des euvirons de Macon. Macon (reprinted with corrections from Macon, Bulletin société science naturelle, vol. 2, 1902-6).

Lomnicki, A. M. Mszywiol. Rozpiorka wieloksztaltna na raku stawowym. Kosmos Lwów Roczn, vol. 31, pp. 249–256, 4 figs. (Polish.)

Loppens, K. Bryozoaires marins et fluviatiles de la Belgique. Annals Societé Zoologique malacologique Belgique, vol. 41, pp. 286-321, 50 figs.

LOPPENS, K. Bryozoaires nouveau pour la faune belge. Bulletin Société Royale zoologique malacologique Belgique, vol. 40, p. 49.

LOPPENS, K. Contributions à l'étude des Bryozoaires de Belgique. Société zoologique malacologique Belgique, vol. 41, pp. 130-136.

LOPPENS, K. Plumatella repens L. vivant dans l'eau saumatre. Bulletin Société Royale zoologique malacologique Belgique, vol. 40, pp. 49, 50.

Loppens, K. Sur quelques variétés de Membranipora membranacea L. vivant dans l'eau saumatre Annals Biol. lacustre, vol. 1, pp. 40-42, 4 figs.

MERCINI, C. L' Infratius del M. Melbe presso Perugia. Pisa, Proc. verb. Soc. tosc. sc. nat., vol. 15, pp. 49, 50.

MARQUAND, E. D. The Zoophytes (Hydroida and Polyzoa) of Guernsey. (Including records by A. M. Norman.) Guernsey, Transactions Society Natural Science, pp. 164-176.

MAYER, PAUL. Bryozoa und Brachiopoda (Bibliography for 1905). 2 vol. Jahresber., Berlin, pp. 1-3.

Neviani, A. Briozoi viventi e fossili illustrati da Ambrogio Soldani neli' opera Testaceographia ac zoophytographia parva et micoscopia (1789-1798). Bollitino Societiá Geologica Italiana, vol. 25, pp. 765-785.

Neviani, A. Briozoi ritenuti nova de Mitili. Bollettino Societiá Zoologique Italiana, ser. 2, vol. 7 fasc. 4-6, pp. 184-190, 1 fig.

Nichols, A. A new Irish Polyzoon (Hypophorella expansa Ehlers). Irish Naturalist, vol. 15, p. 87. Nordgaard, O. Bryozoa from the second Fram expedition 1898-1902. Report second Norwegian arctic expedition of the Fram, no. 8, pp. 1-44, pls. 1-4.

Nordgaard, O. Die Bryozoen des westlichen Norwegens. Bergen Museum Meeres-fauna von Bergen, pp. 73-112, 2 pls.

NORMAN, A. M. Greenlandic Polyzoa. Annals Magazine Natural History, ser. 7, vol. 17, p. 90-93. OKA, ASAJIRO. Tansui-Kokemushi noichi shin shu. (On a new species of fresh-water Polyzoa.) Döbu-

tsugaku Zasshi, Tokyo, vol. 6, 1908, pp. 307-310.

RETZIUS, GUSTAF. Die spermien der Bryozoen. Biol. Untur., new ser., vol. 13, pp. 45–48, pl. 1, 1 fig. Pace, R. M. On the early stages in the development of Flustrella hispida (Fabricius) and on the existence of a "yolk nucleus" in the egg of this form. Quarterly Journal Microscopical Science, vol. 50, pp. 435–478, 4 pls.

RIDEWOOD, W. G. A new species of Cephalodiscus, from the Cape Selas. Maritime Investigation South

African Department of Agriculture, vol. 4, pp. 173-192, 3 pls., 5 figs.

RÖMER, OTTO. Untersuchungen über die Knospung. Degeneration und Regeneration von einigen marinen ectoprocten Bryozoen. Zeitschrift wissenschaftliche zoologie, vol. 84, pp. 446–478, 2 pls. Sacco, Federico. Les étages et les faunas du bassin tértiaire du Piemont. Boll. geol. soc. France,

vol. 5, pp. 893-916.

Sacco, Federico. La questione comiocenica dell'Appenino. Roma, Bollittino Sociétiá Géologica

Italiana, vol. 25, pt. 1, pp. 65-127.

Schefotieff, A. Die Pterobranchier, anatomische und histologische, untersuehunger über Rhabdopleura normanii Allmann und Cephalodiscus dodecalophus M. Int. 1 Teil . . . Zoologische Jahr. Abt. Anat., vol. 23, pp. 463-534, 9 pls.

Seeliger, Oswalp. Ueber die Larven und Verwandtschaftsbeziehungen der Bryozoen. Zeitschrift wissenschaftliche Zoologie, vol. 84, pp. 1-78, 4 pls., 4 figs.

Seely, H. M. Beekmantown and Chazy formations in the Champlain Valley. Report Geological Survey Vermont, vol. 5, pp. 174-187 with plates.

Siemiradzki, Jósef. Monografia warstw paleozoicznych Podola. Kraków. Bull. Intern. Acad., pp. 23-32.

Stemtradzki, Jósef. Die Paleozoischen Gebilde Podoliens. Wien, Beitrg. Pal. Geol., Vest-Ung., vol. 19, pp. 173-286.

Sildermann, Samuel. Untersuchungen über den feineren Bau von Alcyonidium mytili. Archives Naturges. Jahrg. 72, vol. I, p. 265-310, 2 pls.

Thevenin, A. Types du Prodome de Paleontologie stratigraphique universelle de D'Orbigny. Annales de paleontologie, vol. 1, pp. 1-7, pls. 8, (fig. 1) 9, (fig. 11), 12 (fig. 111).

ULRICH, E. O. Pleistocene Bryozoa. Maryland Geological Survey, Pliocene-Pleistocene, pp. 210-212, with figs.

VADISZ, M. ELEMÉR. Über die obermediterrane Fauna von Budapest. Rákos, Földtaui Közlöny Budapest, vol. 36, pp. 256–283, 323–351, with plates.

Vaney, C., and Conte, A. Recherches sur le *Rhabdopleura normanii* Allman. Anatomie, bourgeonnement et affinities. Revue suisse Zoologique, vol. 14, pp. 143, 183, 4 pls.

WATERS, A. W. Bryozoa from Chatham Island and d'Urville Island, New Zealand . . . Annals Magazine Natural History, ser. 7, vol. 17, pp. 12-23, pl. 1.

Weeks, Fred Boughton. Bibliography and index of North American geology, paleontology, petrology, and mineralogy for the years 1901–1905, inclusive. Bulletin U. S. Geological Survey, no. 301.

Weltner, W. Pectinatella magnifica (Leidy) bei Berlin. Archives Naturges, Jahrg. 72, vol. 1, pp. 259-264, 3 figs.

WHITE, HAROLD J., OSBORNE and TREACHER, LLEWELLYN. The phosphatic chalk of Winterbourne and Boxford (Berkshire). Quarterly Journal Geological Society, London, vol. 62, pp. 499-522.

Whiteaves, J. F. 2. The fossils of the Silurian (Upper Silurian) rocks of Keewatin, Manitoba, the northeastern shore of Lake Winnipegosis, and the lower Saskatchewan River. Ottawa, Geological Survey Canada, Pal. Fossils, vol. 3, pp. 243-98.

Wilcox, A. W. Locomotion in young colonies of Pectinatella magnifica. Biological Bulletin of the Marine Biological Laboratory, Woods Hole, Massachusetts, vol. 11, pp. 245-252, 2 pls.

Woods, Henry. The Cretaceous fauna of Pondoland (the Polyzoa by W. D. Lang). Cape Town, Annals South African Museum, vol. 4, pp. 275-350.

I907.

ANNANDALE, N. The fauna of brackish ponds at Port Canning, Lower Bengal, pt. 6. Observations on the Polyzoa, with further notes on the ponds. Records of the Indian Museum, vol. I, pp. 197-205, 3 figs.

Annandale, N. The fauna of brackish ponds at Port Canning, Lower Bengal, pt. 1. Introduction and preliminary account of the fauna. Record Indian Museum, Calcutta, vol. 1, pt. 1, pp. 35-43. Annandale, N. Further notes on a Polyzoon from the Himalayas. Records of the Indian Museum,

vol. 1, pp. 145-148, figs. 3.

Annandale, N. Notes on the fresh-water fauna of India, no. 12. The Polyzoa occurring in Indian fresh and brackish pools. Journal and Proceedings Asiatic Society Bengal, vol. 3, pp. 83-93, 1 pl. Annandale, N. Statoblasts from the surface of a Himalayan pond. Records of the Indian Museum,

vol. 1, p. 177.

Arnold, Ralph. Geology and oil resources of the Summerland district, Santa Barbara County, California. (With notes and illustrations of Mollusca from the Eocene, Miocene, and Pliocene and Bryozoa from the Pliocene.) Bulletin U. S. Geological Survey, no. 321, pp. 1-93, pl.

Apstein, C. Das Plancton im Colombo-See auf Ceylon. Sammelausbeute von A. Borgert, 1904–1905 Zoologische Jahrbüch. Abtheilungen Systematik, vol. 25, pp. 201–244.

BATHER, F. A. Nathorst's use of collodion imprints in the study of fossil plants. Geological magazine, London, ser. 2, dec. 5, vol. 4, pp. 437-440.

Beauchamp, P. de. Quelques observation sur les conditions d'existence des êtres dans la Baie de Saint-Jean-de-Luz et sur la côte avoisinante. Archives zoologie expérimental et général, ser. 4, vol. 7, Notes No. I, pp. 4-16.

Bogolerow, M. Wachstum und Leben der Kolonien der Tendra zostericola au den Gläsern der Aqua-

rien. Zoologischer Anzeiger, vol. 32, pp. 305-316, 7 f.gs.

Bonnevie, Kristine. Untersuchungen über Keimzellen, 11. Physiologische Polyspermie bei Bryozoen. Jenaische Zeitschrift Naturwissenschaft, vol. 42, pp. 567-598, 4 pls.

Boule, Marcelin, and Thèvenin, Armand. Types du Prodome de Paléontologie Stratigraphique Universelle de d'Orbigny (Contd.). Annales Paléontologie Paris, vol. 2, pt. 4, pp. 161-172=25-36,

pls. 23, 24, and 10.

Brehm, V. Über das Vorkommen von Diaptomus tatricus Wierz. in den Ostalpen und über Diaptomus kupelwieseri nov. sp. Zugleich eine Mitteilung über die neue biologische Station in Lunz Zoologischer Anzeiger, vol. 31, pp. 319-328.

Canu, F. Les Bryozoaires fossiles des Terrains du Sud-Ouest dela France. 1. Aquitanien. Bulletin Société Géologique France, ser. 4, vol. 6, pp. 510-518, pl. 12, 13.

Calvet, Louis. Bryozoaires (of the *Travailleur* and *Talisman* expeditions). Expédition scientifique *Travailleur* et *Talisman*, 1906, pp. 355–495, pls. 26–30.

Снадwick, H. C. Report on the marine biological station at Port Erin. Liverpool Proceedings Transactions Biological Society, vol. 21, pp. 28-38.

CRIMENKOW, W. Zur Frage über den geologischen Bau der Umgebung von Chwalynsk und über die Kreide-Ablagerungen im Wolgagebiet des nördlichen Teils des Gouv. Saratow. Annuaire geologique et minèralogique de la Russie Novo Aleksandrija, vol. 9, pp. 115–130.

CLARKE, JOHN M. Some new Devonic fossils. Bulletin New York State Museum, no. 107, pp. 153-

291, with text figs.

Davies, A. Morley. Collodion as a preservative for fossils. Geological Magazine, London, ser. 2, dec. 5, vol. 4, pp. 524-525.

Dendy, Arthur. On the occurrence of Fredericella sultana in New Zealand. Transactions New Zealand Institute, vol. 39, pp. 221-222.

Destinez, P. Contribution a la faune du calcaire carbonifere. Liége, Annales Société géologique Belgique, vol. 34 (1906–1907) (B 97–B 100).

Destinez, P. Quatrième note sur la faune du calcaire noir (V 1 a) de Petit-Modave. Liége, Annales Société géologique Belgique, vol. 34 (B 62–B 64) (B 65–B 67).

Dollfus, Gustave F. Faune malacologique du miocène supériur de Beaulieu (Mayenne). (Étage redonien.) Paris, Comptes Rendus Association française advancement science, vol. 35 (Lyon, 1906), pt. 2, pp. 304-315.

Dublin, L. 1. The history of the germ cells in Pedicellina americana. Annals New York Academy Science, vol. 17, p. 583.

FARQUHAR, H. Note on the bipolarity of littoral marine fauna. Wellington, Transactions New Zealand Institution, vol. 39, pp. 131-135.

FILLIOZAT, M. Bryozoaires Crétacés de Vendome. Bulletin Société Géologique France, ser. 4, vol. 7, pp. 391–399, pl. 13, 14.

Fucini, A. Ancora sopra l'età del marmo giallo di Siena. Pisa, Atti societa toscana scienze naturali, vol. 23, pp. 3-7.

GIRTY, G. H. Descriptions of new species of Upper Paleozoic fossils from China. Proceedings U. S. National Museum, vol. 33, p. 37-48.

Gornon, C. H. Mississippian (Lower Carboniferous) formations in the Rio Grande Valley, New Mexico. American Journal Science, ser. 4, vol. 24, pp. 58-64.

GREGORY, J. W. The rotiform Bryozoa of the Isle of Wight. Geological Magazine, dec. 5, vol. 4, no. 10, pp. 442-443.

Grabau, A. W., and Shimer, H. W. North American index fossil. Bryozoa. School of Mines Quarterly, vol. 28, no. 1, Bryozoa pp. 20-83, figs. 176-217.

HAYDEN, H. H. The stratigraphical position of the Gangamopteris beds of Kashmir. Record Geological Survey India, Calcutta, vol. 36, pp. 23-39, pls. 4-9.

HIND, WHEELTON. Life zones in British Carboniferous rocks. Part 2, The fossils of the Millstone grits and Pendleside series. Naturalist, London, pp. 17-23.

Joly, Henry. Les fossiles du Jurassique de la Belgique avec description stratigraphique de chaque étage. Mémoires Museum Belgique, Bruxelles vol. 5, pp. 1-156, pls. 1-6.

JONKER, H. G. Lijst van Geschriften welke handelen over of van belang zijn voor de Geologie van Nederland (1734-1906). Amsterdam, Verhandlungen Koninklijke akademie van wetenschappen, 2d section, vol. 13, no. 2, pp. 7 and 154.

JUKES-BROWNE, A. J., and Else, W. J. A list of the type fossils and figured specimens in the Museum of the Torquay Natural History Society, Plymouth. Report Devon. Association, ser. 2, vol. 9,

pp. 399-409.

KAMMERER, PAUL. Über Schlammkulturen. Archiv Hydrobiologie, Stuttgart, vol. 2, pp. 500-526. KAMMERER, PAUL. Wiedererweckung kleiner Tiere und Pflanzen aus getrocknetem Schlamm (reprint of preceding). Blätter Aquarienkunde. Magdeburg, 18, pt. 23, pp. 227-228; pt. 24, pp. 235-237; pt. 25, pp. 243-248, 3 figs.; pt. 26, pp. 258-259.

Karakasch, N. S. Le crétacé inférieur de la Crimée et sa faune. (Russe) St. Petersbourg, Travaux Société naturelle Sect. géologique, vol. 32, 5, pp. 1-442, 454-482; Rés. franc., pp. 443-453, 28 pls. Kellner, K. Bericht über die Embryologie von Oikopleura. Zoologischer Anzeiger, vol. 31, pp.

653-654, 3 figs.

Kittl, Ernst. Die Triasfossilien vom Heureka Sund. Kristiania, Report of the Second Norwegian Arctic Expedition in the Fram, 1898–1902, no. 7, pp. 1–44, 3 pls.

Kluge, H. Beiträge zur Kenntnis der Bryozoen des Weissen Meeres. St. Petersburg, Bulletin Academie Science, ser. 6, vol. 1, pp. 658-659 (Russian).

Kluge, H. Zur Kenntnis der Bryozoen von West-Grönland. St. Petersburg, Bulletin Academie Science, ser. 6, vol. 1, p. 703.

KLUGE, HERMANN. Kritische Erörterungen zu den bryozoologischen Arbeiten von K. Chworostansky. Archives Naturgeschichk, Berlin, vol. 73, pt. 1, pp. 181–204.

Kohler, W. Latentzustände als Trutzmittel gegan die Unbilden des Klimas (Contd). Blätter Aquarienkunde, Magdeburg, vol. 18, pt. 42, pp. 417-418.

LANG, W. D. The evolution of Stomatopora dichotomoides. Geological Magazine, dec. 5, vol. 4, no. 511, pp. 20-23, text figs. 6.

Lang, W. D. A tabular view of the Cretaceous Polyzoa of the family Idmoniidae. Geological Magazine, new ser., dec. 5, vol. 4, no. 513, pp. 122-132.

Lemoine, Paul. Les variations de facies dans les terrains sédimentaires de Madagascar. Paris, Bulletin Société géologique, ser. 4, vol. 8, pp. 30-41.

Le Roux, Marc. Recherches biologiques sur le lac d'Annecy. Annales Biol. lacustre, Bruxelles, vol. 2, pp. 220-387.

Levinsen, G. M. R. Sur la regeneration totale des Bryozoaires. Oversigtover del glgl. Danskl. videnskabernes selskavs fornandlinger, no. 4, pp. 151, 159, 1 pl.

Loppens, K. Sur les caracterès Distinctifs entre Alcyconidium gelatinosum et Alc. hirsutum. Annals Société Zoologique malacologique Belgique, vol. 42, pp. 169–174.

Lühe, M. Die Bryozoen Ostpreussens. Schriften. physikalisch-ökonomischen Gesellschaft Königsberg, Jahrg. 47, pp. 281-285.

Lindiger, L. Ein neuer Fundort von Cristatella mucedo Cuv. Abhandlungerndes naturhistorishen Gesellschaff zu Nurnberg, vol. 15, p. 84.

MATZDORFF, CARL. (Jahresbericht über) Bryozoa für 1904. Archives Naturgeshichte, Berlin 67, vol. 2, Heft 3, 1901 (1907), pp. 597-621.

Nelli, Bindo. Il Miocene del Monte Titano nella Republica di S. Marino. Roma, Bollettino Sociétà Géologica Italiana, vol. 26, pt. 2, pp. 239-322, pls. 8-10.

Neviani, Antonio. Di un libro poco noto sugli Zoofite a Litofiti del Mediterraneo dell' abate Francesco Maratti, Pt. I (Briozoi). Bollettino Societiá Zoologique Italiana, ser. 2, vol. 8, pp. 102-118.

NEVIANI, ANTONIO. I primo studi anatomici sui briozoi. Atti Congr. Natural. italiana, 1906, pp. 786-788.

NICHOLS, A. R. Contributions to the natural history of Lambay. Polyzoa. Irish Naturalist, Dublin, vol. 16, Nos. 1-2, pp. 82-83.

Nordgaard, O. Bidrag til faunaens histoire i Trondhjemsfjorden. Kgl. Norse Videnskabors Selskabs Skrifter, No. 7, 43 pp., figs.

Nordgaard, O. Bryozoch von dem norweigischen Fischereidampfer "Michael Sars" in den Jahren 1900-1904, gesammelt. Bergens Museum Aarbok, no. 2, 20 pp., 1 pl.

NORMAN, A. M. On some British Polyzoa. Annals Magazine Natural History, ser. 7, vol. 20, pp. 207-212, 1 pl., 1 fig.

OKA, ASAJIRO. Eine dritte art von Pectinatella. Zoologischer Anzeiger, vol. 31, pp. 716-718, 3 figs. Oka, Asajiro. Zur Kenntnis der Süsswasser-Bryozochfauna von Japan. Annotationes Zoologicae Japonenses, vol. 6, pp. 117-123, 3 figs.

PACE, S. On an improved system of recording for use in Faunistic work. Zoologischer Anzeiger, Leipzig, vol. 32, pp. 385-391.

Poche, Franz, Einige Ergänzungen zu den Indices neuer Gattungs und Untergättungsnamen des Zoological Record (für 1901–1905). Zoologischer Anzeiger, Leipzig, vol. 31, pp. 705–707, vols, 37-42.

REED, F. R. COWPER. Sedgwick Museum notes. New fossils from Haverfordwest, 7. Geological Magazine, new ser. 5, vol. 4, p. 208-211, 1 pl.

RICHARD, J. Campagne scientifique de la Princesse-Alice (1907). Liste des Stations. Monaco, Bulletin Institut Ocean, no. 106 (1-11), map.

RICHARDSON, LINDSALL. The Inferior Oolite and Contiguous Deposits of the Bath-Doulting District (with Appendices (1) by S. S. Buckman, (2) by L. R. Richardson and J. L. Walker, (3) by L. Richardson and J. L. Walker, (4) by L. Richardson and J. L. Walker, (5) by L. Richardson and J. L. Walker, (6) by L. Richardson and J. L. Walker, (7) by L. Richardson and J. Walker, (8) by L. Richardson and J. Walker, (8) by L. Richardson and (8) by Richardson and (8) by Richardson and (8) by Richardson and (8 ardson). London, Quarterly Journal Geological Society, vol. 63, pt. 252, 1907, pp. 383-436, pls. 28-29, 4 pls.

Ridewood, W. G. Pterobranchia. Cephalodiscus. National Antarctic Expedition, vol. 2, no. 5, 67 pp., 7 pls., 17 figs.

Ridewood, W. G. On the development of the plumes in buds of Cephalodiscus. Quarterly Journal Microscopical Society, vol. 51, pp. 221-252, 11 figs.

ROULE, LOUIS. Considérations sur la faune marine du Port de Bonifacio (Bryozoa determined by L. Calvet). Bulletin société zoologique, vol. 32, pp. 40-45.

Rousselet, Charles. Zoological results of the third Tanyanika Expedition conducted by Dr. W. A. Cunnington, 1904-5. Report on the Polyzoa. Proceedings Zoological Society London, 1907, pp. 250-257, 2 pls.

Skorikov, A. Quelques faits concernant la biologie d'un étang situé dans le Jardin de la Tauride St. Pétersbourg. St. Petersbourg, Bulletin Acadamie Science, No. 10, ser. 6, 7, pp. 119-126 (Rus-

DE STEFANI, C. I terreni e le acque cloro-sodiche della salute in Livorno. Pisa, Atti Societa toscana scienze natural, vol. 23, pp. 88-124.

Steuer, Adolf. Nenere Arbeiten über Plankton mit besonderer Berücksichtigung des Zooplanktons. Wien, Verhandlungen Zoologische Botanischen Gesellschaft, vol. 57, pp. 40, 62.

STUCKENBERG, A. Die Fauna der obercarbonischen Suite des Wolgadurchbruches bei Samara. St. Petersbourg, Mémoires comité géologique, ser. 2, vol. 23, 1905 (1907) (xix+144 pp.), 13 pls. (Russian with German summary, 111-144 pp.)

THORNELY, LAURA R. Report on the Marine Polyzoa in the collection of the Indian Museum. Records Indian Museum, vol. 1, pp. 179-196, 8 figs.

ULRICH and BASSLER. See WELLER.

Vadasz, M. Elemér. A ribicei felsőmediterrán korszaki korallpad faunajaról. Über die övermediterrane Koralleubank von Rivica. Földtani Közlöny, Geologische Mittheilungen, Budapest, vol. 37, pts. 9-11. (Hungarian, pp. 368-373; German, pp. 420-425.)

Verrill, Addison E. The Bermuda Islands. Pt. 4, Geology and Paleontology, and pt. 5, An account of the Coral Reefs. New Haven, Connecticut, Transactions Academy Arts and Science, vol. 12, pp. 45-348, pls. 16-40.

Vogl, Viktor. Adatok a fóti álsómediterran ismeretéhez. Beiträge zur Kenntnis des Untermediterrans von Fót. Földtani Közlony, Budapest, vol. 37, pt. 1-8 (Hungarian, pp. 243-246; German, pp. 303-307.)

Walton, H. G. Notes on Hislopia lacustris Carter. Records Indian Museum, vol. 1, p. 177-178. Waters, A. W. Tubucellaria, its species and ovicells. Journal Linnean Society London, vol. 30, pp. 126-133, 2 pls.

- Weller, Stuart. A report on the Cretaceous Paleontology of New Jersey (new species Bryozoa by Ulrich and Bassler). Geological Survey of New Jersey, Paleontology 4, text pp. 1-187, plates.
- Wesenberg-Lund, C. On the occurrence of Fredericella sultana Blumenb. and Paludicella Ehrenbergii van Bened. in Greenland. Meldelelser om Grönland Köbenhavn, vol. 34, pp. 63-75.
- Wiren, A. Zoologien i Uppsala 1882-1907. En aterblick Zool. Stud. tillägn. T. Tullberg (Naturvet. Students ällsk. Uppsala), pp. 1-41.
- Ziegler, H. E., et alii. Zoologisches Wörterbuch. Erklärung der zoologischen Fachausdrücke . . . pt. 1 (A. F.) xvi+208 pp.
- ZIMMERMAN, HANS. Tierwelt am Strande der blauen Adria. Eine Naturwissenschaftliche Skizze aur Erlangung einer Ubersicht der Fauna von Rovigno (Istrien), sowie zur Einführung in die Sammeltechnik. Zs. Natw., Stuttgart, ser. 5, vol. 16, pp. 293–321.
- ZSCHOKKE, F. Ubersichtüber die Tiefenfauna des Vierwaldstättersees. Archive Hydrobiologie Stuttgart, vol. 2 (1907), 1906, pp. 1-8.

1908.

- Annandale, N. Corrections as to the identity of Indian Phylactolaemata. Records Indian Museum vol. 2, p. 110.
- Annandale, N. The fauna of brackish ponds at Port Canning, Lower Bengal. Pt 7. Further observations on the Polyzoa, with the descriptions of a new genus of Entoprocta. Records of the Indian Museum, vol. 2, pp. 11-19, 7 figs.
- Annandale, N. Three Indian Phylactolaemata. Records of the Indian Museum, Calcutta, vol. 2, pt. 2, pp. 169-174.
- Beutler, Karl. Beitrag zur Kenntniss der cyclostomen Bryozoen der alteren Tertiarschichten des südlichen Bayern. Paleontographica, vol. 54, Lief. 5, pp. 205–250, pls. 23, 24, and 7 text figs.
- Braem, F. Die geschlechtliche Entwicklung von Fredericella sultana nebst Beobachtungen über die weitere Lebensgeschichte der Kolonien. Zoologica, Heft 52, pp. 1-37, 7 pl. 1 fig.
- Braem, F. Die Spermatozoen von Paludicella und Triticella. Zoologischer Anzeiger, vol 33, pp. 280-281.
- Braem, F. Die Spermatozoen der Süssawasser-Bryozoen. Zoological Anzeiger, vol. 32, pp. 671-773, 2 figs.
- Braem, F. Ueber die Umwandlung plasmatischer Granula zu halbmondförmigen Körpern. Anatomische Anzeiger, vol. 33, pp. 360-364, 1 fig.
- Canu, F. Iconographie des Bryozoaires fossiles de l'Argentine (Première partie). Anales del Museo Nacional de Buenos Aires, vol. 17 (ser. 3, vol. 10), pp. 245-341, 13 pls.
- Canu, F. Les Bryozoaires fossiles des terrains du Sud-Ouest de la France. II. Lutécien. Bulletin Société Géologique France, ser. 4, vol. 8, pp. 382, 390, pls. 6, 7.
- CHATWIN, CHARLES P. and WITHERS, THOMAS H. The zones of the chalk in the Thames Valley between Goring and Shiplake. With an appendix by George J. Hinde. London, Quarterly Journal Geological Society, vol. 64, pp. 390—420.
- Chirica, Const. Note asupra Bryozoairilor din Romania. Mem. Assoc. romana Inaintarea Respand St. 2, pp. 468–474.
- CLARKE, JOHN M. Early Devonic history of New York and eastern North America. New York State Museum, Mem. 9, no. 1, pp. 1-366, with plates.
- Couffon, O. Le Miocène en Anjou (Supplément). Bulletin de la société études scientifiques d'Angers (Maine-et-Loire), vol. 37, pp. 49-58.
- CUMINGS, E. R. The stratigraphy and paleontology of the Ordovician rocks of Indiana. 32d Annual Report Department Geology Natural Resources Indiana, pp. 605-1190.
- Czwiklitzer, R. Die Anatomie der Larve von *Pedicellina cchinata*. Arbeiten zoologischen Instituten. Wien, vol. 17, pp. 157-186, pl., 2 figs.
- Filliozat, Marius. Nouveaux Bryozaires cheilostomes de la Craie. Bulletin Société Géologique France, ser. 4, vol. 8, pp. 554-560, 1 pl.
- Filliozat, Marius. Sur les Synchronismes Crétacés par les bryozoaires la Craie de Vendome. Bulletin Société Archéologique, Scientifique et Litteraire des Vendomois, vol. 47, pp. 255-257.
- GIRTY, G. H. The Guadalupian fauna. U. S. Geological Survey Professional Paper 58, pp. 1-649, with pls.

GREIG, M. J. Invertébres du fond. In Duc d'Orleans, Croisière Oceanographique dans la mer du Grönland. Bruxelles, pp. 503-567.

Hennig, Anders. Gotlands Silur-Bryozoer. 3. Arkiv. för Zoologie K. Svenska Vetenskapsakademien Stockholm, vol. 41, no. 21, pp. I-64, pls. 1-7.

HOPKINSON, JOHN. Dates of publication of the separate parts of Gmelin's edition (13th) of the "Systema Naturae" of Linnaeus. London, Proceedings Zoological Society, 1907, pp. 1035-1037.

Kluge, H. Beiträge zur Kenntnis der Bryozoen des Weissen Meeres. Annals Museum Zoology, St. Petersbourg, vol. 12, pp. 215-540, 2 figs.

Kluge, H. Zur Kenntnis der Bryozoen von West-Grönland. Annals Museum Zoology, St. Petersbourg, vol. 12, pp. 546-554.

LANG, W. D. Polyzoa and Anthozoa from the Upper Cretaceous limestone of Needs Camp, Buffalo River. Annals South African Museum, vol. 7, pp. 1-11, 1 pl.

Lee, G. W. Notes on fossils from Prince Charles Foreland. Proceedings Royal Physical Society Edin-

burgh, vol. 17, no. 4, pp. 149-166, pl. viii.

LEVANDER, K. M. Zur Kenntniss der Verbreitung der Süsswasser Bryozoen in Finland. Meddel. Société Fauna and Flora fennica, Häft 34, pp. 97-106.

LOPPENS, K. Les Bryozoaires d'eau douce. Annales Biologie Lacustre, vol. 3, pp. 141-183, 31 pls.

MAPLESTONE, C. M. Further descriptions of the Tertiary Polyzoa of Victoria, pt. 10. Proceedings Royal Society Victoria, vol. 21, new ser., pt. 1, pp. 233-239, pls. 7, 8.

MASTERMAN, A. S. On the Diplochorda. Pt. 5. Certain points in the structure of Tornaria. Quarterly Journal Microscopical Science, vol. 52, pp. 481-493, 1 pl.

MATZDORFF, CARL. Bryozoa für 1905. (Jahresbericht) Archives Naturges schichte. Berlin, 69, vol. 2, Heft 3, 1906 (1908), XIII, pp. 1-8.

Montgomery, T. H. On the Morphology of excretory organs of Metazoa. Proceedings American Philosophical Society, vol. 47 (Polyzoa, pp. 553, 603, 604, 606).

Mourlon, Michel. Le Calcaire Carhonifère et les dépôts postprimaires qui le ecouverent dans la vallée de l'Escaut, entre Tournai et Antoine. Bruxelles, Bulletin Société géologique, vol. 22, Pr.-verb. pp. 89-105.

NICHOLS, A. R. Bryozoa (of Duhlin district). Handbook British Association Advancement of Science, pp. 202-204.

OKA, ASAJIRO. Ueber eine neue Gattung von Süsswasserbryozoen (Stephanella). Annotationes Zoologicae Japonenses, vol. 6, pp. 277-285, 1 pl.

REED, F. R. COWPER. The Devonian faunas of the northern Shan States. Paleontologia Indica. Memoirs Geological Survey India, Calcutta, vol. 2, no. 5, pp. 1-183, pls. 1-20.

REED, F. R., COWPER and REYNOLDS, H. S. Silurian fossils from certain localities in the Tortworth Inlier. Bristol, Report Naturalists Society, ser. 4, vol. 2, pp. 32-40.

RICHARDSON, L. On the Phyllis collection of Inferior-Oolite fossils from Doulting. Geological Magazine London, ser. 2, dec. 5, vol. 5, pp. 509-517.

ROBERTSON, ALICE. The incrusting Chilostomatous Bryozoa of the west coast of North America. University of California Publications, Zoology, vol. 4, no. 5, pp. 253-344, pls. 14-24.

Rowe, Arthur W. The zones of the white chalk of the English coast. V. The Isle of Wight. The maps and descriptive appendix by C. Davies Sherborn . . . and a note on certain silicious nodules by G. T. Prior. London, Proceedings Geological Association, vol. 20, pp. 209-35, 2, pls. 8, 23, maps.

Schimkewitsch, W. Die Methorisis als embryologischer Prinzip. Zoologischer Anzeiger, vol. 33, pp. 585-598.

SMITH, JOSEPH. The Polyzoa—their place in nature, with notes on some peculiarities in structure. Manchester, Report Microscopical Society, 1907 (1908), pp. 64-79, pls. 2-3.

SOLLAS, IGERNA B. G. A new fresh-water Polyzoon from South Africa. Annals Magazine Natural History, ser. 8, vol. 2, pp. 264-273, 8 figs.

STEINMANN, GUSTAV, and WILCKENS, OTTO. Kreide und Tertiar fossilien aus den Magellans ländern, gesammelt von der Schwedischen Expedition 1895-1897. Arkiv für Zoologie, Stockholm, vol. 4, no. 6, 118 pp.

STIASNY, G. Beobachtungen über die marine Fauna des Triester Golfes im Jahre 1907. Zoologischer Anzeiger, Leipzig, vol. 32, pp. 748-752.

Taeger, Heinrich. Die geologischen Verhältnisse des Vertesgebirges. Mitt. Jahrb. Ungar, Geol. Ans., vol. 17, pp. 1-275, pls. 1-12.

Tuéel, IlJalmar. Om utvecklingen af Sveriges zoologiska hafsstation Kristineberg och om djurlifvet i angränsande haf och fjordar. Arkiv. Zoologie, Stockholm, vol. 4, no. 5, pp. 1-136, 5 pls., 3 maps.

Thiébaud, Maurice. Contribution à la biologie du Lac de Saint-Blaise. Annales Biologie Lacustre, Bruxelles, vol. 3, pp. 54-140, pls. 1-4.

THOMPSON, J. Zoophytes in the Humber. Naturalist, London, p. 454.

Waters, A. M. A subfossil Polyzoon from Calcutta. Record Indian Museum, vol. 2, p. 109-110.

Young, George William. The chalk area of western Surrey, London, Proceedings Geological Association, vol. 20, pp. 422-455.

Zykoff, W. Das Plancton des Flusses Irtisch und seiner Nebenflüsse Bukon u. Tabol, Zoologischer Anzeiger, Leipzig, vol. 33, pp. 103-112.

1909.

Annandale, N. Materials for a revision of the Phylactolaematous Polyzoa of India. Records of the Indian Museum, vol. 5, pp. 37-57.

Annandale, N. A new species of Fredericella from Indian Lakes. Records of the Indian Museum, vol. 3, pp. 373-374.

Annandale, N. Preliminary note on a new genus of Phylactolaematous Polyzoa. Records of the Indian Museum, vol. 3, pp. 279-280, 1 fig.

Bassler, R. S. The Netterroth collection of Invertebrate fossils. Miscellaneous Collections, Smithsonian Institution, vol. 52, pp. 121-152.

Bedor, M. Sur la faune de l'Archipel Malais (Résumé). Revue Suisse Zoologique, vol. 17 pp. 143-169.

Billard, Armand. Révision des espèces types d'hydroïdes de la collection Lamouroux. Annales des Sciences naturelles, ser. 9, vol. 9, pp. 307-334.

BLOOMFIELD, E. N. Annual notes on the local fauna, flora, etc., Hastings and E. Sussex. Nat., vol. 1, pp. 186-191.

Brydone, R. M. Notes on new or imperfectly known chalk Brydona (Polyzoa). Geological Magazine, dec. 5, vol. 6, pp. 337-339, 398-400, 3 pls., 1 fig.

Calver, Louis. Bryozoaires. Expédition Antarctique Française (1903-1905) commandée par Dr. Jean Charcot. Sciences Naturelles: documents Scientifiques, Paris, pp. 1-49, pls. 1-111.

Calvet, Louis. Voyage de MM. M. Bedot et C. Pictet dans l'Archipelmalais. Bryozoaires d'Amboine. Note sur Bugula dentata (Lamouroux) et Retepora denticulata Busk. Revue Suisse Zoologie Genève, vol. 14, pp. 617-621, 1 pl.

Canu, F. Étude sur la répartition géologique des Bryozoaires. Comptes Rendus Academie Science, Paris, vol. 148, pp. 532-534.

Canu, F. Bryozoaires fossiles des Terrains du Sud-Ouest de la France. III, Burdigalien; IV, Helvetien. Bulletin Société Géologique de France, ser. 4, vol. 9, pp. 442-458, pl. 15-18.

Canu, F. Les Bryozoaires fossiles du Miocène moyen de Marse-Matrouh en Marmarique. Comptes Rendus Academie Science, Paris, vol. 148, pp. 959-960.

CLARKE, JOHN M. Early Devonic history of New York and eastern North America. Memoirs New York State Museum, no. 9, pt. 2 (=Report 62, 4), pp. I-250, pls. 1-34.

Combes, Paul. La faune halolimuique Africaine. Cosmos, Paris, vol. 61, pp. 705-707.

DAVIDSON, W. C. The fresh-water Polyzoan Cristella mucedo from Kilmacolm. Glasgow Naturalist, vol. 2, pp. 15, 16.

Goddard, E. J. Australian fresh-water Polyzoa, pt. 1. Proceedings Linnean Society New South Wales, vol. 34, pp. 487–496, 1 pl.

Gregory, J. W. Catalogue fossil Bryozoa in Department Geology British Museum. Cretaceous, vol. 2, pp. 346, 9 pls.

GREGORY, J. W. New species of Cretaceous Bryozoa. Geological Magazine, new ser., dec. 5, vol. 6, p. 61-66.

HARMER, S. F. (Avicularia and vibracula.) Rept. 78th Meeting British Association Advancement of Science, 1908, pp. 715-731. (Presidential address.) Hartmeyer, Robert. Bryozoen, Moostierchen. In: Die Süsswasser fauna Deutschlands, hrsg. v. Brauer, Heft. 19, Jena, pp. 49-58.

HEATH, ALICE. Notes on Marine Polyzoa collected during the year June, 1908, to May, 1909. Journal

Torquay Natural History Society, vol. 1, pp. 15-16.

Henneguy, F. Sur un épithélium à fibres muscularies striées. Comptes Rendus Academie Science Paris, vol. 148, pp. 134-138, 3 figs. Un épithélium à fibres musculaires striées. Comptes Rendus Ass. Anat. Reun., vol. 11, pp. 301-302.

HINDE, GEORGE J., and Gossling, Frank. Fossils from the chalk, exposed in a road-trench, near Croham Hurst, South Croydon. Croydon Proceedings Microscopical Natural History Cl. 1907-8

(1909), pp. 183-184.

Knipowitsch, N. M. Rapport sur les collections zoologiques faites pour le Musée Zoologique de l'Académie Imp. des Sciences par N. M. Knipowitsch et S. A. Pavlovič dans la mer Baltique durant l'été 1908. Ann. mus. zool., vol. 14, pp. 0131-0245 avec 1 carte.

LEE, G. W. A Carboniferous fauna from Nowaja Semlja, collected by Dr. W. S. Bruce. Edinburgh Transactions Royal Society, vol. 47, pp. 143-186, pls. 1 and 11.

Leighton, Douglas, (in) Morey's guide to the natural history of the Isle of Wight. London, William Wesley, pp. 235-239.

Levinsen, G. M. A. Morphological and systematic studies on the Cheilostomatous Bryozoa, pp. 1-vii, 1-431, 27 lithographic pls., 6 text figs.

MAPLESTONE, C. M. Polyzoa from the Gilbert Islands. Proceedings Royal Society Victoria, n.s., vol. 21, pp. 410-419, 3 pls.

MAPLESTONE, C. M. The results of deep sea investigations in the Tasman sea. The exposition of H. M. C. S. *Miner*. Polyzoa. Records of the Australian Museum, vol. 7, no. 4, pp. 267-273, pls. 7-78.

Matzdorff, Carl. Bryozoa fur 1906. (Jahresbericht) Archives Naturges., vol. 73, p. 2, Heft 3, 1907 (1909), XIII (1-15).

Moderg, Joh. Chr., and Grönwall, K. A. Om Fyledalens gotlandium. (On the gotlandium of Fyledalen, Skane.) Lund Univ. Ärsskr., new ser., 5 Afd., vol. 2, no. 1 (=Fysiogr. Sällsk. Handl., new ser., vol. 20, no. 1, pp. i-x, 1-S6, 1 map, 6 pls.

Nilus, G. Notiz über Loxosoma murmanica und Loxosoma brumpti n. sp. St. Petersburg Travaux Société pat. Comptes Rendus scances, vol. 40, livr. 1, pp. 157-166; deutsch résumé, pp. 167-169.

Société nat. Comptes Rendus seances, vol. 40, livr. 1, pp. 157-166; deutsch résumé, pp. 167-169.

NORMAN, A. M. The Polyzoa of Madeira and neighboring islands. Journal Linnean Society Zoology,
London, vol. 30, pp. 275-314, pls. 33-42.

NORMAN, A. M. The Celtic Province: Its extent and its marine fauna. Transactions Hertford Natural History Society, vol. 14, pp. 19-32.

PARONA, C. F.; CREMA C.; and PREVER, P. L. La fauna coralligena del Cretaceo dei Monti d'Ocre nell' Abruzzo Aquilano. Roma, Mem. serv. descr. Carta Geol. d'It. R. Com. Geol. Regno, vol. 5, pt. 1, pp. 1–242, pls. 1–27.

PRZIBRAM, HANS. Experimental Zoologie. 2. Regeneration. Leipzig and Wien, Franz Deuticke, pp. 1-338, pls. 1-16.

RAMOND, G.; COMBES, PAUL; and MORIN, MAURICE. Études géologiques dans Paris et sabanlieue. Note sur le gite fossilifère du Guespel. Paris Comptes rendus association française avanc. sci., vol. 37 (1908), pp. 476–493.

Retzius, Gustaf. Weiteres zur Kenntnis der Spermien der Bryozoen. Biol. Untersuch., vol. 14, p. 75-76, 1 pl.

ROAF, HERBERT E. Digestive enzymes of invertebrates. London, Report British Association Advancement Science, 1908 (1909), pp. 746-747.

SHEPPARD, T. Catalogue of specimens in the "Lether" collection and of the Cornbrash fossils in the Hull Museum. Hull Transactions Science Natural Cl. 4, pp. 71-80.

Siemiradzki, Josef. Zbiory L. Zejsznera. z kieleckiego dewonu. Kraków Spraw. Komk. fizyogr. vol. 43, pp. 62-94, 2 pls. (see also Kraków Bul. Int. Acad. pp. 765-770.)

SOMMER, KARL. Die fauna des Culms von Königsberg bie Giessen. Neues Jahrbucher Mineralogie, Stuttgart, vol. 28, pp. 611-660, pls. 27-30.

Sütoin, K. Sarajärven Eläimisto. Helsingfors Acta Société Fauna et Flora Fennica, vol. 29.

Walton, H. J. Large colonics of Hislopia lacustris. Records Indian Museum, vol. 3, pp. 295-296.

12184—23—Bull. 125——16

WATERS, A. W. Reports on the Marine Biology of the Sudanese Red Sea, XII. The Bryozoa. Cheilostomata. Journal Linnean Society London, vol. 31, pp. 123-181, 9 pls. (See Cyclostomata in 1910.)

Weller, Stuart. Kinderhook faunal studies 5. The fauna of the Fern Glen formation. Bulletin Geological Society of America, vol. 20, pp. 265-332, pl. 10-15.

ZSCHIESCHE, ALFRED. Untersuchungen über die Metamorphose von Alcyonidium mytili. Zoologische Jahrbücher, Jena, Abth. für Anatomie, vol. 28, pp. 1-72, 5 pls.

1910.

Annandale, N. Contributions to the fauna of Yunnan based on collections made by G. Coggin Brown.
B. Sc., 1909-1910. Pt. 1. Sponges and Polyzoa. Record of the Indian Museum, vol. 5, pp, 197-199.

Annandale, N. Note on a freshwater sponge and Polyzoon from Ceylon. Spolia Zeylanica Colombo, vol. 7, pp. 63-64, 1 pl.

Banfield, Arthur. Premature hatching of Cristella mucedo. Knowledge, new ser., vol. 7, p. 487, 1 fig.

Bower, C. R., and Farmery, G. R. The zones of the lower chalk of Lincolnshire. With a list of new records from the red chalk of the county. London, Proceedings Geological Association, vol. 21, pp. 333-359, pl. 27.

Brydone, R. M. Notes on new or imperfectly known chalk Polyzoa. Geological Magazine, dec. 5, vol. 7, pp. 4-6, pl. 3, pp. 76-78, pl. 8, pp. 145-147, pl. 14, pp. 258-260, pl. 21, pp. 390-392, pl. 30, pp. 481-483, pl. 36.

BUDDENBROCK, WOLFGANG V. Beiträge zur Entwicklung der Statoblasten der Bryozoen, Zeitschrift wissenschaftliche Zoologie, vol. 96, pp. 477~524, 3 pls. 8 figs. Zoological Anzeiger, vol. 35, pp. 534–538.

Canu, F. Bryozoaires des Terrains du Sud-Ouest de la France. 5, Lutetien; 6, Bartonien. Bulletin Société Géologique de France, ser. 4, vol. 10, pp. 840-855, pls. 16-19.

CANU, F. Bryozoaires des terrains tertiaires des environs de Paris, Annales de Paleontologie, vol. 2 (1907), pp. 57-89, 137-160, 8 pls., 8 figs.; vol. 3 (1908), pp. 61-104, 2 pls.; vol. 4 (1909), pp. 101-140, 4 pls; vol. 5 (1910), pp. 89-112, 4 pls.

Canu, F. Liste des Bryozoaires de la craie de Royan. Bulletin Société Géologique France, vol. 10, pp. 62-65.

CLOUGH, C. T., LEE, G. W. (and others). The geology of East Lothian, including parts of the counties of Edinburgh and Berwick. Memoir Geological Survey Scotland, pp. 1-226; pal. app., pp. 206-217.

Drake, Henry C., and Sheppard, Thomas. Classified list of organic remains from the rocks of the East Riding of Yorkshire. Proceedings Yorkshire Geological Society, vol. 17 pp. 4-71.

Fraas, E. Der Petrefactensammler. Ein Leitfaden zum Sammeln und Bestimmen der Versteinerungen Deutschlands. Stuttgart, pp. 1–276, pls. 1–82.

Girty, G. H. New genera and species of Carboniferous fossils from the Fayetteville shale of Arkansas.

Transactions New York Academy Science, vol. 20, pp. 189-238.

Hentschel, Ernst. Bryozoa für 1907. (Jahresbericht) Archives Naturges, Berlin, vol. 74, pt. 2, Heft 3, 1908 (1910), XIII. pp. 1-14.

Hentschel, Ernst. Bryozoa für 1908. (Jahresbericht) Archives Naturges Berlin, vol. 75, pt. 2. Heft 3, 1910, XIVa, pp. 1-9, Ber. Natg. nied. Tiere Berlin, new ser., vol. 25, 1910, pp. 1-40.

Harmer, S. F. Polyzoa. Encyclopaedia Brittanica. Ed. 11, vol. 22, pp. 42-45.

Janisevskij, M. E. Die Fauna des Unteren Kohlenkalkes von Chabarny, Bezirk Orsk, Gouv. Orenburg. Tomsk. lzv. technol. Inst., vol. 17, no. 1, pp. 1-305, 21 pls.

Korschelt, E., und Heider, K. Lehrbuch der vergleichenden Entwicklungs-geschichte der wirbelbsen Thiere. Vierter Abschnitt. Ungeschlechtliche Fortpflanzung und Regeneration. IX, Ungeschlechtliche Fortpflanzung, Jena, pp. 632-694.

LOPPENS, K. Catalogue des bryozoaires d'eau douce avec une note sur Victorella pavida. Annales Société malacologique Belgique, vol. 44, pp. 97-110.

LOPPENS, K. Fauna aquatica Europeae. Les Bryozoaires d'eau douce d'Europe. Annals Biologie lacustre, vol. 4, pp. 139-161, 16 figs.

MAPLESTONE, C. M. On the growth and habits of *Biporac*. Proceedings Royal Society Victoria, new ser., vol. 23, pp. 1–7, 1 pl.

Maplestone, C. M. On a new species of Cellepora from the south Australian coast. Proceedings Royal Society Victoria, new ser., vol. 23, pt. 1, pp. 39-41, 3 pls.

Maplestone, C. M. Observations on Parmularia obliqua and a fossil species. Proceedings Royal Society Victoria, new ser., vol. 23, pp. 42–43, 1 pl.

Martynow, A. Rapport préliminaire sur un voyage à station biologique de Roscoff. Prot. sèances Soc. Nat. Univ. Varsovie. Annals, vol. 22, p. 34-89.

NORMAN, A. M. Museum Normanianum, or a catalogue of the invertebrata of Europe and the Arctic and North Atlantic Oceans, which are contained in the collection of the Rev. Canon A. M. Norman. XIII, Polyzoa. Durham, T. Caldeleugh & Son.

Pace, S., and Pace, R. M. A bibliography and review of recent publications relating to the biology of the British and neighboring marine areas (i. e., of the North Atlantic and Arctic Oceans, Mediterranean Sea, etc.). London Bureau of British Marine Biology, ser. 2, nos. 1 and 2 (not paged titles 1-174).

Peach, B. N., Lee, G. W., and others. The geology of the neighborhood of Edinburgh. Memoirs Geological Survey Scotland, pp. 4-445; paleont. app., pp. 369-412.

Reed, F. R. Cowper. Sedgwick Museum notes. New fossils from the Dufton shales. London Geological Magazine, dec. 5, vol. 7, pp. 211-220, 294-299, pls. 16, 17, 23, 24.

Richardson, L. On a fuller's earth section at Combe Hay, near Bath. London, Proceedings Geological Association, vol. 21, pp. 425–428.

Robertson, Alice. The Cyclostomatous Pryozoa of the west coast of North America. University of California Publications, vol. 6, no. 12, pp. 225-284, 8 pls.

ROEHRICH, O. Description d'un aleyonidium nouveau. Archives de Zoologie experimentale et generale, ser. 5, vol. 5 (notes et rev., pp. 164-167, 3 figs.

Schröder, Olaw. Buddenbrockia plumatellae, eine neue Mesozoenart aus Plumatella repens L. und Pl. fungosa Pall. Zeitschrift Wissenschäftliche Zoologie, Leipzig, vol 96, pp. 525–537, 2 pls.

Schröder, O. Eine neue Mesozoenart (Buddenbrockia plumatellar n. s. n sp.) und Plumatella repens L. und Pl. fungosa Pall. Sitzungberichte Heidelberger Akademie der Wissenschaften, Kl. Abh. 6, pp. 1-8.

Skorikov, A. S. Zoologische Untersuchung des Wassers des Ladoga-Sees als Trinkwasser. St. Petersberg, pp. 1-123, 1 map and 1 plate.

Vinassa de Regny, Paolo. Fossili ordoviciani del nucleo centrale Carnico. Catania Mem. Accademia Giocnia scienze natural., ser. 5, mem. 12, pp. 1-48, 3 pls.

WALTHER, JOHANNES. Die Sedimente der Taubenbank im Golfe von Neapel. Abhandlungen der Kgl. Akademie der Wissenschaften zu Berlin, Abth. 3, pp. 1-49.

Waters, A. W. Reports on the Marine Biology of the Sudanese Red Sea, . . . XV. The Bryozoa. (Pt. I Cyclostomata, Ctenostomata and Endoprocta). Journal Linnean Society London Zool., vol. 31, pp. 231-256, 2 pls.

Woods, Rev. F. H. Marine biology at Redcar. Naturalist, London, pp. 646, 408-410.

WILLEM, VICTOR. Les "Néphridies" des Bryozoaires Phylactolémides. Comptes Rendus Association française Advancement Science, vol. 38, pp. 709-711.

ZSCHOKKE, F. Die Tiefenfauna hochalpiner Wasserbecken. Basel, Verhandlungen der naturforschenden Gesellschaft, vol. 21, pp. 145–152.

1911.

Annandale, N. Fresh-water sponges, Hydroids and Polyzoa. Fauna of British India. Bryozoa pp. 161–251, pls. 3–5.

Annandale, N. Systematic notes on the Ctenostomatous Polyzoa of fresh water. Records of the Indian Museum, Calcutta, vol. 6, pt. 4, pp. 193-201, pl. 13.

Bassler, R. S. The early Paleozoic Bryozoa of the Baltic Provinces. Bulletin 77, United States National Museum, 382 pages, 13 pls., text figs.

Bassler, R. S. Corynotrypa, a new genus of tubuliporoid Bryozoa. Proceedings U. S. National Museum, vol. 39, pp. 497-527, 27 figs.

Bassler, R. S. Bryozoa of the Middle Devonic of Wisconsin. Geological and Natural History Survey Wisconsin, Bulletin no. 21, pp. 49-67, pls. 5-11.

Braem, F. Die Variation bei den Statoblasten von Peetinatella magnifica. Archiv Entricklungs mechanik Organismen, Leipzig, vol. 32, pp. 314-348.

Braem, F. Beiträge zur Kenntniss der Fauna Turkestans auf Grund des von Pedaschenko gesammelten Materials. VII Bryozoa und deren Parasiten. Travaux Société Nat. St. Petersbourg, Sect. zool., vol. 42, fasc. 2, pt. 1, Zool. et Physiol. (Victorella continentalis, n. sp.)

Braem, F. Pterobranchier und Bryozoen. Zoologischer Anzeiger, vol. 38, pp. 546-551, 2 figs.

BRYDONE, R. M. Notes on New or imperfectly known chalk Polyzoa. Geological Magazine, dec. 5, vol. 8, no. 4, pp. 153-156, pls. 9, 10.

Calvet, Louis. Diagnoses de quelques espàces nouvellesde Bryozoaires Cyclostomes. Bulletin de l'Institute oceanographique, no. 215, pp. 1–9, 6 text figs.

Calvet, Louis. Sur deux espàces nouvelles de Bryozoaires de la Méditerranée. Archives Zoologie experimentale et generale, ser. 5, vol. 8, no. 3, p. lvii-lxi, 2 figs.

Canu, F. Les Bryozoaires fossiles des Terrains du Sud-Ouest de la France. VI Bartonien (suite). Bulletin Géologiqué Société France, ser. 4, vol. 11, pp. 444-445, pls. 7, 8.

Canu, F. Inconographie des Bryozoaires fossiles de l'Argentine, pt. 2, Anales del Museo Nacional de Buenos Aires, vol. 21 (ser. 3, f. XIV), pp. 215–292, pls. 1–12.

Fabiani, Ramiro. Paleontologia dei colli Berici. Roma, Mem. Soc., vol. 40, no. 3, 15, pp. 45–250, pls. 1–6.

Fric, Anton. Studien im Gebiete der Böhmischen Kreideformation. Ergänzung zu Band I. Illustriertes Verzeichniss der Petrefacten der cenomanen Korycaner Schichten. Archive der Naturwissenschaftlichen Landesdurchforschung von Bohmen. Prag. Vol. 15, no. 1, pp. 1–101.

GORTANI, M. Contribuzioni allo studio del Paleozoico carnico, IV. La fauna mesodevonica di Monu-

menz. Pl. Italiana, Pisa, vol. 17, pp. 141-228, pls. 16-20.

Guerin-Ganivet, Madame G. «Contributions à l'étude des Bryozoaires des côtes armoricaines. 1. Bryozoaires provenant du Haut Fond de la Chapelle et recueilles en 1900 par l'expédition de la Vienne. Travaux scientifiques du Laboratoire de Zoologie de Concarneau, vol. 3, fasc. 2, pp. 1-12, 2 pls.

GUERIN-GANIVET, MADAME G. Contributions à l'étude des Bryozoaires des côtes armoricanies. II. Bryozoaires provenant de la rade de Brest et recueillis par les freres Crouan. Travaux Scientifiques du Laboratoire de Zoologie de Concarneau, vol. 3, fasc. 5, pp. 1–7.

GUERIN-GANIVET, MADAME G. Étude préliminaire des Bryozoaires rapportés des côtes septentrionales de l'Europe par l'expedition du *Jacques-Cartier* en 1908. Bulletin Institut oceanographique Monaco, no. 207, 27 pp.

GIRTY, GEORGE II. The fauna of the Moorefield shale of Arkansas. Bulletin 439, U. S. Geological Survey, pp. 1-148, pls. 1-15.

HARMER, S. F. The terms Polyzoa and Bryozoa. Proceedings Linnean Society London, pp. 70-71.
HERDMAN, W. A. Note on J. V. Thompson's use of the term "Polyzoa." Proceedings Linnean Society London, pp. 62, 63.

Hennio, Anders. Le Conglomérat pleistocène à Pecten de l'île Cockburn. Wissenschaft. Ergebnisse der Schwed Südpolar-Expedition 1901–1903, Stockholm, 3 Lief, pp. 1-72, pls. 5.

Hurrell, II. E. Distribution of the Polyzoa in Norfolk waters. Transactions Norfolk and Norwich Naturalists Society, vol. 9, pp. 197-205.

Köhler, W. Pectinatella magnifica Leidy im Tegeler See. Internationale Revue Hyrobilogie, Leipzig, vol. 4, biol. Suppl. no. 5, pp. 6-7, 1 pl.

Kranz, W. Das Tertiar zwischen Castelgomberto, Montecchio. Maggiore, Creazzo und Monteviale im Vicentin. Stuttgart, Neues Jahrbuch Mineralogie, vol. 32, pp. 701–729.

LOHMANN, H. Die Cyphonautes der nordischen Meere. Nordisches Plankton, Lief. 13, no. 9, pp. 31-40, 8 tigs.

LUCAS, ROBERT. Bryozoa für 1909. (Jahresbericht) Archiv Naturgeschichte, Berlin, 76, 1910, vol. 6, Heft 1 (1911), pp. 117-143.

Maplestone, C. M. Further descriptions of the Tertiary Polyzoa of Victoria, pt. 11. Proceedings Royal Society Victoria, vol. 23, new ser., pt. 2, pp. 266-284 + 37-47.

MAPLESTONE, C. M. The results of deep sea investigations in the Tasman Sea. 1. The expedition of H. M. C. S. Miner. No. 5. Polyzoa, supplement. Records of the Australian Museum, vol. 8, pp. 118, 119, pls. 34, 35.

MORTENSEN, TH. A new species of Entoprocta, Loxosomella antedonis, from northeast Greenland. Københaven Danmark-Ekspeditionen til Grønlands Nordøstkyst, 1906-1908, vol. 5 Nr. 8. Reprint of Meddelelser Grønland, vol. 45, pp. 399-406, pl.

NICHOLS, A. R. Polyzoa from the coasts of Ireland, Fisheries, Ireland Scientific Investigations, 1910, no. 1, 37 pp., 1 pl.

Palk, Mary. On an enigmatic body in certain Bryozoa. Zoologischer Anzeiger, vol. 38, pp. 209-212,

Papp, Karl. Trias-Korallen aus dem Bakony. In Resultate der wissenschaftlichen Untersuchungen des Balaton (Plattensee). Anhang, 1 volume for 1911, pp. 1-23, 1 pl., Wien.

RITCHIE, J. On an Entoproctan Polyzoon (Barentsia benedeni) new to the British fauna, with remarks on related species. Transactions Royal Society Edinburg, vol. 47, pp. 835-848, 1 pl.

Stebbing, T. R. R. The terms Polyzoa and Bryozoa. Proceedings Linnean Society London, 123d session, pp. 61-62. Note on Thompson's use of the "Polyzoa" by Herdman, pp. 52-53. On Johnvaughan Thompson and his Polyzoa and on Vaunthompsonia, a genus of Sympoda, by Stebbing, p. 64-70. Note by Harmer, p. 70-71; by Waters, pp. 71-72.

Teller, Edgar E. A synopsis of the type specimens of fossils from the Paleozoic formations of Wiscon

sin. Milwaukee, Bulletin Wisconsin Natural History Society, vol. 9, pp. 170-271.

VINASSA DE REGNY, P. Trias-Tabulaten Bryozoen und Hydrozoen aus den Bakony. In Resultate der wissenschaftliche des Balaton (Plattensee) anhang I, volume for 1911, pp. 1-22, 2 pls.

WATERS, A. W. (The terms Polyzoa and Bryozoa.) London Proceedings Linnean Society, pp. 71-72.

AGATZ, JOSEPH. Knospung und Regeneration bei den Bryozoen. Diss. Strassburg. Bamberg (Druck v. w. Gärtner), pp. 1-30, 4 pls.

Andrussov, N. Die fossilen Bryozoenriffe der Halbinseln Kertsch und Taman. Kiev, Lief. 1,909 (pp. 1, 48 and 6 pls.); Lief, 2, 1911 (pp. 48-88 and 5 pls.); Lief. 3, 1912 (pp. 89-144 and 3 pls.).

Annandale, N. The occurrence of Entoprocta in Indian waters. Records of the Indian Museum Calcutta, vol. 7, pt. 2, p. 205.

Annandale, N. Fauna Symbiotica Indica. No. 1, Polyzoa attached to Indo-Pacific Stomatopods. Records of the Indian Museum Calcutta, vol. 7, pp. 123-126.

Annandale, N. Fauna Symbiotica Indica. No. 3. Polyzoa associated with certain Gangetic tortoises. Records of the Indian Museum Calcutta, vol. 7, pp. 147-150.

Annandale, N., and Kemp, Stanley. Observations on the invertebrate fauna of the Kumaon Lakes, with special reference to the sponges and Polyzoa. Records of the Indian Museum Calcutta, vol. 7, pp. 129-145.

Barroso, Manuel G. Briozoos de la Estacion de Biologia Maritima de Santander. Trabajos del Museo de Ciencias naturalés, no. 5, Madrid, pp. 1-63.

Braem, F. Nachträgliches über die Variation der Statoblasten von Statoblasten von Pectinatella. Arch. Intro. Mech. Leipzig, vol. 35, pp. 46-55.

BRYDONE, R. M. Notes on new or imperfectly known chalk Polyzoa. Geological Magazine, dec. 5, vol. 9, no. 1, pp. 7-8, pl. 1; no. 4, pp. 145-147, pl. 7; no. 7, pp. 294-296, pls. 14, 15; no. 10, pp. 433-435, pl. 22.

CALVET, LOUIS. Sur un Bryozoaire cténostome (Watersia paessleri n. g. n. sp.) parasitant le cormus d'une Synascidie. Comptes Rendus Academy Science, Paris, vol. 154, pp. 243-245. Apropos de Watersia paessleri, Bryozoaire parasite, p. 395.

CANU, F. Les Bryozoaires fossiles des Terrains du Sud-Ouest de la France. VI. Bartonein-Auversien. Bulletin Société Géologique France, ser. 4, vol. 12, pp. 623-630, pls. 20, 21.

Canu, F. Étude comparée des Bryozoaires Helvétiens de l'Egypte avec les Bryozoaires vivants de la Mediterranée et de la mer Rouge. Memoires de l'Institut Egyptien, vol. 6, fasc. 3, pp. 185-236,

CHAPMAN, FREDERICK. Note on a collection of Tertiary limestones and their fossil contents, from King Island, Melbourne. Memorial National Museum, vol. 4, pp. 39-51, pls. 6, 7.

Crawshay, L. R. On the fauna of the outer western area of the English Channel. Plymouth Journal Marine Biological Association, new ter., vol. 9, pp. 292-393.

CUMINGS, EDGAR R. Development and systematic position of the Monticuliporoids. Bulletin Geological Society America, vol. 23, pp. 357-370, pls. 19-22.

Cumngs, Edgar R., and Galloway, J. J. A note on the Batostomas of the Richmond series. Proceedings Academy Science Indiana, 1911, pp. 147-167, 7 pls.

Gravier, C. Sur une éspèce nouvelle de Cephalodiscus provenant de la seconde Expédition antartique française. Bulletin Histoire naturelle Paris, pp. 146-150, 1 fig.

Gravier, C. Sur la repartition geographique des especes actuellement connues du genere Cephalodiscus MacIntosh. Bulletin Museum Histoire naturelle Paris, pp. 151-153.

Gravier, C. Sur les Ptérobranches rapportés par la seconde Expédition antaretique française et sur un Crustacé parasite de l'nn d'eux. Comptes Rendus Academy Science Paris, vol. 154, pp. 1458-1440

Guerin-Ganivet, G. Contributions a l'étude des Bryozoaires des côtes Armoricaines, III. Bryozoaires de la region de Concarneau et de l'Archipel de Glenan. Travaux scientifique du Laboratoire de Zoologie de Concarneau, vol. 4, pp. 1–28.

HASPER, M. On a method of rearing larvae of Polyzoa. Journal Marine Biology Association Plymouth, new ser., vol. 9, pp. 435-436.

HOLTEDAHL, OLAF. On some Ordovician fossils from Boothia Felix and King William land collected during the Norwegian expedition of the Gjoa, Captain Amundsen, through the North West Passage. Kristiania Skr. Vid. selsk, vol. 9, pp. 1-11, pls. 1-4.

Lecointre, Georges. Sur quelques Bryozoaires nouveaux ou peu connus du Cénomanien du Mans. Bulletin Société Géologique, ser. 4, vol. 12, pp. 349–355, figs.

Lee, G. W. The British Carboniferous Trepostomata. Memoirs Geological Survey of Great Britains Paleontology, vol. 1, pt. 3, pp. 135-195, pls. 14-16.

Levinsen, G. M. R. Studies on the Cyclostomata operculata. Mémoires Academie Royale Science, et Lettres de Danemark, ser. 7, vol. 10, pp. 1-52, 7 pls., and 2 text figs. (Vid. Selsk. Skr.)

Lucas, Robert. Bryozoa für 1910. Archiv für Naturgeschichte, Berlin, vol. 77 (1911), pt. 6, Heft 1 (1912), pp. 99-116.

Moleanov, L. A. Zur Fauna der Wirbellosen der Seliger-Sees. St. Petersburg, Der Süsswasser Naturf, Gessellschaft, vol. 3, pp. 146–153.

Nichols, A. R. Clare Island Survey, pt. 53, Polyzoa. Proceedings Royal Irish Academy, vol. 31, no. 53, 14 pp.

Nordgaard, O. Revision av universitetsmuseets samling av norske Bryozoer. Kgl. norske Videnskabers Selskabs Skriften, 1911, no. 3, pp. 1–28.

Nordgaard, O. Bryozoaires de la Campagne arctique de 1907, de Duc d'Orleans. Bruxelles, vol 8, 42 pp., 1 map.

OSBURN, RAYMOND C. Bryozoa from Labrador, Newfoundland, and Nova Scotia. Proceedings U. S. National Museum, vol. 43, pp. 275-289, pl. 34, no. 1933.

OSBURN, RAYMOND C. Bryozoa of the Woods Hole Region. Bulletin Büreau of Fisheries, vol. 30 (1910), pp. 205-266, pls. 18-31.

RUEDEMANN, RUDOLF. The Lower Siluric shales of the Mohawk Valley. New York State Museum, Bulletin 162, pp. 1-151, pls. 1-10.

THORNELY, LAURA R. Marine Polyzoa of the Indian Ocean. Transactions Linnean Society London, Zoology, vol. 15, pp. 137-157, pl. 8.

Ulmer, Georg. Süsswasser-Bryozoen von Äquatorial-Afrika. (In: Wiss, Ergebnisse d. D. Zentral-Afrika Expedition 1907-8, vol. 4, Lfg. 10.) Leipzig (Klinkhardt und Biermann), pp. 285-290.

Waters, A. W. A structure in Adeonella (Laminopora) contorta Michelin, with remarks on the Adeonidae. Annals Magazine Natural History, ser. 8, vol. 9, pp. 489, 500, pls. 10, 11.

1913.

Annandale, N. The Polyzoa of the Lake of Tiberias. Calcutta, Journal Asiatic Society, Bengal, vol. 9, pp. 223-228, pl. 7.

Bassler, R. S. Bryozoa in Zittel's Textbook Paleontology (English edition), edited by Charles R. Eastman. Ed. 2, pp. 314-355.

Braem, F. Die Keimung der Statoblasten von Pectinatella und Cristatella (I. Morpholog, Vorgänge), Zoologica, Stuttgart, vol. 67 (=Band 26), pp. 35-64, 5 pls.

Brydone, R. M. Notes on new or imperfectly known chalk Polyzoa. Geological Magazine, dec. 5, vol. 10, no. 3, pp. 97-99; no. 5, pp. 196-199, pl. 7; no. 6, pp. 248-250, pl. 8; no. 10, pp. 436-438, pl. 14.

Canu, F. Bryozoaires fossiles des terrains éocéniques du Plà de la Gargara près Aiguafrede (Lutécien). Bulletin de l'Institut Catalona d'Histoire naturelle, no. 7, pp. 1-4, pl. 2a.

Canu, F. Études morphologiques sur trois nouvelles familles de Bryozoaires. Bulletin Société Géologique de France, ser. 4, vol. 13, pp. 132-147, text figs.

Canu, F. Les Bryozoaires fossiles des Terrains du Sud-Ouest de la France, VII, Lutécien. Bulletin Société Géologique France, ser. 4, vol. 13, pp. 298–303, pls. 4, 5.

Canu, F. Contributions à l'étude des Bryozoaires fossiles. IV-XII. Pliocène d'Alger, etc. Bulletin Société Géologique France, ser. 4, vol. 13, pp. 124-131.

Canu, F. Contributions à l'étude des Bryozoaires fossiles. XIII. Bryozoaires jurassiques. Bulletin Société Géologique France, ser. 4, vol. 13, pp. 267–276, pls. 2, 3.

Cumings, E. R., and Galloway, J. J. The stratigraphy and paleontology of the Tanner's Creek section of the Cincinnati series of Indiana. Department Geology and Natural Resources Indiana, 37th Annual Report, pp. 353-478, 20 pls.

Filliozat, Marius. Sur le genre "Ceriopora" Goldfuss, 1827. Comptes rendus du Congrès des Société Savantes, pp. 154–157.

GERWERZHAGEN, ADOLF. Beiträge zur Kenntnis der Bryozoen. 1. Das Nervensystem von Cristatella mucedo Cuv. Zeitschrift wissenschaftliche Zoologie, Leipzig, vol. 107, pp. 309–345, 3 pls.

GERWERZHAGEN, ADOLF. Untersuchungen an Bryozoen. (Vorl. Mitt.) Sitzungsberichte der Heidelberger Akademie der Wissenschaften, Abt. B, Abh. 9, pp. 1-16.

GIRTY, GEORGE H. A report on Upper Paleozoic fossils collected in China in 1903-4. Washington, D. C., Carnegie Institute Publications, no. 54 (Research in China, 3), pp. 295-334, pls. 27-29.

Guérin-Ganivet, G. Bryozoaires de la Mission arctique commandée par Ch. Bénard. Société d'océanographie du Golfe de Gascogne, fasc. 7, 46 pp.

HARMER, SIDNEY F. The Polyzoa of waterworks. Proceedings Zoological Society, London; pp. 426-457, pls. 62, 63.

Herwig, Ernst. Beiträge zur Kenntnis der Knospung hei den Bryozoen. Diss. Marburg (Druck v. R. Friedrich), 36 pp., 2 pls.

Kettner, K. Ueber das neue Vorkommen der untersilurischen Bryozoen und anderen Fossilien in den Ziegelei Pernikarka bei Kossire. Bulletin Int. Acad. St. Bohème, 22 pp., 2 pls., 9 figs.

Lano, W. D. Report of a visit to the exhibits of Polyzoa and Corals in the Geological Department of the British Museum of Natural History. Proceedings Geologists' Association, vol. 24, pt. 3, pp. 168–173.

LARGER, Dr. RENÉ. La contre évolution ou dégénérescence par l'hérédité pathologique; cause naturelle de l'extinction des groupes animaux. Essai de paléopathologie genéralé comparée. Bulletin et Mémoires de la Société d'Anthropologie de Paris.

Maplestone, C. M. Further descriptions of the Tertiary Polyzoa of Victoria, pt. 12. Proceedings Royal Society Victoria, new ser., vol. 24, pp. 355, 356, pl. 27.

MAPLESTONE, C. M. New or little known Polyzoa. Proceedings Royal Society Victoria, vol. 25, new ser., pp. 357-362, pl. 28.

Munroe, Madeline. Description of some new forms of Trepostomatous Bryozoa from the Lower Carboniferous rocks of the North Western Province (of England). London, Quarterly Journal Geological Society, vol. 68, pp. 574-579.

Schröder, Olaw. Ueber einen einzelligen Parasiten des Darmepithels von Plumatella fungosa Pallas. Zoologisher Anzeiger, Leipzig, vol. 43, pp. 220-223.

Summer, Francis B.; Osburn, Raymond C.; and Cole, Leon J. A hiological survey of the waters of Woods Hole and vicinity. Section 3. A catalogue of the marine fauna of Woods Hole and vicinity. Washington, D. C., Bulletin Bureau Fisheries, vol. 31 (1911), pp. 549-794.

ULRICH, E. O., and BASSLER, R. S. Bryozoa of the middle and Lower Devonic deposits of Maryland. Maryland Geological Survey, Lower Devonic, pp. 259-290, pls. 46-51; Middle and Upper Devonian, pp. 123-124, pl. 7, figs. 7-12.

WACKERHEIM, M. Moostierchen im Aquarium (Fredericella). Blätter Aquarienkunde, Stuttgart, vol. 24, pp. 298–299.

- Waters, A. W. Marine Fauna of British East Africa and Zanzibar. Bryozoa-Cheilostomata. Proceedings Zoological Society London, 1913, pp. 458-537, pls. 64-73. (See Cyclostomata, 1914.)
- Weber, R. Plumatella repens . . . (Mikrophotogr. Bilder). Mikrokosmos, Stuttgart, vol. 7, pp. 105-107.
- WOLFER, OTTO. Die Bryozoen des schwäbischen Jura. Paleontographica, vol. 60, pp. 115-173, 5 pls.

1914

- BRYDONE, R. M. Notes on new or imperfectly known chalk Polyzoa. Geological magazine, dec. 6, vol. 1, no. 3, pp. 97-99, pl. 4; no. 8, pp. 345-347, pl. 26; no. 11, pp. 480-483, pl. 35.
- Canu, F. Bryozoaires des Terrains du Sud-Ouest de la France. VIII. Rupelien de Gaas. Bulletin Société Géologique de France, ser. 4, vol. 14, pp. 465–474, pls. 14, 15.
- Canu, F. Contributions à l'étude des Bryozoaires fossiles. XIV. Bryozoaires du Stampien. Bulletin Société Géologique France, ser. 4, vol. 14, pp. 147–152, pl. 4.
- FAURA y SANS M. Sobre la presencia de un briozoa vivente, la "Cupularia canariensi?" Busk descubierto en los terrenos miocenices de Catalana. Boletín de la Real Sociedad española de Historia natural, pp. 397, 398.
- Lano, W. D. Some new genera and species of Cretaceous Cheilostome polyzoa. Geological Magazine, dec. 6, vol. 1, pp. 436-444, pl. 34.
- Lang, W. D. On Herpetopora a new genus containing three new species of Cretaceous Cheilostome Polyzoa. Geological Magazine, dec. 6, vol. 1, no. 595, pp. 5–8, pl. 2.
- OSBURN, RAYMOND C. The Bryozoa of the Tortugas Islands, Florida. Publication Carnegie Institution of Washington, no. 182, pp. 181-222, 23 text figs.
- WATERS, A. W. The Marine Fauna of British East Africa and Zanzibar. Cyclostomata. Ctenostomata, and Endoprocta. Proceedings Zoological Society London, 1914, pp. 831-858, pl. 1-4, text fig.

1915.

- Barroso, Manuel Gerónimo. Contribución al conocimiento de los Briozoos marinos de España. Boletín de la Real Sociedad española de Historia natural, pp. 413-420.
- Cumings, E. R., and Galloway, J. J. Studies of the morphology and histology of the Trepostomata or Monticuliporoids. Bulletin Geological Society America, vol. 26, pp. 349-374, pls. 10-15. (Review by Canu in Cossman's Revue Critique de Paléozoologie 1916, no. 1, p. 22.)
- GIRTY, GEORGE 11. Faunas of the Boone limestone at St. Joe, Arkansas. U. S. Geological Survey, Bulletin 598, Bryozoa, pp. 8-11.
- GIRTY, GEORGE H. Fauna of the Batesville sandstone of northern Arkansas. U. S. Geological Survey, Bulletin 593, Bryozoa, pp. 28-38.
- GIRTY, GEORGE H. Fauna of the Wewoka formation of Oklahoma. U. S. Geological Survey, Bulletin 544, Bryozoa, pp. 44-48.
- HARMER, S. F. The Polyzoa of the Siboga Expedition. Pt. I. Entoprocta, Ctenostomata, and Cyclostomata. Mono. XXVIII, Results Explorations Siboga, 180 pp., 12 pls. (Reviewed by Canu in Cossman's Revue Critique de Paléozoologie, 1916, no. 1, p. 24.)
- LANO, W. D. New uniserial Cretaceous Cheilostome Bryozoa. Geological Magazine, dec. 6, vol. 2, no. 11, pp. 496-504, pl. 17.

1916.

- BRYDONE, R. M. Notes on new or imperfectly known chalk Polyzoa. Geological Magazine, Dec. 6, vol. 3, no. 3, pp. 97-100, pl. 6; no. 6, pp. 241-243, pl. 10; no. 8, pp. 337-339, pl. 14; no. 10, pp. 433-435, pl. 18.
- Bassler, R. S. Bryozoa of the Upper Cretaceous deposits of Maryland. Maryland Geological Survey, Upper Cretaceous, pp. 736-745, pl. 46.
- Canu F. Bryozoaires (reviews) in Cossman's Revue Critique de Paléozoologie. Vingtième année, no. 1, pp. 22-27.
- Canu, F. Les Bryozoaires fossiles des Terrains du Sud-Ouest de la France. IX. Aquitanien. Bulletin Société Géologique France, ser. 4, vol. 15, pp. 320-334, pls. 3, 4.
- Canu, F. Bibliographie primitive relative aux Bryozoaires. Bulletin de la Société Géologique de France, ser. 4, vol. 14, pp. 287-292.

Canu, F. Bibliographie paléontologique relative aux Bryozoaires du Bassin de Paris. Bulleti de la Société Géologique de France, ser. 4, vol. 14, pp. 293–305, 381–410.

Gregory, J. W. Age of the Norseman limestone, Western Australia. Geological Magazine, dec. 6, vol. 3, no. 7, pp. 320, 321, text figs.

Lano, W. D. Revision of the "Cribrimorph" Cretaceous Polyzoa. Annals and Magazine Natural History, ser. 8, vol. 18, pp. 81–112.

LANG, W. D. Calcium carbonate and evolution in Polyzoa. Geological Magazine, dec. 6, vol 3, no. 620, pp. 73-77.

Levinsen, G. M. R. Danmark-Ekspeditionen til Grönlands Nordöstkyst, 1906–1908, vol. 3, no. 16; Bryozoa, pp. 435–472, pls. 19–24.

RAYMOND, PERCY E. Expedition to the Baltic Provinces of Russia and Scandinavia. Bulletin Museum Comparative Zoology, Harvard College, vol. 56, no. 3, pp. 179–286, 8 pls.

TWENHOFEL, W. H. Expedition to the Baltic Provinces of Russia and Scandinavia, 1914. Bulletin Museum Comparative Zoology, Harvard University, vol. 56, no. 4, pp. 289-354, 5 pls.

Waters, A. W. Some species of Crisia. Annals and Magazine of Natural History, ser. 8, vol. 18, pp. 469-477, pl. 16.

- 1917.

Barroso, Manuel Gerónimo. Notes sobre Briozoos. Boletín de la Real Sociedad Española de Historia Natural, vol. 17, pp. 494-499.

Brydone, R. M. Notes on new or imperfectly known chalk Polyzoa. Geological Magazine, dec. 6, vol. 4, no. 2, pp. 49-53, pl. 3; no. 4, pp. 145-148, pl. 9; no. 11, pp. 492-496, pl. 32.

Canu, F., and Bassler, R. S. A Synopsis of American Early Tertiary Cheilostome Bryozoa. United States National Museum, Bulletin 96, pp. 1–87, pls. 1–6.

Canu, F. Les Bryozoaires fossiles des Terrains du Sud-Ouest de la France. X. Burdigalien. Bulletin Société Géologique de France, ser. 4, vol. 16, pp. 127-152, pls. 2, 3, text figs.

Faura, Sans M., and Canu, F. Sur les Bryozoaires des terrains tertiaires de la Catalogne. Treballs de l'Institucio Catalana d'Historia Natural, vol., 1916, pp. 1-137, 9 pls:

FAURA, SANS M. Caracterización de la fauna briozoaria del Maestrichiense en el monte Perdido (Pirineos Centrales del alto Aragón). Boletín de la Real Sociedad de Historia natural, vol. 17, pp. 191-194.

LANG, W. D. The genotypes of certain Polyzoan genera. Geological magazine, dec. 6, vol. 4, no. 4, pp. 169-174.

LANG, W. O. On some new Cenomanian and Turonian Polyzoa. Geological Magazine, dec. 6, vol. 4, no. 6, pp. 256-258, pl. 17.

Mather, K. F. The Tertiary fauna of Wolf Island, Ontario. Ottawa Naturalist, vol. 31, pp. 33-40, pl. 1.

Okada, Yaichiro. A report on the Cyclostomatous Bryozoa of Japan. Reprint from the Annotationes Zoologicae Japonenses, vol. 9, pt. 3, pp. 355-360.

1918.

Barroso, Manuel Gerónimo. Notas sobre Briozoos. Boletín de la Real Sociedad Española de Historia Natural, vol 18, pp. 206-216, 307-309, 407-409.

BRYDONE, R. M. Notes on new or imperfectly known Cretaceous Polyzoa. Geological Magazines dec. 6, vol. 5, no. 1, pp. 1-4, pl. 1.

Canu, F. Hippaliosina, un nouveau genre de Bryozoaires. Compte Rend. Sommaire des Séances de la Société Géologique de France, no. 3, p. 40.

Canu, F. Hippaliosina. Un nouveau genre de Bryozoaires. Bulletin de la Société Géologique de France, ser. 4, vol. 18, pp. 88-94.

Canu, F. Les Ovicelles des bryozoaires cyclostomes, Etudes, surquelques familles nouvelles et anciennes. Bulletin Société Géologique de France, ser. 4, vol. 16, pp. 324-335, pl. 9.

Kindle, E. M., and Whittaker, E. J. Bathymetric check list of the marine invertebrates of eastern Canada. Department of the Naval Service, 8 George V, sessional papers, no. 38a, pp. 229-294.

Nordgaard, O. Bryozoa from the Arctic regions. Tromso Museums Aarshefter, vol. 40 (1917), nr. 1, pp. 1-99.

12184-23-Bull, 125-17

NORDGAARD, O. Bryozoa in the papers and collection of Dr. J. E. Gunnerus. Det Kgl. Norske Videnskabers Selskabs Skrifter, 1917, no. 5, pp. 1-5.

WATERS, ARTHUR WM. Some collections of the Littoral Marine Fauna of the Cape Verde Islands, Bryozoa. Journal Linnean Society Zoology, vol. 31, pp. 1-15, 4 pls.

WATERS, ARTHUR WM. Some Mediterranean Bryozoa. Annals and Magazine of Natural History, ser. 9, vol. 2, pp. 96-102, pl. 12.

Yanagi, Naokatsu, and Okada, Yaichiro. On a collection of Japanese Cheilostomatous Bryozoa, I. Annotationes Zoologicae Japanenses, vol. 9, pt. 2, pp. 407–429, pl. 6.

1919.

Barroso, Manuel Geronimo. Notas sobre Briozoos Españoles. Boletin de la Real Sociedad Española de Historia natural., vol. 19, pp. 200-204, 340-347.

Canu F., and Bassler, R. S. Bryozoa of the Canal Zone and related areas. Bulletin U. S. National Museum, no. 103, pp. 117-122, pl. 53.

Canu, F. Etudes sur les Ovicelles de la famille du Corymboporidae Smitt, 1866 (3d contribution). Bulletin de la Société géologique de France, ser. 4, vol. 17 (1917), pp. 348, 349, pl.

Canu, F. Les Bryozoaires fossiles des terrains du Sud-Oucst de la France. XL Rupelien (=Stampien). Bulletin de la Société Géologique de France, ser. 4, vol. 17 (1917), pp. 350-361, 2 pls.

Canu, F., and Bassler, R. S. Fossil Bryozoa from the West Indies. Pub. 291, Carnegie Institution of Washington, pp. 73-102, 7 pls.

CANU, F. Etudes sur les Ovicelles des Bryozoaires cyclostomes (2d contribution). Bulletin de la Société géologique de France, ser. 4, vol. 17 (1917), pp. 345-347, pl.

LANG, W. D. Old age and extinction in fossils. Proceedings Geologists' Association, vol. 30, pp. 102-113.
LANG, W. D. The Kelestominae; a subfamily of Cretaceous Cribrimorph Polyzoa. Quarterly Journal Geological Society, vol. 74, pp. 204-220.

Lang, W. D. The Pelmatoporinae; an essay on a group of Cretaceous Polyzoa. Philosophical Transactions of the Royal Society of London, Series B, vol. 209, pp. 191–228.

Marcus, Ernst. Notizen über einiges Material mariner Bryozoen des Berliner Zoologischen Museums. Sitzungsberichte der Gesellschaft Naturforschender Freunde, Berlin, no. 7, pp. 255–284.

OSBURN, RAYMOND C. Bryozoa of the Crocker Land Expedition. Bulletin American Museum of Natural History, vol. 40, pp. 603-624.

Vinassa de Regny, Paola. Fossili ordoviciani del Capolago (Seekopf) presso il Passo di Volaia (Alpi Carniche). Paleontographica Italica, vol. 21 (1915), pp. 97-116 (1-20), pl. 12, 13.

WATERS, ARTHUR WM. Batopora (Bryozoa) and its allies. Annals and Magazine Natural History, ser. 9, vol. 3, pp. 79-94, pl.

1920.

CANU, F. Les Bryozoaires fossiles de la region des Corbiéres. Bulletin Société Geologique de France, ser. 4, 18, pp. 294-314, 6 pls.

BARROSO, MANUEL GERONIMO. Notes sobre Briozoos Españoles, VIII. Boletin de la Real Sociedad Española de Historia Natural, vol. 20, pp. 353-362.

Canu, F., and Bassler, R. S. North American Early Tertiary Bryozoa. Bulletin 106, U. S. National Museum, 2 vols., 879 pp., 162 pls.

Canu, F. Bryozoaires Crétacés des Pyrénées. Bulletin Société Geologique de France, ser. 4, vol. 19, pp. 186-211, 2 pls.

Cipolla, F. Nota preventiva sui Bryozoi fossili di Alta villa (Palerme). Bolletino della Societa di Scienze Naturali ed Economiche, pp. 1-12.

Marcus, Ernst. Mittelmeer-Bryozoen des Berliner Zoologischen Museums, Sitzungsberichte der Gesellschaft Naturforschender Freunde, Berlin no. 2, pp. 65-108.

Okado, Yaichoro. Notes on some species of *Retepora* and *Adeonella* occurring in Japan, with descriptions of one new variety and five new species. Annotationes Zoologicae Japonenses, vol. 9, p. 5. pp. 613-634, 1 pl.

VINASSA DE REGNY, PAOLA. Sulla classificazione dei Treptostomidi. Atti della Societa Italiana di Science Naturali, vol. 59, pp. 212-231.

1921.

Barrosa Manuel Geronimo. Notas Sobre algunas especiés de Briozoos de España (Especiés del Golfo de Valencia). Real Sociedad Española de Historia Natural, vol. del 50 Ann., pp. 1-11.

Bekker, Hendrick. The Kuckers stage of the Ordovician rocks of N. E. Estonia. Acta et Commentationes Universitatis Dorpatensis, A. II, 1, pp. 1-92, 12 pls.

Bretnall, Rex W. Studies on Bryozoa. Records of Australian Museum, Sydney, vol. 13, no. 4, pp. 157-162.

CIPOLLA, FRANCESCO. I Briozoi Pliocenici di Altavilla presso Palermo. Giornale della Societa di Scienze Natural ed Economiche di Palermo, vol. 32, pp. 185, pls. 8.

CORYELL, HORACE N. Bryozoan faunas of the Stones River group of central Tennessee. Proceedings Indiana Academy of Science for 1919, pp. 261–340, 14 pls.

Duvergier, J. Note sur les Bryozoaires du Néogène de l'Aquitaine. Actes de la Société Linnéenne de Bordeaux, ser. 72, pp. 1–41, 4 pls.

MARCUS, ERNST. Bryozoen von den Juan-Fernandez Inseln in Skollsberg. C. The Natural History of Juan Fernandez and Easter Isles, vol. 3, pp. 93-124.

Marcus, Ernst. Indo-Pacifische Bryozoen aus dem Riksmuseum in Stockholm. Arkiv für Zoologi K. Svenska Vetenskapsakademien, vol. 14, no. 7, pp. 1–23, pls. 2.

MARCUS, ERNST. Papers from Dr. Th. Mortensens Pacific Expedition, 1914-16. VI. Bryozoen von den Auckland-und Campbell Inseln, Saertryk af Vidensk. Medd. fra Dansk Naturh. Foren., vol. 73, pp. 85-121, 1 pl.

Marcus, Ernst. Einiges über Bau und Entwicklung der Meeresbryozoen. Schriften der zoologischen Station Büsum für Meereskunde, pp. 22–27.

Marcus, Ernst. Bryozoen. In results of Dr. E. Mjobergs Swedish Scientific Expeditions to Australia 1910–13. Köngl. Svenska Vetenskaps akademiens Handlingar, vol. 61, No. 5, pp. 1–34, 2 pls.

Marcus, Ernst. Über die Verbreitung der Meeresbryozoen. Zoologischer Anzeiger, vol. 53, pp. 205-221.

Okado, Yaichoro. Notes of some Japanese Chilostomatous Bryozoa. Annotations Zoologicae Japon enses, vol. 10, pt. 3, pp. 19-32.

ROBERTSON, ALICE. Report on a collection of Bryozoa from the Bay of Bengal and other eastern seas Records of the Indian Museum, vol. 22, pl. 1, no. 8, pp. 33-65, 11 figs.

WATERS, ARTHUR WM. Observations upon the relationships of the (Bryozoan) Selenariadae, Conescherellinidae, etc., Fossil and Recent. Journal Linnean Society, Zoology, vol. 34, pp. 399-427, 2 plates.

Barroso, Manuel Geronimo. Notas Sobre briozoos marinos espagnoles No. X (Especies de Mahon, Baleares). Boletin de la Real Sociedad espanola de Historia Natural, vol. 22, pp. 88-101, 8 figs.

Canu, F., and Bassler, R. S. Studies on the Cyclostomatous Bryozoa. Proc. U. S. National Museum, vol. 61, pp. 1-160, 28 pls.

Canu, F. Bryozoa in Fossiles Crétacés de la cote orientale, Paleontologie de Madagascar. Annales de Paléontologie, vol. 11, pp. 16–30, 2 pls.

Marcus, Ernst. Südafrikanische Bryozoen aus der Sammlung des Gothenburger Museum. Göteborgs Kungl. Vetenskaps och Vitterhets Samhalles Handlingar, vol 25, pp. 1-45.

MARCUS, ERNST. Bryozoen von den Aru-Inseln. Abhandl. d. Senckenb. Naturf. Gesellsch., vol. 35, pp. 421–446, 2 pls.

Waters, A. W. On Mediterranean Tervia and Idmonea (Bryozoa). Annals and Magazine Natural History, ser. 9, vol. 10, pp. 1–16, 2 pls.

1923.

Boro, Folke. On the structure of Cyclostomatous Bryozoa, Arkiv för Zoologi. K. Svenska Vetenskaps-Akademien, vol. 15, no. 11, pp. 1-17.

Marcus, Ernst. Referat über die historische und moderne Auffassung des Baues und der systematischen Stellung der Bryozoengattung Adeona. Verhandlungen der zoolog.-botan. Gesellschaft in Wien, vol. 72, pp. 42-61.

APPENDIX.

TREMOGASTERINA TRUNCATOROSTRIS, new species.

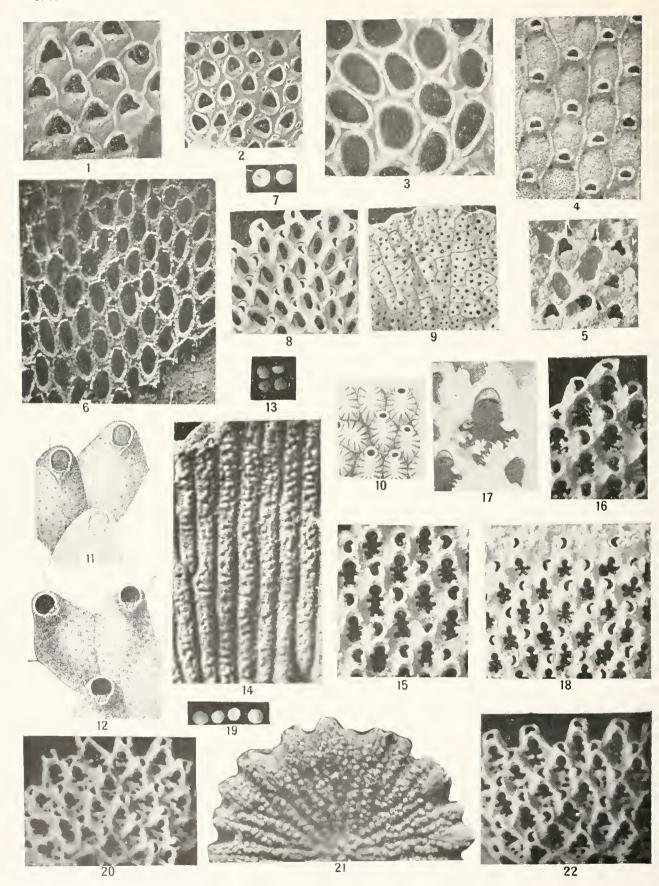
Plate 47, Fig. 6.

Description. The zoarium is bilamellar. The zooecia are distinct, separated by a furrow, elongated, elliptical; the frontal is perforated by a trifoliate orifice resulting from the coalescence of three large pores arranged in a triangle: a large salient rounded mucro is present and is prolonged somewhat above the aperture. The aperture is large, semicircular. On each side there are two large, straight avicularia, with pivot, the beak of which is truncated and forms a transverse straight line. The ovicell is large salient globular, finely granular.

 $\label{eq:measurements} \begin{array}{ll} \textit{Measurements.} - \Delta \text{pertura} \\ |\textit{ha} = 0.17 - 0.20 \text{ mm.} \\ |\textit{la} = 0.17 - 0.20 \text{ mm.} \end{array} \\ \begin{array}{ll} \textit{Zooecia} \\ |\textit{lz} = 0.35 - 0.40 \text{ mm.} \\ \end{aligned}$ $\textit{Affinities.} - \text{This new species differs from } \textit{Tremogasterina horrida, new species,} \\ \end{array}$

Affinities.—This new species differs from Tremogasterina horrida, new species, and T. problematica Canu, 1911, in its trifoliate frontal pore arising manifestly from the coalescence of three pores. It approaches in its ovicell Escharipora? mucronata Smitt, 1872, but differs from it in its large truncated avicularia and in its frontal pores united together. It is indeed an intermediate species between the forms with a large single frontal pore and those with three pores in a triangle. It is also probable that Smitt's species is also a Tremogasterina. If the studies on the living species prove this to be correct, Smitt's species could then be considered the type of this American genus whose structure has always remained mysterious.

Occurrence.—Miocene (Bowden horizon): Santo Domingo (very rare). Holotype. Cat. No. 68685, U.S.N.M.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

Fig. 1. Floridina pyripora Canu and Bassler, 1919. (p. 56.)

Surface of the incrusting zoarium, \times 20, showing the large poorly defined opesiules, the pyriform opesium and the smooth salient endozooccial ovicell.

Oligocene (Antigua formation): Ritle Butts, Antigua, Leeward Islands.

Fig. 2. Callopora dumerilii Savigny-Audouin, 1826. (See also pl. 2, fig. 23; pl. 12, fig. 12.) (p. 40.) Portion of the specimen, × 20, separated by Canu and Bassler in 1919 as var. lata.

Oligocene (Anguilla formation): Southwest side Crocus Bay Bluff, Anguilla, Leeward Islands.

Fig. 3. Ogivalina mutabilis Cann and Bassler, 1949. (p. 37.)

The incrusting type specimen, \times 20, illustrating the large, irregular opesia, small endozooccial ovicell, the small gymnocyst and the supposed avicularium.

Oligocene (Emperador limestone): Near Empire, l'anama Canal Zone.

Fig. 4. Calpensia impressa Moll, 1803. (p. 83.)

Surface of the incrusting zoarium, \times 20, referred to this species.

Oligocene (Antigua formation): Carlisle marl pit, Antigua, Leeward Islands.

Fig. 5. Floridina fusifera Canu and Bassler, 1919. (p. 56.)

The incrusting type specimen, \times 20, exhibiting the small opesium, large, rounded opesiules, prominent polypidian convexity, large onychocellarium and small fusiform avicularia.

Oligocene (Antigua formation): Rifle Butts, Antigua, Leeward Islands.

Fig. 6. Conopeum lacroixii Busk, 1852. (p. 26.)

Portion of the incrusting zoarium, \times 20, showing the characteristic interopesial cavities and small tuberosities on the mural rim.

Miocene (Bowden marl): Bowden, Jamaica.

Figs. 7-9. Cupuladria canariensis Busk, 1859. (p. 28.)

7. Two of the small, free, conical zoaria, natural size.

8. The cellulliferous, convex surface, \times 20, illustrating the membraniporoid opesium and the vibracula.

9. Concave, basal surface, \times 20, showing the characteristic polygonal areas marked by pores.

Miocene (Gatun formation): Banana River, Costa Rica.

Fig. 10. Puellina radiata carolinensis Gabb and Horn, 1862. (p. 90.)

Gabb and Horn's illustration of this variety.

Eocene (Jacksonian) of South Carolina.

Fig. 11. Hippoporina lata Smitt, 1872. (p. 131.)

View of several zooccia, much enlarged (after Smitt).

Recent: Gulf of Mexico.

Fig. 12. Porella bella Busk, 1860. (p. 147.)

Smitt's illustration of Escharella landsborovi.

Recent: Gulf of Mexico.

Figs. 13-17. Cupularia haidingeri Reuss, 1847. (p. 77.)

13. The small discoidal zoaria, natural size.

14. The inner side, \times 20, showing the tuberose, bifurcating ribs.

15, 16. Portions of the celluliferous side, \times 25, illustrating the two symmetrical condyles and the denticular processes.

17. A zooccium, \times 50, showing the structure of the spinose processes which are flat, claviform, and finely denticulated.

Redonnian: Pigeon Blanc, France.

Fig. 18, Cupularia doma D'Orbigny, 1852. (See also pl. 15, figs. 1-5.) (p. 77.)

Zooccia, × 25, showing the two symmetrical spinules forming condyles and three irregular ununited spinules.

Mediterranean: Oran, Algeria.

Figs. 19-22. Cupularia reussiana Manzoni, 1869. (p. 78.)

19. Several, small cupuliform zoaria, natural size.

20. Zooecia, imes 25, showing the trifoliate opesium and that the condyles are not united to the spinules.

21. Inner side, \times 20, illustrating the large tuberosities.

22. View of zooccia, \times 25, in which some of the spinules are not united together.

Sicilian: Farnesina, Italy.

Fig. 1. Membranipora vaughani Canu and Bassler, 1919. (p. 23.)

The incrusting zoarium, imes 20, showing the large rounded interzooccial tubercles.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Figs. 2, 3. Acanthodesia savarti forma monilifera Canu and Bassler, 1919. (p. 32.)

The hollow cylindrical zoarium, natural size and \times 20. The beaded structure of the mural rim is apparent.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Fig. 4. Hemiseptella lata Canu and Bassler, 1919. (p. 70.)

Part of the incrusting type specimen, \times 20, with some zooccia showing the characteristic inferior denticle

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Figs. 5-7. Nellia oculata Busk, 1852. (p. 55.)

- Normal zooecia with tubercles, complete, + 25.
- 6. Fragment showing large zooecia, > 25; the tubercles are replaced by pores.
- 7. View of fragment, \times 25, showing two sides of the zoarium.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Figs. 8-10. Corynostylus ellipticus Canu and Bassler, 1919. (p. 84.)

- 8. Several fragments of the articulated zoarium, natural size.
- 9. Anterior face of zoarium, \times 25, showing the characteristic unilamellar growth with two rows of zooccia.
 - 10. Posterior face, \times 25.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Figs. 11-13. Corynostylus labiatus Canu and Bassler, 1919. (p. 84.)

- 11. Complete segment of the articulated bilamellar zoarium, natural size.
- 12. Photograph of the same, \times 25. The salient opesial lip is shown.
- 13. Section, < 20, exhibiting the interior of the zooccia in the middle row and a longitudinal section in the outer rows.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Fig. 14. Thalamoporella granulata Levinsen, 1909. (p. 61.)

Small fragment, \times 20, identified with this recent species.

Lower Miocene (Bowden horizon): Rio Gurabo, Santo Domingo.

Figs. 15-19. Cupularia umbellata Defrance, 1823. (p. 80.)

- 15. The discoid zoaria, natural size.
- 16. Photograph of the inner tuberose side, \times 25.
- 17. Ancestrular region, \times 25, in which the ancestrula is covered over by a normal zooecium.
- 18. Ancestrular region of another zoarium, \times 25; the zooccia are arranged alternately and in inverted order.
- 19. Another view of the ancestrular region, \times 25; here the ancestrula is a membraniporoid zooecium

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Figs. 20, 21. Smitting? brevis Canu and Bassler, 1919. (p. 146.)

- 20. Surface of the bilamellar zoarium, \times 20.
- 21. Ovicelled zooccia of another fragment, \times 20.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

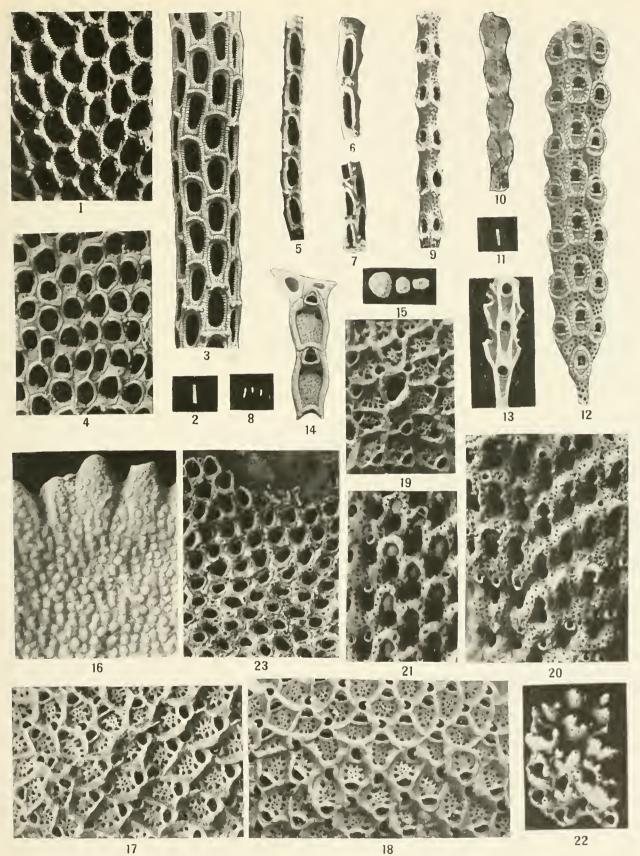
Fig. 22. Rhynchosoon vaughani Canu and Bassler, 1919. (See also pl. 4, figs. 1, 2.) (p. 155.) A small fragment, × 20.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Fig. 23. Callopora dumerilii Savigny-Audouin, 1826 (see also pl. 1, fig. 2; pl. 12, fig. 12.) (p. 40)

An example, × 20 showing thick mural rims and regenerated zooceia.

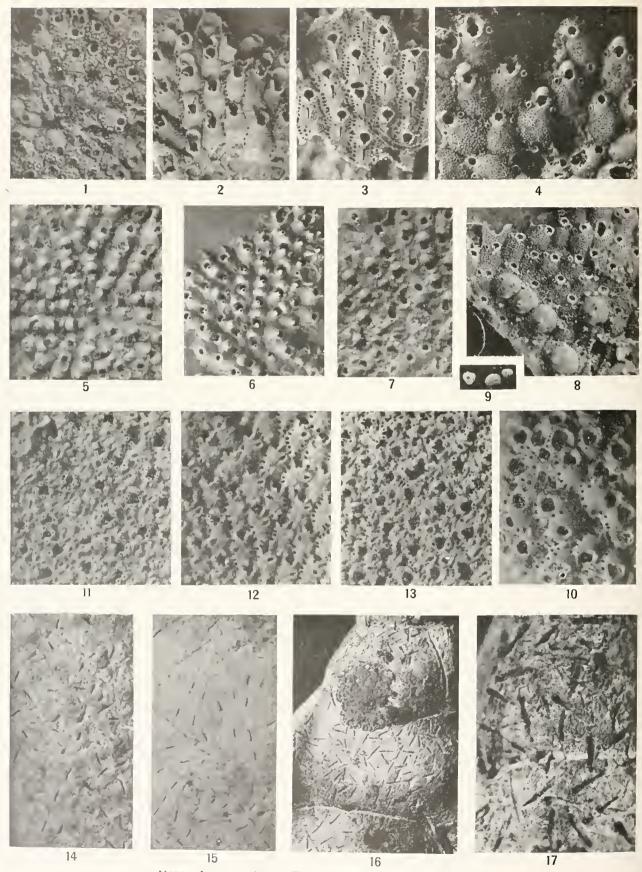
Lower Miocene (Bowden marl): Bowden, Jamaica.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

U. S. NATIONAL MUSEUM

BULLETIN 125 PLATE 3



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

PLATE 3.

Fig. 1. Hippodiplosia baccata Canu and Bassler, 1920. (p. 131.)

View, > 20, of Miocene example of this species.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Fig. 2. Rhamphostomella granulosa, new species. (p. 151.)

The incrusting zoarium, \times 20, showing the granular pleurocyst, small areolar pores, and the aperture with its triangular avicularium.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Fig. 3. Smittina ophidiana Waters, 1877. (p. 144.)

A small incrusting zoarium, > 20. The large median avicularium, two frontal pores, rectangular lateral pores, and granular ovicell are to be noted.

Miocene (Bowden marl): Bowden, Jamaica.

Fig. 4. Mastigophora granulosa, new species. (p. 172.)

Ovicelled and unovicelled zooccia of the incrusting zoarium, \times 20, illustrating their large size, granular surface, and the position of the vibracula at the level of the rimule.

Lower Miocene (Bowden marl): Bowden, Jamaica,

Figs. 5-7. Aimulosia brevis, new species. (p. 140.)

5. The incrusting zoarium, imes 20, illustrating the zooecia in the ancestrular region.

Lower Miocene (Bowden marl): Bowden, Jamaica.

6. A typical example, \times 20, with the usual short zooecia bearing very salient median axicularia.

7. Marginal portion of a zoarium, × 20, exhibiting larger zooccia.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida.

Fig. 8. Stylopoma minuta, new species. (p. 104.)

Portion of the incrusting type specimen, \times 20. The ovicell entirely covering the apertura and the small size of the zooccia are to be noted.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Figs. 9, 10. Holoporella hemispherica, new species. (p. 176).

9. Several of the small, free, hemispherical zoaria, natural size.

10. Zooecia. imes 20, illustrating their large size and convex frontal, surrounded by large areolar pores.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Figs. 11-13. Rhynchozoon verrueulatum Smitt, 1872. (p. 157.)

11. Central portion of the incrusting zoarium, \times 20, showing indistinct zooccia.

12. Marginal zooecia, ×20, with outlines well preserved.

13. Another aspect of the zooecial surface, \times 20.

Phocene Caloosahatchee marl: Shell Creek, De Soto County, Florida.

Figs. 14, 15. Terebripora sincfilum, new species. (p. 15.)

14. A normal zoarium, × 20, illustrating deeply perforated old zooccia and absence of canals.

15. Portion of a young zoarium, > 10, showing both the axes and the zooecia. The latter have not perforated the shell very deeply.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Figs. 16, 17. Terebripora elongata, new species. (p. 15.)

16. The type specimen, \times 3 and a small portion, \times 10, illustrating the very large size and elongate shape of the zooccia.

Lower Miocene (Bowden marl): Bowden, Jamaica.

PLATE 4.

- Figs. 1, 2. Rhynchozoon vaughani Canu and Bassler, 1919. (See also pl. 2, fig. 22.) (p. 155.)
 - 1. The unilamellar zoarium, $\sqrt{20}$, showing young condition in which the zooccia are distinct.
 - 2. Old condition of zoarium, \times 20. The zooecia are indistinct and not oriented.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

- Figs. 3-12. Metrarabdotos colligatum Canu and Bassler, 1919. (p. 162.)
 - 3.5. Different aspects of the base of the zoarium, \times 6.
 - 6. Three fragments, natural size, preserving base.
 - 7. Portion of zoarial surface, \times 20, with oviceHs well preserved.
 - 8. Base of a zoarium, \times 20, showing calcification and disappearance of the apertures.
 - 9. Broad zooecia, \times 20, characteristic of the aged condition.
 - 10. Narrow zooecia, \times 20, of youthful condition.
 - 11. Normal zooccia, \times 20.
 - 12. Longitudinal section, \times 20, exhibiting the interior of the zooecia.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo,

Fig. 13. Palmicellaria cf. inermis Jullien, 1882. (p. 149.)

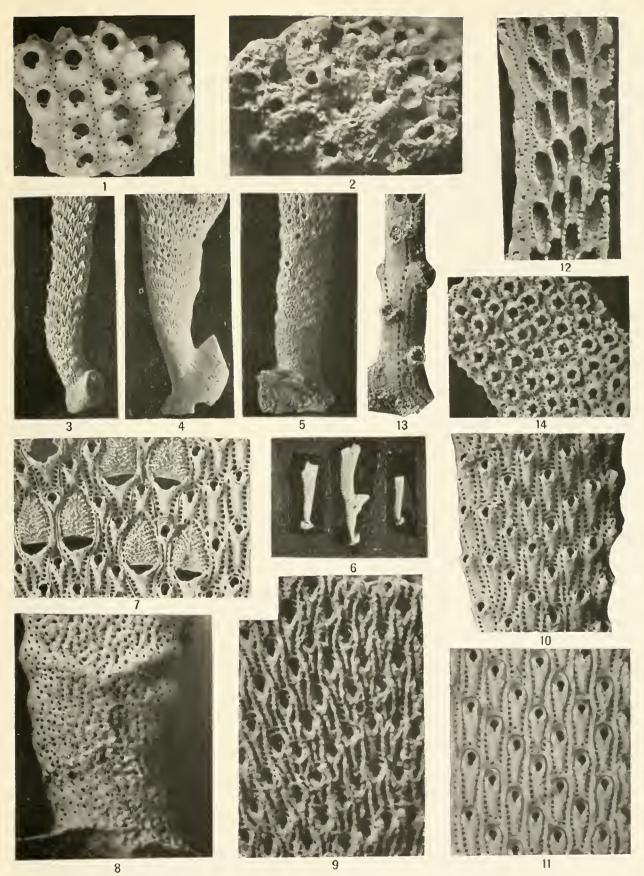
The branching fragment referred doubtfully to this species, \times 20.

Lower Miocene (Bowden horizon): Santo Domingo.

Fig. 14. Rhynchozoon curtum, new species. (p. 156.)

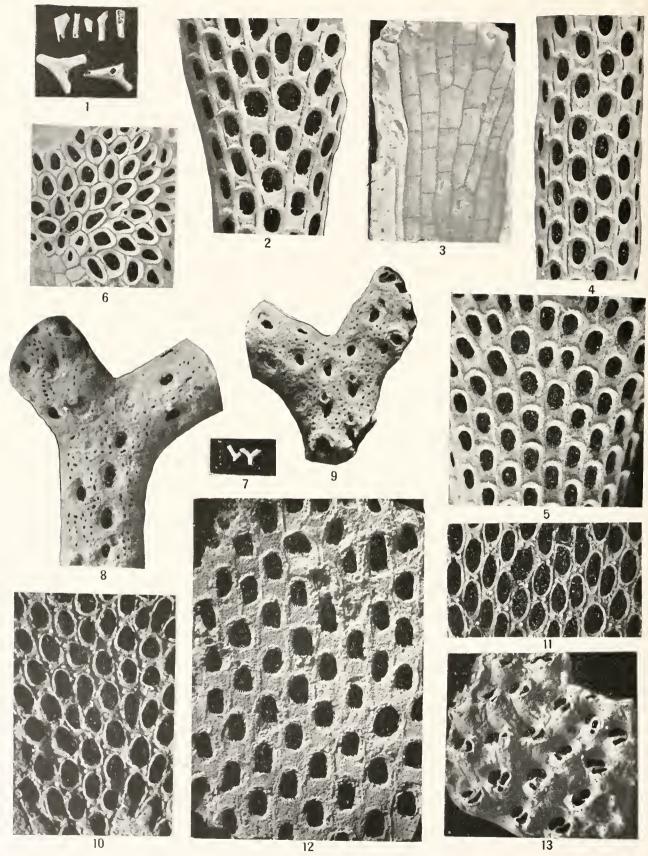
The unilamellar zoarium, \times 20, illustrating the characteristic short zooecia.

Lower Miocene (Bowden horizon): Santo Domingo.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 248.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 249.

PLATE 5.

- Figs. 1-5. Acanthodesia savarti forma texturata Reuss, 1847. (See also pl. 46, figs. 8, 9.) (p. 32.)
 - 1. Fragments of the unilamellar, hollow cylindrical zoarium, natural size.
 - 2. Zooecia, \times 20, with traces of delicate spines in the openium.
 - 3. Inner side of zoarium, × 20, showing rectangular form of zooccia.
 - 4. Zooecia, \times 20, with a distinct rim around the opesium.
 - 5. Another surface, \times 20, showing the large size of the first zooecium of a new row.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Fig. 6. Conopeum orale Cann and Bassler, 1919. (p. 26.)

Ancestrular portion of the type specimen, \times 20, incrusting a shell. The rare and irregular interopesial cavities, the entire, oval opesium and the flat mural rim are illustrated.

Lower Miocene (Bowden marl): Bowden, Jamaica.

- Figs. 7-9. Gemelliporella punctata Canu and Bassler, 1919. (p. 111.)
 - 7. Two fragments of the free cylindrical zoarium, natural size.
 - 8. An example, \times 20, showing the development of the tremocyst.
 - 9. Another fragment, imes 20, exhibiting the form of the aperture, ovicells (broken) and arrangement of the areolae.

Lower Miocene (Bowden marl): Bowden, Jamaica.

- Figs. 10-II. Membraniporina tenella Hincks, 1880. (p. 24.)
 - 10. The incrusting zoarium, imes 20, illustrating the thin mural rim and the small tubercles on the gymnocyst.
 - H. Another zoarium, \times 20, with narrow zooecia.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Fig. 12. Hemiseptella grandicella Canu and Bassler, 1919. (p. 71.)

Surface of the incrusting zoarium, \times 20. The largest zooecia commence a new row.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Fig. 13. Rhamphostomella laticella Canu and Bassler, 1919. (p. 151.)

The incrusting zoarium, \times 20. The arcolar costules are scarcely visible.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Fig. 1, Labiopora miocenica Canu and Bassler, 1919. (p. 67.)

Fragment of the unilamellar zoarium, - 20. The polypidiam tube and the small pores of the tremocyst are visible.

Lower Miocene (Bowden horizon): Rio Gurabo, Santo Domingo.

Fig. 2, Hippomenella infratelum Canu and Bassler, 1919. (p. 132.)

Base of the bilamellar zoarium, \times 20. One zooccium is regenerated, being replaced by an avicularium.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

Figs. 3-5. Schizopodrella mutabilis Canu and Bassler, 1919. (p. 105.)

- 3. The tubular zoarium, natural size.
- 4. Surface, \times 20, showing indistinct zooecia.
- 5. Another portion of the same zoarium, \times 20, illustrating the shape of the zooccia.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

- Figs. 6-9. Steganoporella parvicella Canu and Bassler, 1919. (p. 62.)
 - 6. Fragments of the unilamellar zoarium, natural size.
 - 7. Zoarial fragment, > 20, illustrating the great irregularity in the zooecia.
 - 8. The usual zooecia, \times 20, with regular arrangement.
 - 9. The most frequent aspect of the zooccia, \times 20. The polypidian tube is very fragile and often broken or altered,

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

- Figs. 10-15. Thalamoporella biperforata Canu and Bassler, 1919. (p. 62.)
 - 10. Fragments of the bilamellar zoarium, natural size.
 - 11. Specimen, -, 20, exhibiting a well-preserved reticulocellarium.
 - 12. An example, \times 20, showing the hollow tubercles worn and replaced by pores. A reticulocellarium is here present.
 - 13. Drawing showing the real form of the aperture, \times 77.
 - 14. A specimen, × 20, with a deformed membraniporoid zooccium near the upper corner.
 - 15. Another similar example, \times 20, with a primoserial membraniporoid zooecinm along the lower edge.

Lower Miocene (Bowden horizon): Cercado de Mao and Rio Cana, Santo Domingo.

Figs. 16-19. Mamillo pora tuberosa Canu and Bassler, 1919. (See also pl. 7, figs. 1-8.) (p. 192.)

- 16. Small fragments of the small, free, conical zoarium, natural size.
- 17. Zoarial fragment, < 20, with large zooecia.
- 18. Fragment, > 20, with small zooccia. The tubercles are well developed.
- 19. Inner side of zoarium, \times 20.

Lower Miecene (Bowden horizon): Rio Cana, Santo Domingo.

Figs. 20, 21. Diaperoecia milneana D'Orbigny, 1839. (p. 202.)

Views of a fragment, natural size and \times 12, referred to this species.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

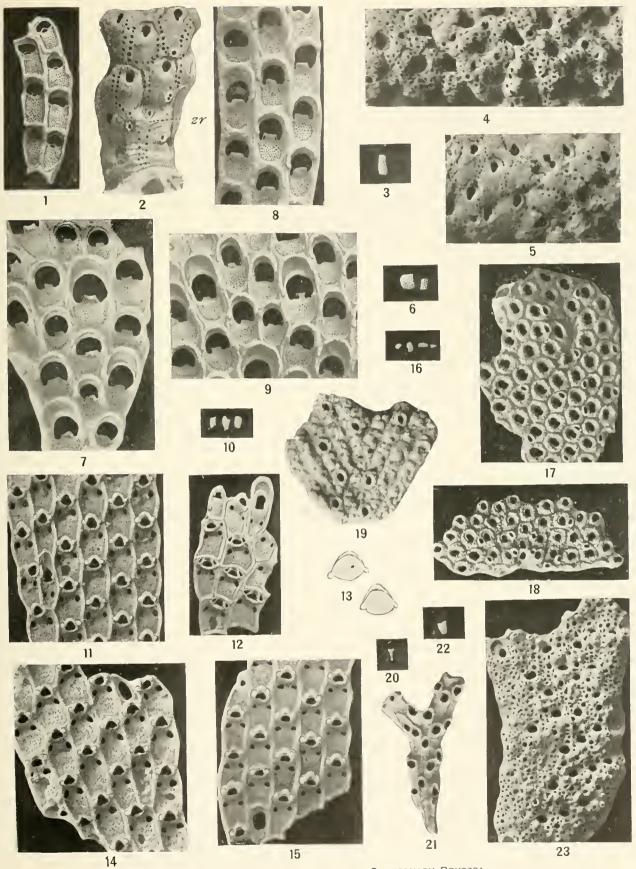
Figs. 22, 23. Adeona porosa Cann and Bassler, 1919. (p. 153.)

Fragment of the free bilamellar zoarium, natural size and \times 20.

In figure 23, three gonoecia with their oral gibbosities are shown in the upper right-hand corner, an avicularian zooecium in the upper left-hand corner, and ordinary zooecia with their ascopores, and small areolae in the rest of the photograph.

Lower Miocene (Bowden horizon): Cercado de Mao, Santo Domingo.

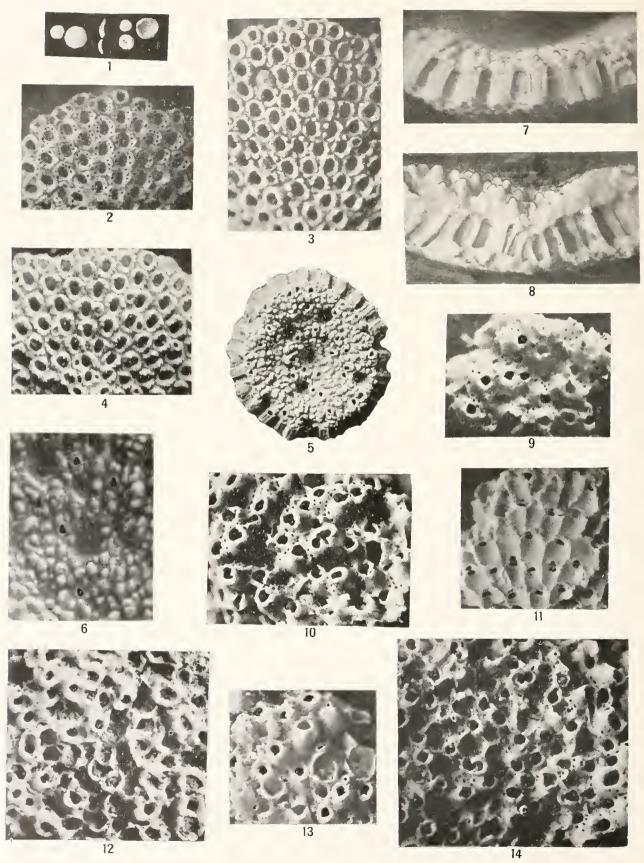
BULLETIN 125 PLATE 6



North American Later Tertiary and Quaternary Bryozoa.

FOR EXPLANATION OF PLATE SEE PAGE 250.

U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

PLATE 7.

- Figs. 1-8. Mamillopora tuberosa Canu and Bassler, 1919. | See also pl. 6, figs. 16-19.) (p. 192.)
 - 1. Group showing the upper, lower, and edge views of the zoarium, natural size.
 - 2. Zoarium with weathered surface, \times 20, illustrating that the tuberosities are hollow.
 - 3. Zooecia, \times 20, with one preserving the ovicell, which is hyperstomial and closed by the oper-culum.
 - 4. Portion of a zoarium, \(\sigma 20, \) with the tuberosities around the zooecial apertures well preserved
 - 5. Inner surface of zoarium, \times 10. The tuberosities, avicularia, and hydrostatic (?) cavities are visible.
 - 6. Portion of figure 5, \times 20.
 - 7. Edge view of fractured zoarium, \times 20, illustrating zooecia open.
 - 8. Similar view, \times 20, but with the zooecia closed.
 - Lower Miocene (Bowden marl): Bowden, Jamaica.
- Figs. 9-14. Holoporella albirostris Smitt, 1872. (See also pl. 32, figs. 6-10.) (p. 174.)
 - 9. Fragment, \times 20, showing zooecia without beak with the typical form of aperture.
 - 10. Zooecia, \times 20, showing interzooecial avicularia. The oral beak is incomplete and in process of formation.
 - 11. Interior of zooecia, \times 20.
 - 12. General aspect of the zooecia with ovicell, \times 20.
 - 13. Marginal zooccia, \times 20. The deep zooccia have a beak bearing a small avicularium which appears then as isolated.
 - 14. Zooecia, \times 20, showing the different aspects of the beak when broken.

Lower Miocene (Bowden marl): Bowden, Jamaica.

PLATE 8.

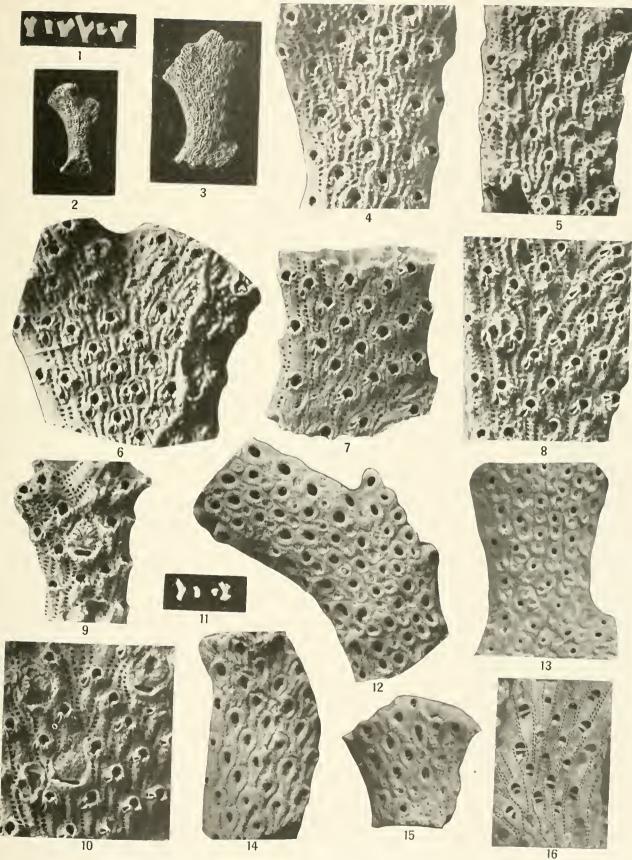
- Figs. 1-10. Metrarabdotos lucrymosum Canu and Bassler, 1919. (p. 164.)
 - 1. A group of fragments, natural size.
 - 2. Fragment, × 6, showing the little expanded base.
 - 3. Another fragment, \times 6.
 - 4. A worn branch, \times 20. The avicularia are absent or little apparent.
 - 5. Zooecia, \times 20, with salient peristomes and large triangular avicularia.
 - 6. Worn zoarial fragment, \times 20.
 - 7. Ordinary zooccia of a branch, \times 20, with short, pointed triangular avicularia.
 - 8. Ordinary zooecia, \times 20, with long avicularia.
 - 9. Branch, \times 20, with ovicelled zooccia. The ovicell is slightly costulated and its frontal is granu-lose.
 - 10. Ovicelled zooccia, \times 20, with the ovicell not costulated.

Lower Miocene (Bowden marl): Bowden, Jamaica.

- Figs. 11-16. Bracebridgia deformis Canu and Bassler, 1919. (p. 160.)
 - 11. Fragments, natural size.
 - 12. The usual aspect of the zooecia, \times 20. The large frontal pore does not perforate the wall.
 - 13. Zooccia at the base of a zoarium, \times 20.
 - 14. Zooecia, \times 20 with the large frontal pore wanting.
 - 15. Zooecia, \times 20, with the peristomial axicularium slightly visible.
 - 16. Interior of zooecia, \times 20. The peristomial avicularium is visible, but the frontal pore does not perforate the zooecial wall.

Lower Miocene (Bowden marl): Bowden, Jamaica.

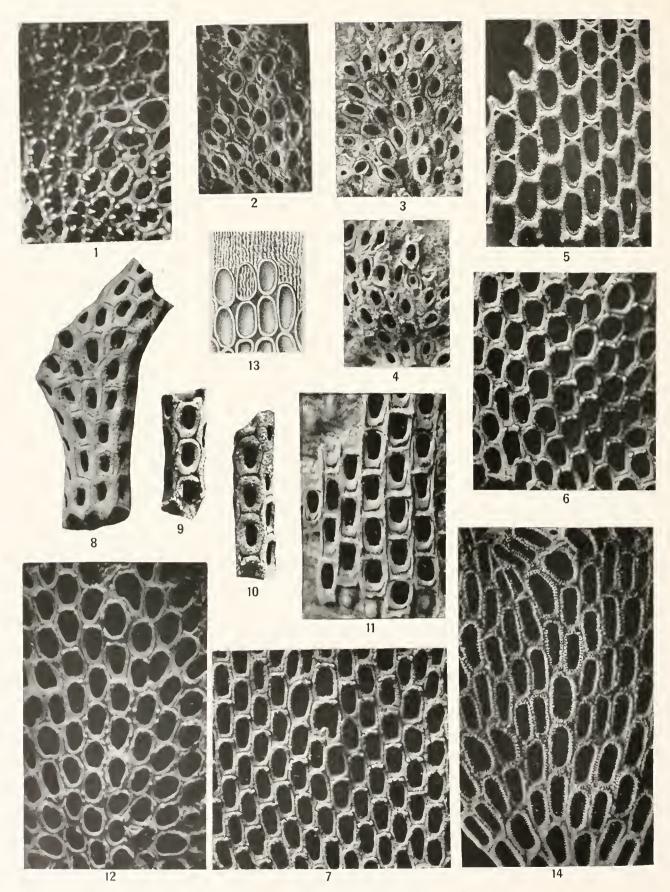
U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

U. S. NATIONAL MUSEUM

BULLETIN 125 PLATE 9



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 253.

Fig. 1. Membranipora spiculata, new species. (p. 21.)

Zooecia of the incrusting type specimen, \times 20, showing the curved spicules.

Miocene: Kuhns, Carteret County, North Carolina.

Figs. 2-4. Pyripora brevicauda, new species. (p. 19.)

2. Portion of the zoarium \times 20. showing several of the fusiform heterozooccia.

3. Another zoarium \times 20, illustrating the flabelliform growth. Normal and deformed zooecia are present.

4. Zooecia, × 20, with several calcified ones present.

Miocene: Kuhns, Carteret County, North Carolina.

Fig. 5. Conopeum nitidulum Ulrich and Bassler, 1904. (p. 27.)

View of the type specimen \times 20 illustrating the small flat cryptocyst and the triangular interopesial spaces.

Miocene (Choptank formation): Pawpaw Point, Maryland.

Figs. 6, 7. Membranipora fossulifera Ulrich and Bassler, 1904. (p. 20.)

6. An irregular specimen \times 20 showing the large interzooecial tubercles, the small spines on the mural rim and some zooecia without tubercles.

Miocene: Kuhns, Carteret County, North Carolina.

7. Photograph \times 20 of the type specimen.

The median channel between the walls, giving rise to the specific name, is apparent.

Miocene (Calvert formation): Reeds, Maryland.

Figs. 8-10. Membraniporina vincularina, new species. (p. 26.)

8. A bilamellar zoarium, \times 20, referred to this species.

Miocene (Yorktown formation): Petersburg, Virginia.

9, 10. The types of the species, fragments of the free rod-like zoaria, \times 20,

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Fig. 11. Acanthodesia rectangularia, new species. (p. 34.)

The incrusting type specimen \times 20, showing the rectangular zooecia with deep cryptocyst.

Miocene (Yorktown formation): Petersburg, Virginia.

Fig. 12. Membranipora tuberimargo, new species. (p. 23.)

Portion of the type specimen \times 20 showing both ancestrula and normal zooccia. The single pair of distal tubercles and two pairs of lateral ones are quite visible on some zooccia.

Miocene (St. Mary's formation): Cove Point, Maryland.

Fig. 13. Membranipora speciosa, Gabb and Horn, 1862. (p. 44.)

The original figure of this undetermined species.

Miocene? Chiriqui, Central America.

Fig. 14. Membranipocina baccata, new species. (p. 25.)

The incrusting, unilamellar zoarium, \times 20, illustrating the numerous granules of the mural rim. Miocene (St. Mary's formation): Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia.

Figs. 1-3. Acanthodesia oblongula Ulrich and Bassler, 1904. (p. 34.)

1. One of the original types of this incrusting species, \times 20. illustrating the prominent inter-zooccial tubercles and the spinose processes (spicules) surrounding the opesium.

Miocene (Calvert formation): Plum Point, Maryland.

2. A specimen, \times 20, showing the interzooccial tubercles joined.

Miocene (Choptank formation): Pawpaw Point, Maryland.

3. Another of the original types, \geq 20, in which there are numerous spinose processes but with the interzooccial tubercles little developed.

Miocene (Choptank formation): Governor Run, Maryland.

Figs. 4-5. Vibracellina pusilla, new species. (p. 35.)

4. Ancestrular region of the incrusting zoarium, \times 20.

Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, North Carolina.

5. A well-developed zoarium, \times 20, illustrating normal and regenerated zooccia and the small but salient vibracula.

Pliocene (Caloosaliatchee marl): Shell Creek, De Soto County, Florida.

Figs. 6-7. Vibracellina simpler, new species. (p. 35.)

6. The incrusting zoarium, \times 20, showing the ancestrular region. The vibracula are very rare but heterozooccia are present.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

7. A small zoarium, 4 20, showing the calcified ancestrula which engenders five zooecia. Several regenerated zooecia are present.

Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, North Carolina,

Fig. 8. Conopeum germanum Ulrich and Bassler, 1904. (p. 27.)

The incrusting type specimen, \times 20, illustrating the interopesial cavities and other characteristics.

Miocene (Choptank formation): Dover Bridge, Maryland.

Fig. 9. Hemiseptella filimargo, new species. (p. 71)

The incrusting type specimen, \times 20. The salient rim separating the zooccia and the two large lateral as well as the smaller spicules of the opesium are evident.

Miocene (Yorktown formation): York River, Virginia.

Fig. 10-14. Membranipora flabellata Canu, 1904. (p. 20.)

10. Surface of an outer lamella, < 20, with the tubercles worn away and the zooecia grouped around a false ancestrula.

Miocene (Yorktown formation): Yorktown, Virginia.

11. Portion of a zoarium, + 20, with the zooecia separated by interopesial cavities.

12. Young zone ia, < 20, with thin mural rims. Other zonecia of this same zonrium are normal, tuberose or separated by interopesial cavities.

13. Zooecia, \times 20, with granulose mural rims.

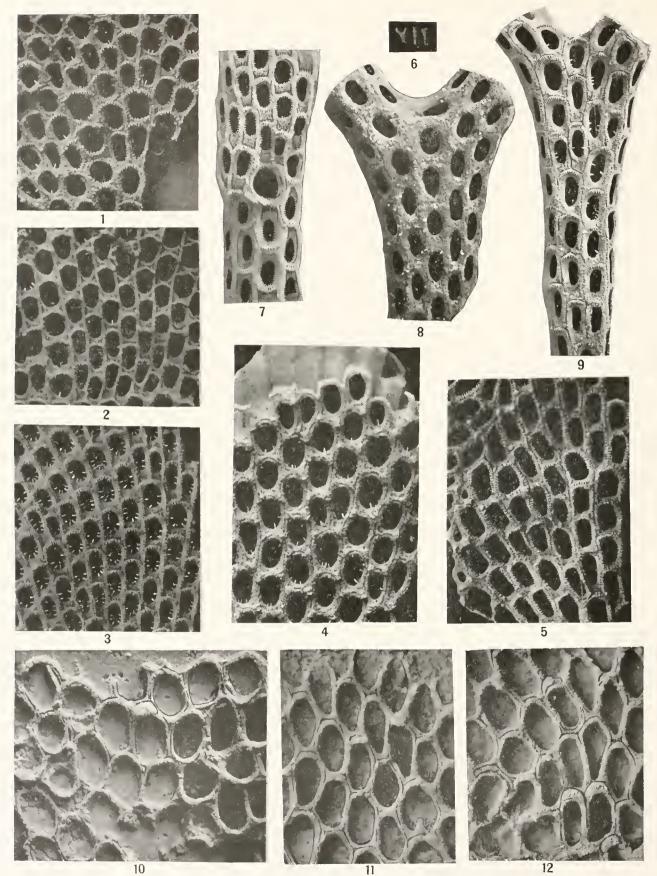
Miocene (Yorktown formation): Near Suffolk, Virginia.

14. Photograph of an inner lamella, \times 20, with the tubercles preserved.

Miocene (Yorktown formation) Yorktown, Virginia.

NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 254.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 255.

PLATE 11.

Figs, 1-3. Acanthodesia savarti Audouin, 1826, forma typica. (p. 31.)

1. An incrusting specimen \times 20, of the typical form of the species, illustrating the considerable micrometric variations. Traces of the spicules in the opesium are visible.

Miocene: Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia.

2. An example < 20, with young zooecia.

3. Surface of a free, multilamellar zoarium, \times 20, with numerous spicules, in the opesium.

Miocene (Yorktown formation): Near Suffolk, Virginia.

Fig. 4. Acanthodesia savarti forma bifoliata Ulrich and Bassler, 1904. (p. 33.)

Surface of the bilamellar type zoarium, \times 20, showing the thin mural rim, the absence of the cryptocyst, the tubercles at the angles and and spicules in the opesium.

Miocene (Choptank formation): Jones Wharf, Maryland.

Figs. 5-9. Acanthodesia savarti forma delicatula Busk, 1859. (p. 33.)

5. An incrusting example, \times 20, referred to this form.

Miocene: Bowler's wharf, 18 miles above Urbana, Virginia.

6. Zoarial fragments, natural size.

7. A branch, \times 20, on which the zooecia are developed in opposite directions.

8. Another branch, \times 20, in which the cryptocyst is small and the superficial ornament is obscured by fossilization.

9. A well preserved branch, \times 20, showing the unequal development of the cryptocyst.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Figs. 10-12. Membrendoccium grande, new species. (p. 36.)

10. Portion of the zoarium \times 20, with irregular zooecia, one showing the endozooecial ovicell.

Miocene (Duplin marl): Wilmington, North Carolina.

11. Marginal, elongated zooecia, \times 20.

12. Another portion of the same zoarium imes 20, with several ovicelled zooecia.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Figs. 1, 2. Membrendoceuni parvicapitatum, new species. (p. 36.)

1. The incrusting type specimen \times 20, with ancestrular, normal, ovicelled and regenerated zooccia.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

2. An Early Miocene specimen, > 20, referred to this species.

Lower Miocene (Bowden marl): Bowden, Jamaica.

Fig. 3. Callopora parvirostris, new species. (p. 41.)

 Λ portion of the type specimen, ≥ 20 , with normal and ovicelled zooccia, exhibiting the small, triangular, suberect avicularium,

Miocene (Calvert formation): I mile south of Parkers Creek, Calvert County, Maryland.

Figs, 4-5. Alderina cesticella, new species. (p. 39.)

4. Ancestrular region of the incrusting zoarium, 5, 20, showing regenerated, calcified and ovicelled zooccia. The ovicells have a prominent collar.

5. Another view of the same zearium, \times 20, exhibiting regenerated zooccia and numerous salient spines on the zooccial walls.

Miocene (Duplin marl); Natural Well, 2 miles southwest Magnolia, Duplin County, North Carolina.

Fig. 6. Ogivalina parvula Ulrich and Bassler, 1904. (p. 37.)

The incrusting type specimen ≤ 20 . A small cryptocyst and often a small convex gymnocyst are present.

Miocene (Calvert formation): Reeds, Maryland.

Fig. 7. Membrani poridra purva, new species. (p. 39.)

The imperfect type specimen, \times 20, with two ovicelled zooecia

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Figs, S. 9. Dakaria torquata D'Orbigny, 1839. (p. 98.)

8. Portion of the incrusting zoarium, × 20, with ovicelled zooccia.

9. Normal zooecia, × 20, illustrating the characteristic broad sinus.

Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia.

Fig. 10. Hippodiplosia bigibbera, new species. (p. 131.)

The incrusting zoarium, \times 20, showing the aperture deformed by two frontal gibbosities.

Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia.

Fig. 11. Callopora lanccolata, new species. (p. 41.)

The incrusting type specimen, \times 20, showing ovicelled and normal zooecia, diatellae and the large lanceolate avicularia.

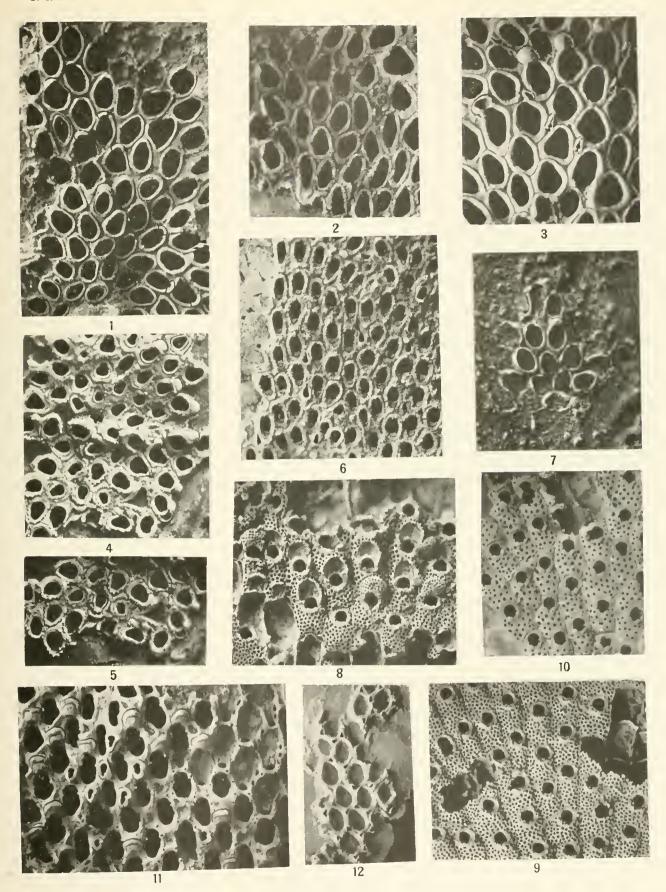
Miocene (Yorktown formation): 3 miles southwest of Petersburg, Virginia.

Fig. 12. Callopora dumerilii Savigny-Andonin, 1826. (See also pl. 1, fig. 2; pl. 2, fig. 23.) (p. 40.)

Photograph of a specimen, \times 20, referred to this species.

Lower Miocene (Chipola marl): Chipola River, Calhoun County, Florida.

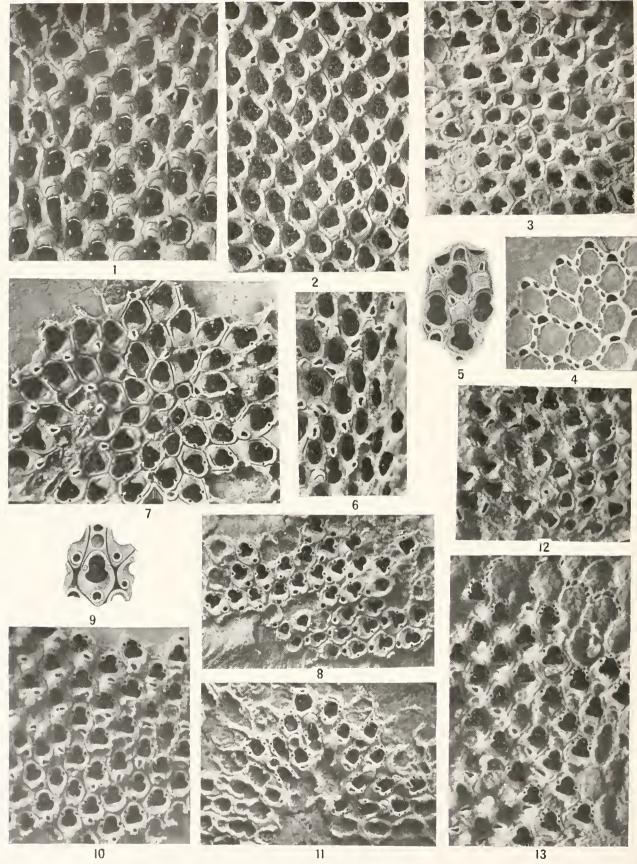
BULLETIN 125 PLATE 12



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 256.

U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

PLATE 13.

Figs. 1-6. Amphiblestrum constrictum Ulrich and Bassler, 1904. (p. 44.)

- 1. Portion of the incrusting zoarium, \times 20, showing numerous ovicelled zooecia, a giant zooecium, and deformed primoserial zooecia.
 - 2. Nonovicelled zooecia, \times 20.
 - 3. Zoarium, × 20. exhibiting the ancestrula, calcified zooecia and regenerated zooecia.
 - 4. Worn zooecia, \times 20, exposing the dietellae.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

- 5. Drawing of several zooecia of the type specimens, \times 17.
- 6. A portion of the type specimen, \times 20, showing zooecia with nontrifoliate opesia. The lateral condules are replaced by serrate denticles.

Miocene (St. Mary's formation): Cove Point, Maryland.

Fig. 7. Amphiblestrum tenuiparietis, new species. (p. 45.)

The incrusting type \times 20, showing the ancestrula and surrounding zooecia, all with thin mural rim.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Figs. 8-10. Ramphonotus agellus Ulrich and Bassler, 1904. (p. 47.)

- 8. The type specimen, \times 20, showing the trifoliate opesium and the avicularia oriented in the zooecial axis.
 - 9. Drawing of a zooecium of the same, more enlarged.

Miocene (St. Mary's formation): Cove Point, Maryland.

10. An example, \times 20, containing both ovicelled and unovicelled zooecia.

Miocene (St. Mary's formation): Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia.

Figs. 11-13. Ramphonotus asperus, new species. (p. 46.)

- 11. Specimen, \times 20, showing ovicelled zooecia, unovicelled zooecia with 6 spines and worn zooecia exposing the dietellae.
 - 12. The incrusting zoarium \times 20, illustrating the ancestrular region with several calcified zooecia.
- 13. Another portion of the same specimen \times 20, showing both ovicelled and unovicelled zooecia, and the dietellae.

Miocene (Yorktown formation) Yorktown, Virginia.

PLATE 14,

Fig. 1. Chaperia partispina, new species. (p. 51.)

The type specimen >, 20, illustrating the absence of large distal spines which are replaced by very small fragile ones.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Figs. 2-4. Chaperia caminosa Ulrich and Bassler, 1904. (p. 54.)

- 2. One of the original types, \times 20, with ovicelled zooecia and the salient triangular avicularium well developed.
 - 3. Another specimen not so well preserved, > 20, showing the ordinary zooecia.

4. View of a zoarium, \times 20, with the ovicells Froken.

Miccene (Choptank formation): Jones Wharf, Maryland.

Fig. 5. Yelumell velongata, new species. (p. 58.)

The incrusting zoarium, \times 20, with large, elongated zooecia, several of which show the ovicell. Miocene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, North Carolina.

Fig. 6. Floridina minima, new species. (p. 57.)

The incrusting zoarium, \times 20, showing both the ancestrular and marginal areas.

Miocene (Duplin marl): Natural Well, 2 miles northwest of Magnolia, North Carolina.

Figs. 7, 8. Floritina regularis, new species. (p. 57.)

7. The external layer of the lamellar incrusting zoarium, \times 20.

Miocene (Duplin marl): Wilmington, North Carolina.

8. A variety of this species, × 20, distinguished by its slightly smaller zooecia.

Miecene (Duplin marl): Natural Well, 2 miles southwest of Magnolia, Duplin County, North Carolina.

Fig. 9. Hemiseptella fistula Ulrich and Bassler, 1904. (p. 73.)

The original type, \times 20, showing the spicules on the inferior border.

Miocene (St. Mary's formation): St. Marys River, Maryland.

Fig. 10. Hemiseptella? lacinia Tuomey and Holmes, 1857. (p. 70.)

The original figure of this doubtful species which has not been rediscovered.

Miocene: Smith's, Goose Creek, South Carolina.

Fig. 11. Hemiseptella rectangulata, new species. (p. 74.)

Photograph of the incrusting zoarium, \times 20, showing the rectangular form of the zoeecia, the minute spinose processes, and the absence of dietellae.

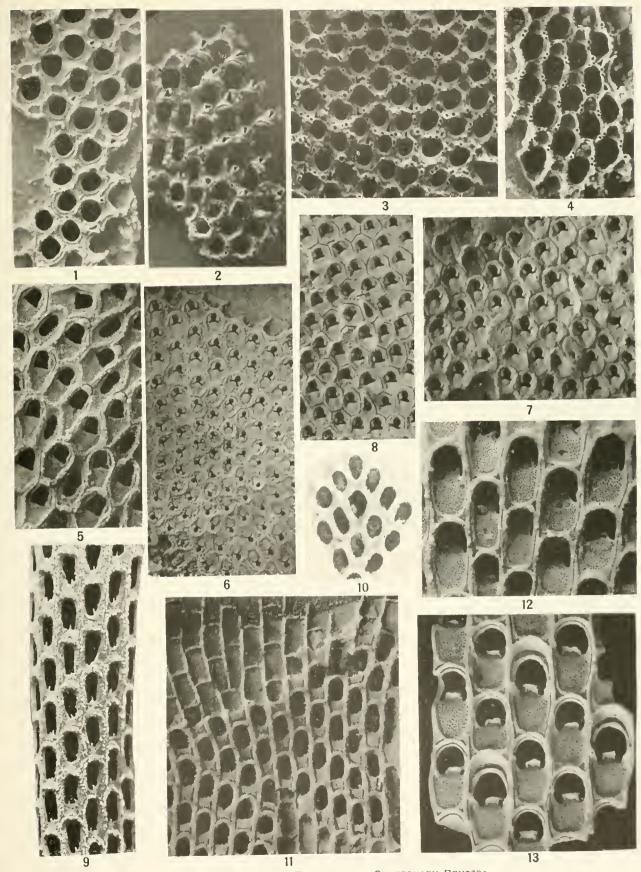
Miocene (Duplin marl): Wilmington, North Carolina.

Figs. 12, 13. Steganoporella magnilabris Busk, 1854. (p. 63.)

12. Surface of a zoarium, \times 20, showing the polypidian tube and the opesiules.

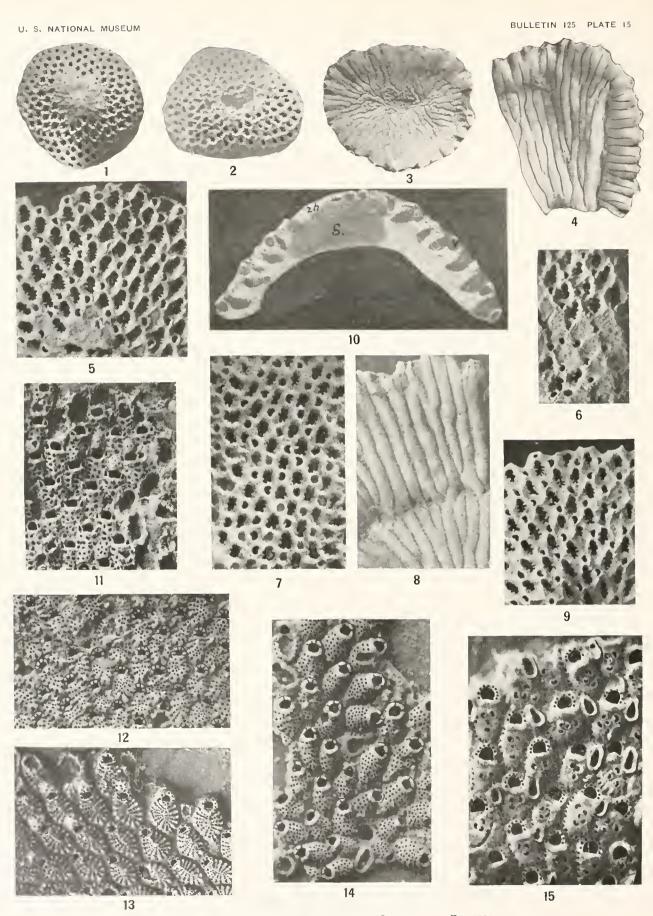
13. An example, \times 20, with both small (a) and large (B) zooecia, and illustrating clearly the large lip under which is a polypidian tube,

Pliocene (Caloosahatchee marl): Monroe County, Florida.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 258



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

- Figs. 1-5. Cupularia doma D'Orbigny, 1851. (See also plate 1, fig. 18.) (p. 77.)
 - 1. A zoarium, \times 10, showing the ancestrular zooecia covering the substratum. These zooecia are calcified and deprived of polypide.
 - 2. Another zoarium \times 10, illustrating that the ancestrular zooccia are easily detached in fossilization.
 - 3. Inner side of zoarium, × 10, which is smooth or slightly granulated.
 - 4. Inner (superior) side of a zoarium with hydrostatic zooecia, × 20. The ribs are smooth and are not radially arranged. This difference in arrangement depends on the form of the substratum on which the larva fixes itself.
 - 5. Celluliferous face \times 20, illustrating the form of the denticles.

Miocene (Duplin marl): Wilmington, North Carolina.

Figs. 6-10. Cupularia denticulata Conrad, 1841. (p. 79.)

- 6. Calcified ancestrular zooecia imes 20. The vibracula alone are visible.
- 7. Center of a zoarium × 20, deprived of calcified zooecia.
- 8. Inner side of a large zoarium \times 20, without hydrostatic zooecia, showing the regular bifurcation of each rib.
- 9. Zooecia \times 20, illustrating the two symmetrical condyles and the vestibular arch above, the auriculated vibracula and the irregular denticles below.
- 10. A median section through a zoarium \times 25, showing the substratum (s) upon which the larva became attached, the calcified hydrostatic zooecia (zh) covering the substratum and the ordinary zooecia surmounted by their vibracula (v).

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Fig. 11. Cribrilina punctata Hassall, 1841. (p. 87.)

The type specimen of *Lepralia marylandica* Ulrich and Bassler, 1904, \times 20, showing its identity with this widespread fossil and recent species.

Miocene (St. Mary's formation): Cove Point, Maryland.

Fig. 12. Puellina radiata forma scripta Reuss, 1847. (See also pl. 35, fig. 1.) (p. 89.)

A well-preserved example, \times 20 with wide zooecia, somewhat approaching ferma rarecosta, Reuss, 1847.

Miocene (Yorktown formation): 1 mile west of Fort Nonsense, Gloucester County, Virginia.

Fig. 13. Puellina innominata Couch, 1844. (p. 90.)

A Miocene example of this widespread species, > 20.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Fig. 14. Cribrilina ligulata, new species. (p. 88.)

The type specimen, \times 25, showing the ancestrula and surrounding zooccia. The spines of the peristome are transformed by coalescence into several small tongues.

Miocene (Calvert formation): 1 mile south of Parkers Creek, Calvert County, Maryland.

Fig. 15. Cribrilina cuspidata, new species. (p. 88.)

The incrusting type specimen, \times 20 showing the line of small lacunae, the few large lacunae and the very large triangular axicularia with rounded leak.

Miocene: Santiago, Cuba.

PLATE 16.

Fig. 1. Trypostego venusta Norman, 1864 (p. 95)

A Miocene example of this widely distributed species, \$\sim 20\$.

Miocene (Puplin marl): Wilmington, North Carolina

Fig. 2. Lacerna mucronata Smitt, 1872. (p. 99.)

 Λ fossil, incrusting example \times 20, referred to this recent species.

Miocene (Duplin marl): Wilmington, North Carolina.

Fig. 3. Arthropoma cornuta, new species. (p. 97.)

The incrusting zoarium, × 20, showing the characteristic sinus and the frontal protul erance.

Miocene (Choctawhatchee marl): Jacl son Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Figs. 4-9. Stephanosella biaperta Michelin, 1842. (p. 99.)

4. The incrusting zoarium, \times 20, illustrating the triangular avicularia.

5. A zoarium, \times 20, with more rounded a vicularia. The tremocyst is detachable from the subjacent olocyst.

6. Interior of the zooecia > 20, illustrating the form of the apertura.

Pleistocene: Rustic Canyon, Santa Monica, California.

7. An example, imes 20, containing both round and triangular axicularia.

Pleistocene: Los Angeles, California.

8. Surface, \times 20, showing several ovicelled zooecia.

Pleistocene: Vero, Florida.

9. A zoarium \times 20 showing the ancestrular zooecia and the thinness of the zooecial walls. The dietellae are very narrow.

Pleistocene: Dead Man's Island off San Pedro, California.

Fig. 10, Schizoporella cumulata Ulrich and Bassler, 1904. (p. 96.)

View of the type specimen, \times 20, showing the irregular arrangement and the piling up of the zooecia.

Miocene (Choptank formation): Jones Wharf, Maryland.

Figs. 11-15. Schizopodrella floridina Osburn, 1914. (p. 106.)

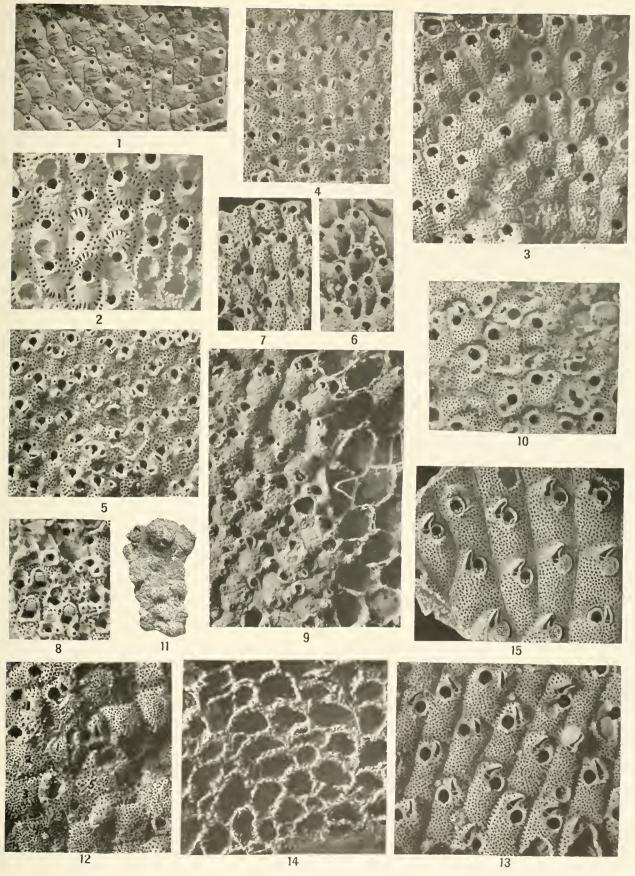
- 11. A zoarium, natural size, composed of many lamellae.
- 12. Non oriented ancestrular zooecia, '× 20.
- -13. Normal aspect of the zooecia, \times 20.—One zooecium has been regenerated with an axicularium occupying the apertura.
 - 14. A transverse section through a multilamellar zoarium, imes 20.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

15. Zooccia \times 20, showing the form of the apertura more clearly. Several of the apertures are closed.

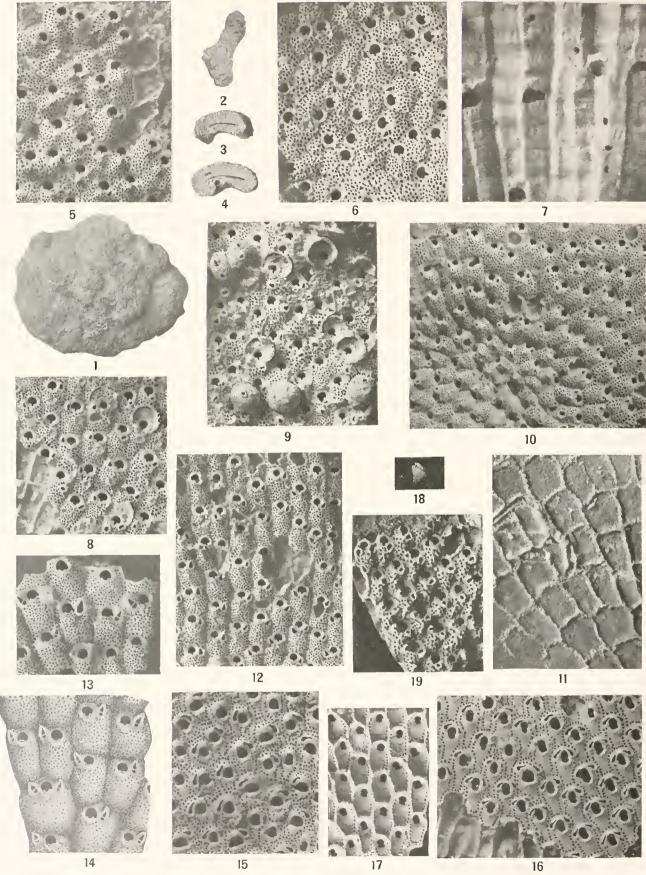
Miocene (Duplin marl): Wilmington, North Carolina.

U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 200.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

PLATE 17.

Figs. 1-12. Stylopoma spongites Pallas, 1766. (p. 102.)

- 1, 2. Two forms of the zoarium, natural size, made up of many lamellae.
- 3, 4. Transverse sections of zoaria, natural size.
- 5. Surface × 20, showing broad unovicelled zooecia.
- 6. Another surface × 20, exhibiting nonoriented zooccia which bear conspicuous avicularia.
- 7. Inferior face of a lamella, \times 20.

Miocene (Duplin marl): Wilmington, North Carolina.

- 8. Normal zooecia with avicularia \times 20. Dietellae are not present. The broken ovicell shows the tremocyst resting on the olocyst.
 - 9. Normal and ovicelled zooecia < 20. The ovicell surrounds and conceals the apertura.
 - 10. Ancestrular portion of a zoarium, \times 20.
 - 11. Zooecial walls \times 20, obtained by abrasion of the surface, showing arrangement of the septules.
 - 12. A portion of a zoarium < 20, with quite elongated zooecia.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida.

Figs. 13, 14. Schizopodrella unicornis Johnston, 1847. (p. 105.)

- 13. Photograph of a well-preserved Miocene specimen, \times 20, showing the form of the apertura and the avicularia clearly.
- 14. Zooecia of the type specimen of $Schizoporella\ subquadrata\$ Uhrich and Bassler, 1904, now considered as a synonym of $S.\ unicornis.$

Miocene (Choptank formation): Governor Run, Maryland.

Figs. 15-17. Hippaliosina rostrigera Smitt, 1872. (p. 167.)

15. Zooecia, \times 20, of a form exhibiting only one avicularium above the aperture.

Miocene (Duplin marl): 1 mile northeast Suffolk, Virginia.

- 16. The normal form of the species, \times 20, with two avicularia.
- 17. Interior of zooecia, \times 20, illustrating the two cardelles and the olocyst.

Pliocene (Caloosahatchee marl): Monroe County, Florida.

Figs. 18-19. Schizopodrella pusilla, new species. (p. 406.)

- 18. The free bilamellar zoarium, natural size.
- 19. Surface, \times 20, showing the small zooecia with large tremopores.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

PLATE 18.

Ftgs. 1-3. Schizop Arella doverensis Ulrich and Bassler, 1904. (p. 107.)

1. A Pliocene example, \times 20, referred to this species.

2. Ancestrular portion of a zoarium, × 20, showing irregular arrangement of zooccia.

Pliocene (Calobahatchee marl): Shell Creek, De Soto County, Florida.

3. The incrusting type of the species, \times 20.

Miocene (Choptank formation): Dover Bridge, Maryland.

Fig. 4. Schizoporella latisimuata Ulrich and Bassler, 1904. (p. 95.)

Photograph of the original incrusting type, \times 20.

Miocene (Choptank formation): Jones Wharf, Maryland.

Figs. 5, 6. Gemellip rella asper, new species. (p. 110).

5. The incrusting zoarium, \times 20, with oriented zooecia and showing both the large and small avicularia.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

6. A Pliocene example, $\times 20$, illustrating nonoriented zooecia, several of which preserve the ovicell.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Figs. 7-9. Hippoporina (?) vestita, new species. (p. 130.)

7. View of the incrusting zoarium, × 20, with the zooecia more distinctly marked than usual.

8. An example, \times 20, showing at the base the calcareous pellicle covering the frontal and at the top, monstrous zooccia. All of the zooccia have indistinct outlines.

9. An example, \times 20, preserving some ovicelled zooecia.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahasse, Florida.

Fig. 10. Hippoporma gibbosa, new species. (p. 120.)

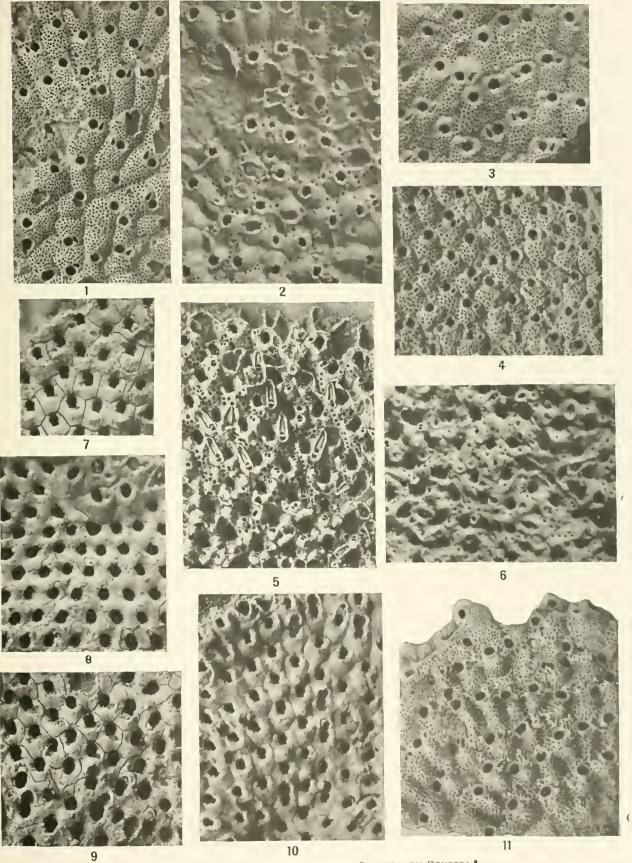
The incrusting type specimen, \times 20, showing the smooth frontal with its gibbosity, the shape of the aperture and the widely open ovicell.

Miocene (Duplin marl): Wilmington, North Carolina.

Fig. 11. Lepcalia montifera Ulrich and Bassler, 1904. (p. 134.)

Photograph of the original type, \times 20.

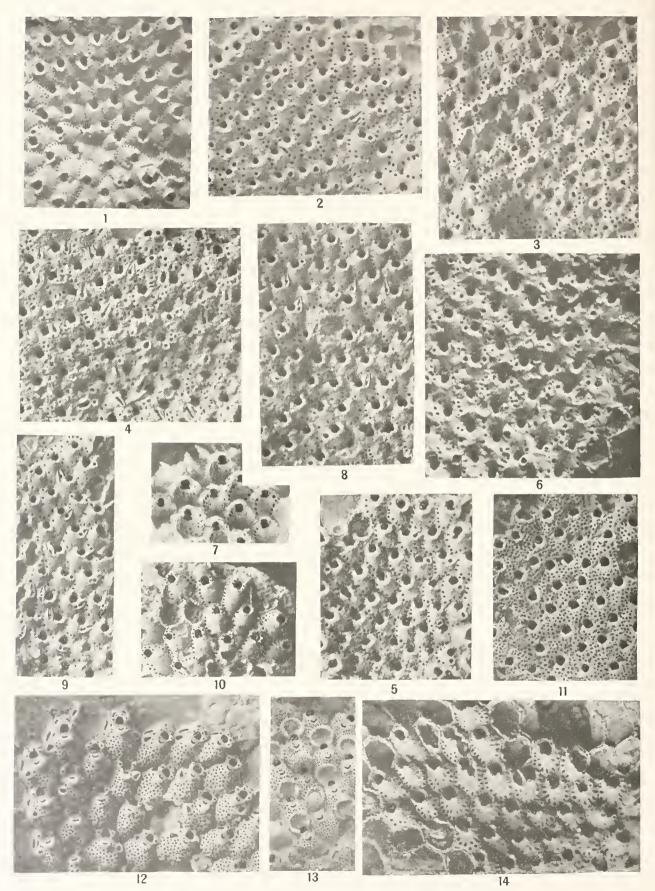
Miocene (St. Mary's formation): St. Marys River, Maryland.



North American Later Tertiary and Quaternary Bryozoa.

FOR EXPLANATION OF PLATE SEE PAGE 262.

BULLETIN 125 PLATE 19



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

Plate 19

Figs. 1-9. Gemelliporella vorax, new species. (p. 111.)

- 1. Ancestrular portion of the incrusting zoarium \times 20. The ancestrula is covered by the pleurocyst of the adjacent zooccia.
- 2. Zooecia, <20, with orbicular axicularia. The arcolar pores and granular pleurocyst are evident.
 - 3. A zoarium, imes 20, with indistinct zooecia and orbicular axicularia.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

- Zooecia, × 20, with large spatulate avicularia. The areolae are limited by short costules.
- 5. Zooecia, \times 20, with large areolar pores and the granulated pleurocyst shown.
- 6. Portion of a large zoarium, × 20, with indistinct zooecia and orbicular avicularia.
- 7. Interior of the zooecia, \times 20, showing the olocyst which supports the pleurocyst.

Miocene (Duplin marl): Wilmington, North Carolina.

8. Zooecia, imes 20, with both large and small avicularia.

Pliocene (Caloosahatchee marl): Monroe County, Florida.

9. A zooecia of a young zoarium, \times 20, with large spatulate avicularia.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida.

Fig. 10. Hippoporella spinosa, new species. (p. 132.)

The incrusting type specimen + 20, showing the smooth frontal with small lateral areolar pores and the spines of the peristome.

Miocene (Yorktown formation): Yorktown, Virginia.

Fig. 11. Cyclicopora? mansfieldi, new species. (p. 138.)

Unovicelled zooecia of the incrusting zoarium, >. 20, showing the suborbicular aperture and the numerous large tremopores.

Miocene (Yorktown formation): 11 miles southwest of Reeds Ferry, Virginia.

Figs. 12, 13. Microporella fissurifera, new species. (p. 121.)

12. Unovicelled zooecia of the incrusting zoarium · 20, exhibiting the dietellae visible along the edge of the zooecia.

Miocene (Yorktown formation): Near Macedonia Church, Essex County, Virginia.

13. Ovicelled zooecia, \times 20.

Miocene (Duplin marl): 10 miles south of Greenville, North Carolina.

Fig. 14. Hippoporella papulifera, new species. (p. 133.)

Unovicelled zooecia of the incrusting zoarium, ≥ 20 , with distinct areolar pores and the characteristic mucro.

Miocene (Choctawhatchee marl). Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

PLATE 20,

Figs. 1-6. Microporella ciliata Linnaeus, 1759. (See also pl. 36, figs. 4, 5.) (p. 119.)

1. Incrusting portion of zoarium, +20, showing the typical zooccia.

2. Bilamellar expansion of the same zoarium, \times 20. The zooccia have large tremopores and no granules.

Miocene: Kuhns, Carteret County, North Carolina.

3. Typical zooecia, × 20, with their frontal provided with granules and small tremopores.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

- 4. The original type specimen of *Microporella pracciliata* Ulrich and Bassler, 1904, \times 28, hitherto distinguished on account of its large tremopores.
 - 5. Several zooecia of the same, further enlarged.

Miocene (Choptank formation): Jones Wharf, Maryland.

6. The type specimen of $Micropovella~in~\theta ata$ Ulrich and Bassler, 1904, now considered a synonym of M.~ciliata.

Miocene (Choptank formation): Jones Wharf, Maryland.

Figs. 7-11. Microporella bifoliata Ulrich and Bassler, 1904. (p. 121.)

- 7. Photograph of the bifoliate type specimen, ≤ 20 , with both oxicelled and unovicelled zooecia,
- 8. A drawing of two ovicelled zooecia.
- 9. A zooecium and an avicularium highly magnified.
- 10. Mature unovicelled zooecia, \times 20.
- 11. Unovicelled zooecia, × 20, illustrating appearance in young specimen.

Miocene (Choptank formation): Cordova, Maryland,

Figs. 12, 43. Cyclocolposa (?) spinifera new species. (p. 136.)

- 12. The incrusting zoarium, \times 20, with ancestrular zooecia. The pleurocyst is incompletely developed.
- 13. A zoarium with mature zooccia, ~ 20 , showing the very thick frontal and the globular smooth ovicell.

Miocene (Yorktown formation): 1 mile west of Fort Nonsense, Gloucester County, Virginia.

Figs. 14, 15, Microporella heragona, new species. (p. 120.)

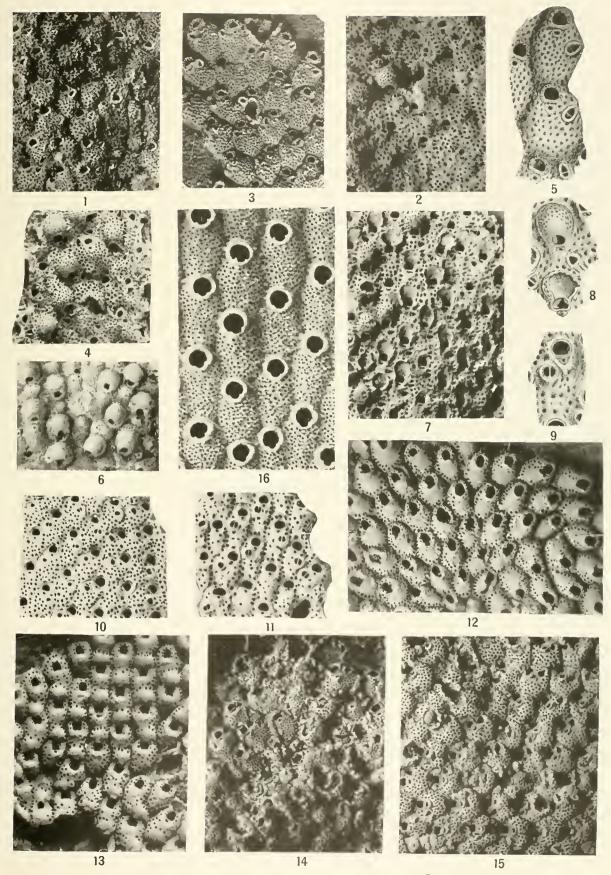
- 14. The incrusting zoarinm, × 20, showing the strongly calcified hexagonal zooccia.
- 15. Another portion of the same specimen, \times 20, illustrating the hexagonal shape of the zooeeia, the large ascopore with its salient peristome and the triangular avicularia.

Miocene (Duplin marl): Darlington Courthouse, South Carolina.

Fig. 16. Dakaria grandis, new species. (p. 97.)

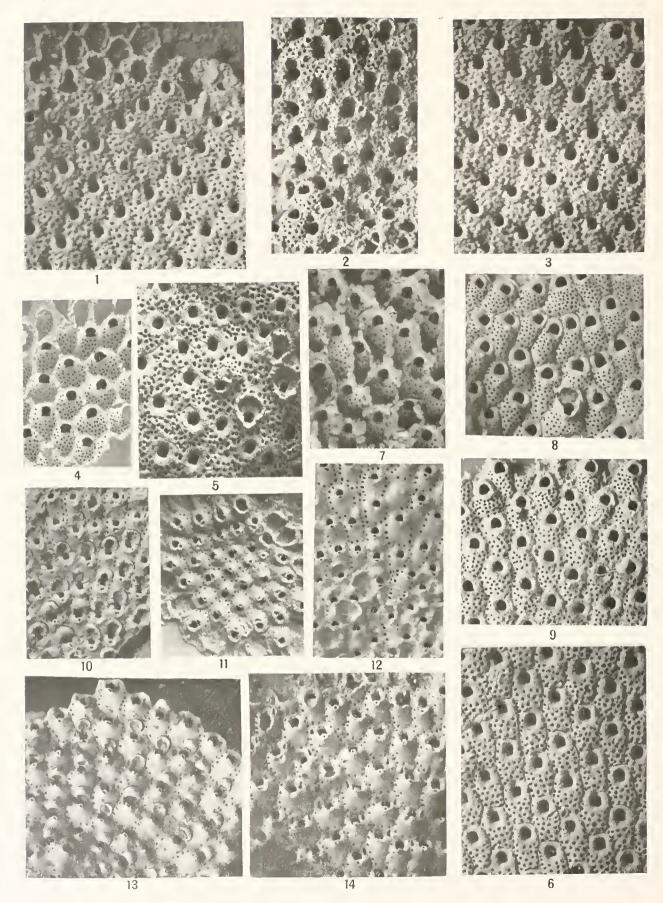
The incrusting zoarium, - 20.

Miocene (Yorktown formation): Yorktown, Virginia.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 264.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

PLATE 21.

Figs. 1-4. Cyclocol posa-tenui parietis, new species. (p. 136.

- 1. The incrusting zoarium, \times 20, with some of the zooecia bearing a round avicularium. The thinness of the zooecial walls may be noted.
 - 2. Ovicelled zooecia, × 20, showing the ovicell deeply imbedded in the distal zooecium.
- 3. Zooecia, \times 20, exhibiting regular arrangement, the two rows of areolae and the granulated pleurocyst.
 - 4. Interior of the zooccia, × 20, illustrating the condyles on each side of the apertura.

Miocene (Duplin marl): Harvey's Mills, Leon County, Florida.

Figs. 5-9, Cyclopericlla rubra, new species. (p. 137.)

- 5. Surface of an incrusting lamella, < 20, with several ovicelled zooecia, showing that the ovicell entirely covers the apertura.
 - 6. Normal zooecia, < 20.
- 7. Interior of the zooecia, \times 20, illustrating the two lateral condyles on which the operculum * oscillates.

Miocene (Duplin marl): Wilmington, North Carolina.

8. Zooecia, \times 20, preserving a frontal gibbosity.

Miocene (Duplin marl): 8 miles east of Snow Hill, North Carolina.

9. A Pliocene example of this species, \times 20; the zooecia are slightly less regular than in the Miocene form.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Figs. 10-14. Aimulosia aculeata, new species. (p. 139.)

10. The incrusting zoarium with ovicelled zooecia, \times 20.

Miocene (Duplin marl): Harvey's Mills, Leon County, Florida.

11. The ancestrula and surrounding zooecia, \times 20. The first zooecium is an avicularium while the other four zooecia are normal.

Miocene (Duplin marl): Muldrow's Mills, South Carolina.

12. A very irregular incrusting zoarium, \times 20.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

13. Zoarium with ovicelled zooccia, imes 20. The orifice of the ovicell is distinct from the apertura.

14. Broad marginal zooecia, × 20, without ovicell.

Miocene (Duplin marl): Wilmington, North Carolina,

PLATE 22.

Figs. 1-6. Leiosella edax, new species. (p. 142.)

- 1. Fragments of the free unilamellar zoarium, natural size.
- 2. A tubular zoarium, \times 20, with the zooecia exhibiting the large rimule spiramen.
- 3. Ovicelled zooecia, \times 20.
- 4. Zooecia, \times 20, in which the small frontal avicularia are replaced by a large avicularium with spatulate mandibles.
- 5. A tubular zoarium, $\lesssim 20$, showing zooecia provided with an irregular rimule spiramen and with small avicularia.
 - 6. Interior of zooecia, < 20, showing the apertura and the peristomial avicularium.

Lower Miocene (Chipola marl): Chipola River, Calhoun County, Florida.

Figs. 7-14. Smittina trispinosa Johnston, 1838. (p. 143.)

- 7. Unovicelled zooccia of a well preserved zoarium, \times 20, with the avicularium small or absent
- 8. Ancestrula and surrounding zooecia of the same zoarium, \times 20.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

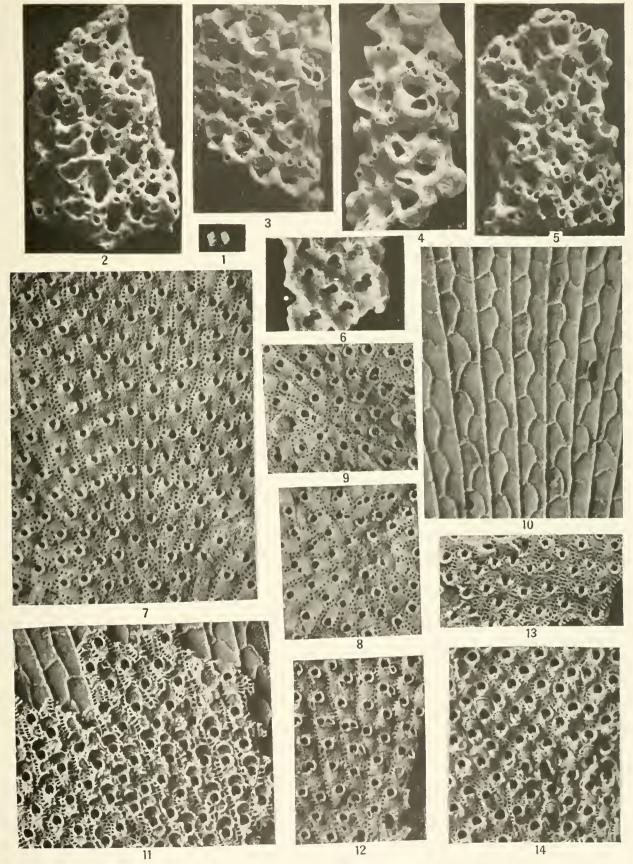
9. Ancestrula region of a Miocene example, \times 20.

Miocene (Duplin marl): Wilmington, North Carolina.

- 10. An abraded surface, \times 20, showing the alternate arrangement of the zooecia between two rectilinear axes without septulae.
- 11. An ovicelled zoarium, \times 20, in which the ovicells are broken. The alternate arrangement of the zooccia is visible.
- 12. Zooecia, \times 20, with salient avicularia.
- 13. Zooecia, \times 20, in which the peristome is formed by two lateral salient lips.
- 14. Zooecia, \times 20, with well-preserved ovicells.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida.

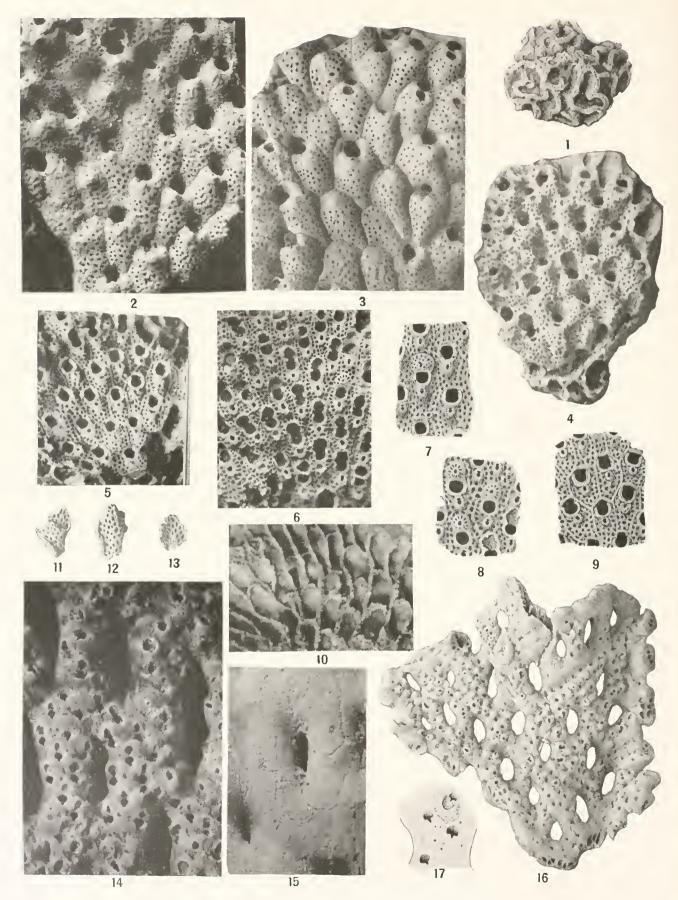
U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 266.

BULLETIN 125 PLATE 23



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 267.

PLATE 23.

- Figs. 1-3. Porella (Palmicellaria) convoluta Ulrich and Bassler, 1904. (p. 150.)
 - 1. The convoluted, bifoliate zoarium, natural size.
 - 2. Zooecia, \times 20, showing the prominent mucro and the avicularium.
 - 3. Surface of another fragment, \times 20, with several closed zooecia.

Miocene (Calvert formation): Reeds, Maryland.

Fig. 4. Porella (Palmicellaria) punctata Ulrich and Bassler, 1904. (p. 150.)

The bifoliate type specimen, \times 20.

Miocene (Calvert formation): Reeds, Maryland.

Figs. 5-10. Porella reversa Ulrich and Bassler, 1904. (p. 148.)

- 5. Portion of the incrusting zoarium, \times 20, showing the zooecia with thin raised borders.
- 6. Another zoarium, \times 20, with ovicelled zooecia.
- 7. Drawing showing usual aspect of the ovicelled and unovicelled zooecia, \times 28.5.
- 8. Zooecia of a specimen with ovicells, \times 28.5.
- 9. Several zooecia, × 28.5, with the boundaries well defined.

Miocene (St. Mary's formation): Cove Point, Maryland.

10. A view of the zooecial walls after abrasion of the frontal, \times 20.

Miocene: Kuhns, Carteret County, North Carolina.

- Figs. 11-17. Retepora doverensis Uhrich and Bassler, 1904. (p. 152)
 - 11-13. Three specimens, natural size.
 - 14. Portion of a type specimen, \times 20, showing the fissured ovicell and the wide irregular rimule piramen.
 - 15. Noncelluliferous side of a zoarium, × 20.
 - 16. One of the original types, \times 8, with ovicelled zooecia.
 - 17. Drawing of several zooecia, \times 30.

Miocene (Choptank formation): Dover Bridge, Maryland.

PLATE 24.

Figs. 1, 2. Advona heckeli Reuss, 1847. (p. 158.)

1. Surface of zoarium, × 20, showing convex, young zooccia.

2. The normal aspect of the zourium, × 20, in which the zopecia are flat.

Miocene (Duplin marl): Wilmington, North Carolina.

Figs. 3, 4. Tremogasterina horrida, new species. (p. 168.)

3. The free, bilamellar zoarium, natural size.

4. Zooecial surface, < 20, illustrating the large median orbicular pore surrounded by a line of arcolar pores, and the immense triangular axicularium.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

Figs. 5-8. Adeonellopsis cocrinella, new species. (p. 161.)

5. Fragments of the bilamellar zoarium, natural size.

6. Surface of the bilamellar zoarium, \times 20, with poorly oriented zooecia.

7. Another surface, \times 20, with more regularly arranged zooccia and showing the frontal perforated by stellate pores.

8. Interior of zooecia, \times 20, illustrating the frontal perforations.

Lower Miocene (Chipola marl): Chipola River, Calhoun County, Florida.

Fig. 9. Lagenipora(?) brevicollis, new species. (p. 171.)

Zooecia of the incrusting zoarium, \times 20, illustrating the form of the apertura, the tremopores, and the occasional small avicularia.

Miocene (Duplin marl): Kuhus, Carteret County, North Carolina

Figs. 10-13. Holoporella parvula, new species. (p. 175.)

10. The free irregularly branched zoarium, natural size.

11. Surface, \times 20, illustrating the very small, cumulate zooecia.

12. Another portion of the type, \times 20, showing the small umbo before the apertura more clearly.

13. Zooccia, \times 20, with areolar pores distinctly visible.

Miocene (Duplin marl): Cape Fear River, 28 miles northwest of Wilmington, North Carolina.

Fig. 14. Holoporella rostrifera, new species. (p. 175.)

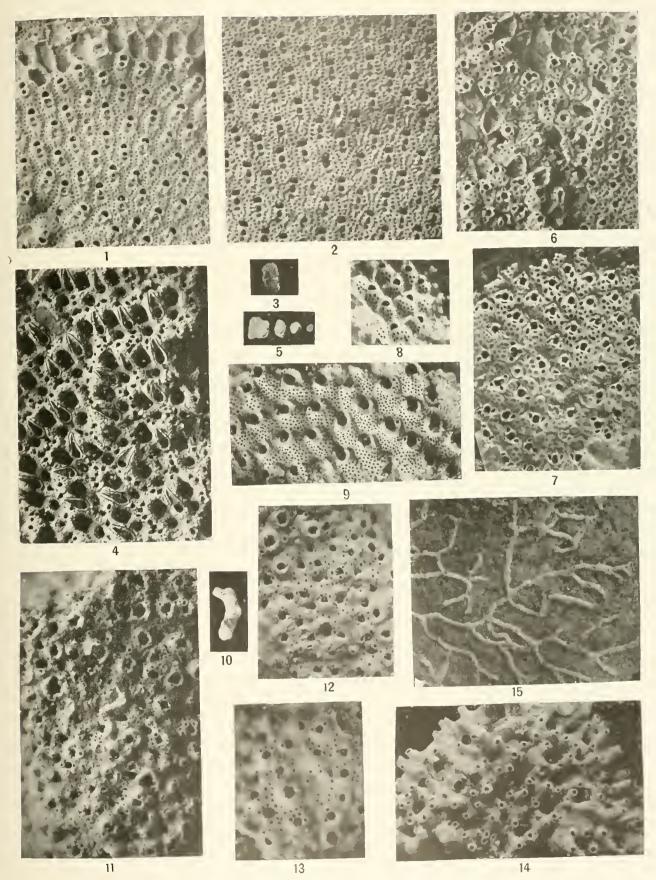
The incrusting zoarium, \times 20, exhibiting the erect, indistinct zooccia with the four salient rostra about the apertura.

Miocene (Yorktown formation): York River, Virginia.

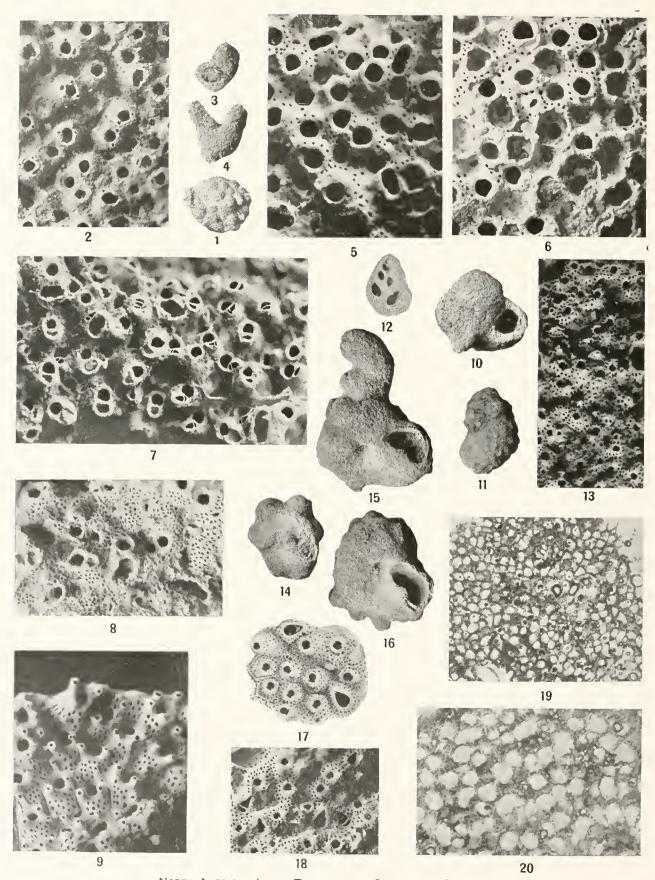
Fig. 15. Aetea anguina (?) Linnaeus, 1758. (p. 49.)

The incrusting network, \times 20, referred doubtfully to this recent species.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

Plate 25.

Figs. 1, 2. Holoporella (?) echinata, new species. (p. 177.)

1. The globular zoarium, natural size.

2. Zooecia, × 20, showing two oral spines and the absence of areolar pores.

Miocene (Yorktown formation): Yorktown , Virginia.

Figs. 3-6. Holoporella orbifera, new species. (p. 177.)

3, 4. Two examples of the irregular, massive zoaria, natural size.

5. Portion of the zoarium, × 20, with large interzooccial avicularia. The areolar pores, the convex frontal, and the form of the apertura are well shown.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

6. Another surface, \times 20, illustrating the usual characters noted.

Miocene (Duplin marl): Wilmington, North Carolina.

Fig. 7. Holoporella massalis, Ulrich and Bassler, 1904. (p. 177.)

Surface of the original type, \times 20.

Miocene (St. Mary's formation): St. Marys River, Maryland.

Fig. 8. Cellepora cribrosa Ulrich and Bassler, 1904. (p. 183.)

Photograph of the type specimen, \times 20.

Miocene (Calvert formation): Reeds, Maryland.

Fig. 9. Holoporella subturrita, new species. (p. 176.)

The incrusting zoarium, × 20, illustrating the four prominent rostra with avicularia, about each

Miocene (Duplin marl): Twenty-eight miles northwest of Wilmington, North Carolina.

Figs. 10-13. Cellepora minuta, new species. (p. 182.)

10-12. Two zoaria, natural size, with a third cut through the middle, showing growth upon gastropod shells.

13. Surface of a zoarium, \times 20, illustrating the small size of the zooecia, a single row of areolar pores and the occurrence of small tubular avicularia only.

Miocene (Duplin marl): Wilmington, North Carolina.

Figs. 14-20. Cellepora maculata Ulrich and Bassler, 1904. (p. 182.)

14-16. Three zoaria, natural size, growing as usual upon gastropod shells.

17. Drawing of the surface, imes 17, illustrating the shape of the apertura and the two kinds of avicularia as well as the areolar pores.

18. Photograph of zooecia, < 20, illustrating the large interzooecial avicularia.

19. A section through a zoarium, \times 10, showing the accumulation of the zooecia.

20. An enlargement of the same, \times 25, showing the arrangement and structure in more detail,

Miocene (Calvert formation): Plum Point, Maryland.

PLATE 26.

Figs. 1-3. Ceriopora virginiana, new species. (p. 193.)

1. The irregular globular zoarium, natural size.

2. Surface, × 12, showing the thin walls of the polygonal zooccia.

3. Portion of the same, \times 25.

Miocene (Yorktown formation): One mile northeast of Suffolk, Virginia.

Figs. 4-6. Atclesopora reptans, new species. (p. 191.)

4. Zoaria, natural size, incrusting a shell.

5. A complete zoarium, \times 12, with the zone of growth showing small lobes.

6. A narrow flabellate zoarium, >, 12, with a thick zone of growth.

Miocene (Duplin marl): Muldrows Mills, 5 miles south of Mayville, Sumter County, South Carolina.

Fig. 7. Proboscina mesleri, new species. (p. 193.)

The type specimen, \times 12, showing the long branches with three rows of zooecia.

Miocene (Duplin marl): Wilmington, North Carolina.

Fig. 8. Idmonea(?) expansa Ulrich and Bassler, 1904. (p. 200.)

The irregular, flabellate, incrusting zoarium, + 12.

Miocene (St. Mary's formation): Cove Point, Maryland.

Figs. 9-12. Theonou glomerata Ulrich and Bassler, 1904. (p. 201.)

9, 10. A large specimen, natural size, and a photograph, \times 4.

11. A small specimen, natural size.

12. Another view of the same, \times 4.

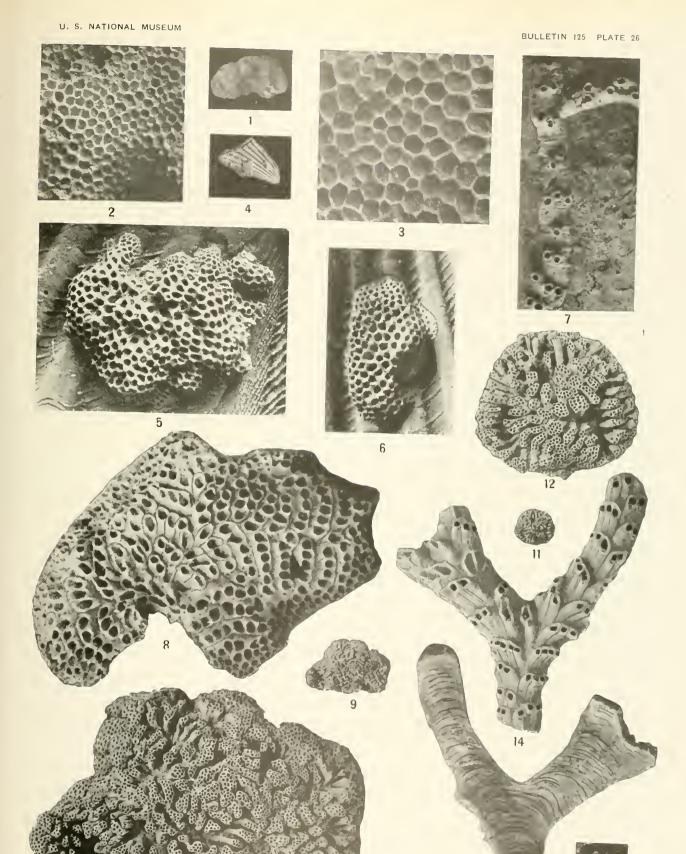
Figs. 13-15. Idmonea planula, new species. (p. 199.)

13. The free zoarium, natural size.

14. Anterior side, \times 12, showing the salient fascicles closely arranged and opposite to each other.

15. Posterior side of the same specimen, × 12, illustrating the flat, transversely striated dorsal.

Miocene (Choctawhatchee marl): Jackson Bluff, Ocklocknee River, 25 miles southwest of Tallahassee, Florida.

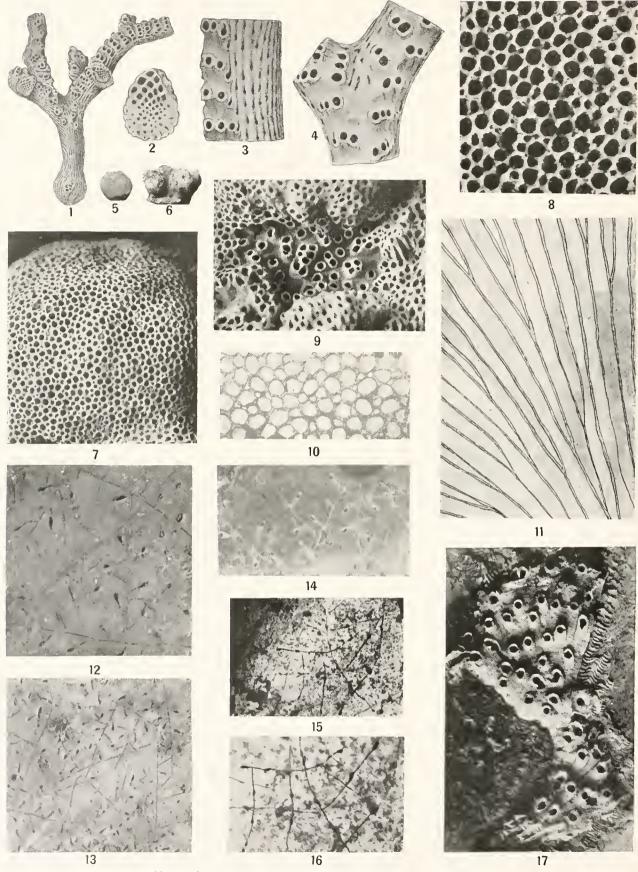


NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

10

15

FOR EXPLANATION OF PLATE SEE PAGE 270.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 271

PLATE 27.

Figs. 1-4. Crisina striatopora Ulrich and Bassler, 1904. (p. 200.)

1. The type specimen, \times 5.

2. View of the end of a branch, \times 17.

3. Lateral view of one of the branches, \times 23.

4. Upper surface of the same branch, \times 17.

Miocene (Choptank formation): Jones Wharf, Maryland.

Figs. 5-11. Tretocycloecia avellana, new species. (p. 206.)

5-6. Two specimens natural size, showing variations in the free spherical zoarinm.

7. Surface of a globular specimen, \times 12.

8. Zoarial surface, \times 25.

9. Surface, \times 12, showing that the ovicell is large and irregular.

10. A tangential thin section, \times 25.

11. A longitudinal thin section, \times 25, showing the hollow walls and the absence of diaphragms. Miocene (Duplin marl): one-half mile above Edenhouse Point, Chowan River, North Carolina.

Figs. 12-13. Spathipora longicauda, new species. (p. 16.)

A portion of the type specimen, \times 20 and \times 10, illustrating the elongated fusiform zooecia with a long peduncle attached to the canalicules at an angle of about 45°.

Miocene (St. Mary's formation): Bowler's wharf, 18 miles above Urbana, Middlesex County, Virginia.

Fig. 14. Spathipora cucullata, new species. (p. 16.)

The type specimen, \times 20, showing the zooecia embedded in the shell substance.

Miocene (Yorktown formation): Beulahland, Virginia.

Figs. 15-16. Terebripora parvicella, new species. (p. 15.)

Two views of the type specimen, \times 10 and \times 20, illustrating the very thin canalicules branching almost at right angles and the slightly oblique arrangement of the zooecia.

Miocene (Duplin marl): two miles southwest of Magnolia, North Carolina.

Fig. 17. Berenicea flabellum? Reuss, 1847. (p. 194.)

The American example, \times 12, referred doubtfully to this European species.

Miocene (Yorktown formation): Weavers Pond, Gloucester County, Virginia.

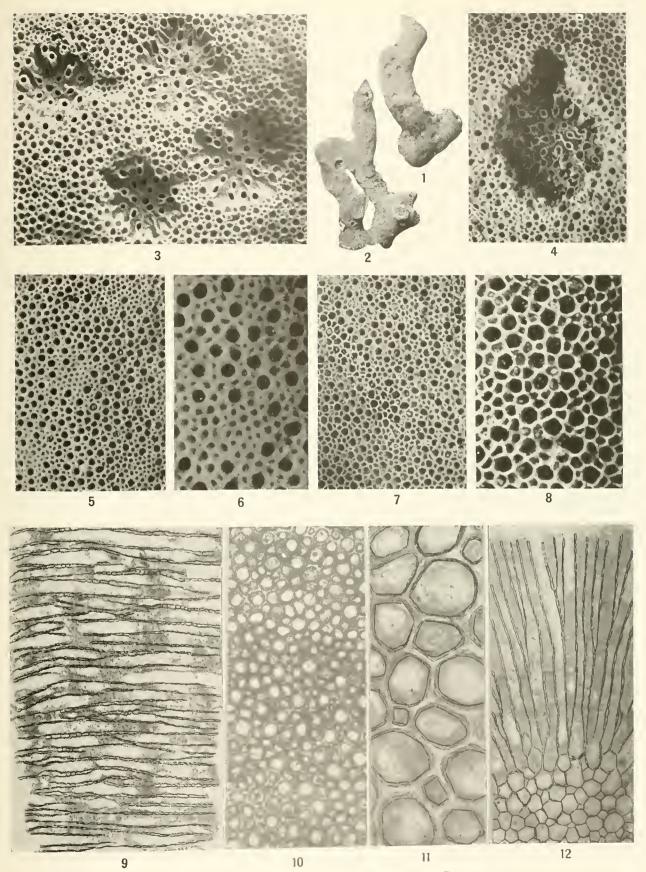
PLATE 28.

Figs. 1-12. Tretocycloccia tortilis Lonsdale, 1845. (p. 206.)

- 1, 2. Fragments of the irregularly branched, cylindrical zoarium, natural size.
- 3. Surface of a specimen, \times 12, with a group of broken ovicells. The superior layer of the ovicells has disappeared in fossilization.
 - 4. A large isolated ovicell, \times 12, with its upper surface broken away.
- 5. Surface of ordinary zeoecia, \times 12, showing the mesopores grouped together in many parts of the branch.
- 6. Same surface, + 25, illustrating the thickness of the zooecial walls.
- 7, 8. Another surface, \times 12 and a portion of the same, \times 25, in which the zooecia have thin walls and the mesopores are arranged irregularly between the apertures.
 - 9. Outer portion of longitudinal thin section, \times 25.
 - 10. Tangential thin section, \times 25.
- 11. Several zooecia of the same, \times 100, showing that the internal walls are much calcified and are separated by a less dense tissue.
 - 12. Portion of a transverse thin section, \times 25.

Miocene (Yorktown formation): Yorktown, Virginia.

U. S. NATIONAL MUSEUM

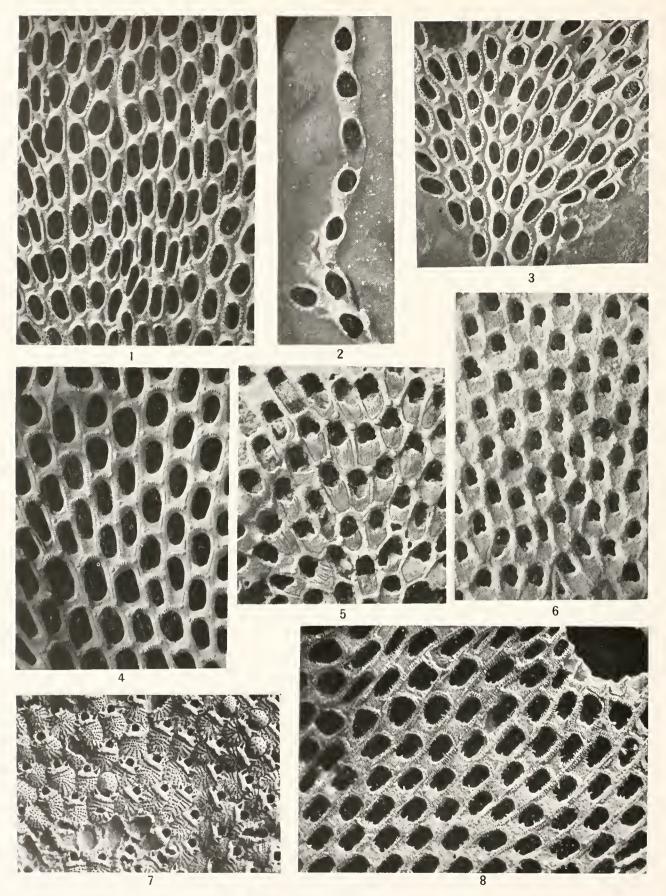


North American Later Tertiary and Quaternary Bryozoa.

FOR EXPLANATION OF PLATE SEE PAGE 272.

U. S. NATIONAL MUSEUM

BULLETIN 125 PLATE 29



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

PLATE 29.

Figs. 1-3. Electra monostachys Busk, 1854. (p. 17.)

1. An expanded zoarium, \times 20.

Pleistocene: Wailes Bluff, near Cornfield Harbor, St. Marys County, Maryland.

2. Unilinear series of zooccia, × 20, with zooccia somewhat larger than usual.

Pleistocene: Santa Barbara, California.

3. A flabelliform zoarium, \times 20.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Fig. 4. Membranipora lacroixii Audouin, 1826. (p. 22.)

The incrusting zoarium, \times 20, showing the finely striated mural rims and the small tubercle at each interzooccial angle.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Figs. 5, 6. Hemiseptella planulata, new species. (p. 74.)

5. Portion of the incrusting zoarium, \times 20, somewhat worn but showing the flat cryptocyst and the tubercles at the interzooccial angles.

6. A better preserved example, \times 20, with the spinous processes visible.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Fig. 7. Puellina crassilabiata, new species. (p. 91.)

The incrusting zoarium, \times 20, showing the thick transverse lip along the edge of the apertura. Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Fig. 8. Hemiseptella granulosa, new species. (p. 74.)

The unilamellar zoarium, \times 20, exhibiting the characteristic irregular opesial proximal border with spinous processes and the granulose mural rim.

Miocene: Charleston, South Carolina.

PLATE 30.

Fig. 1. Dakaria parciporosa, new species. (p. 98.)

The incrusting zoarium, \times 20, exhibiting the form of aperture and the minute tre nopores.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Figs. 2, 3. Microporella tessellata Tuomey and Holmes, 1857. (p. 122.)

2. Zooecia, × 20, showing a large avicularium placed lower than the apertura.

3. Ancestrular region, \times 20, with several worn zooecia exhibiting the dietellae.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Figs. 4, 5. Schizopodrella marginata, new species. (p. 107.)

4. Usual aspect of the incrusting zoarium, \times 20.

5. Zooecia, < 20, with the marginal rims worn away.

Phocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

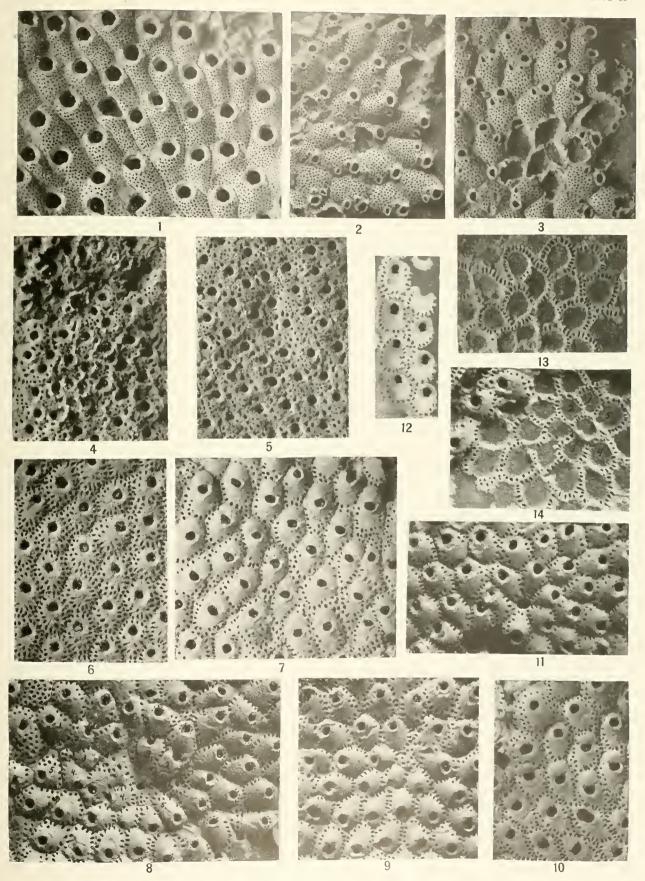
Figs. 6-14. Cyclocolposa perforata, new species. (p. 135.)

6. Normal zooccia of the incrusting zoarium, \times 20.

7. Ovicelled zooccia, \times 20.

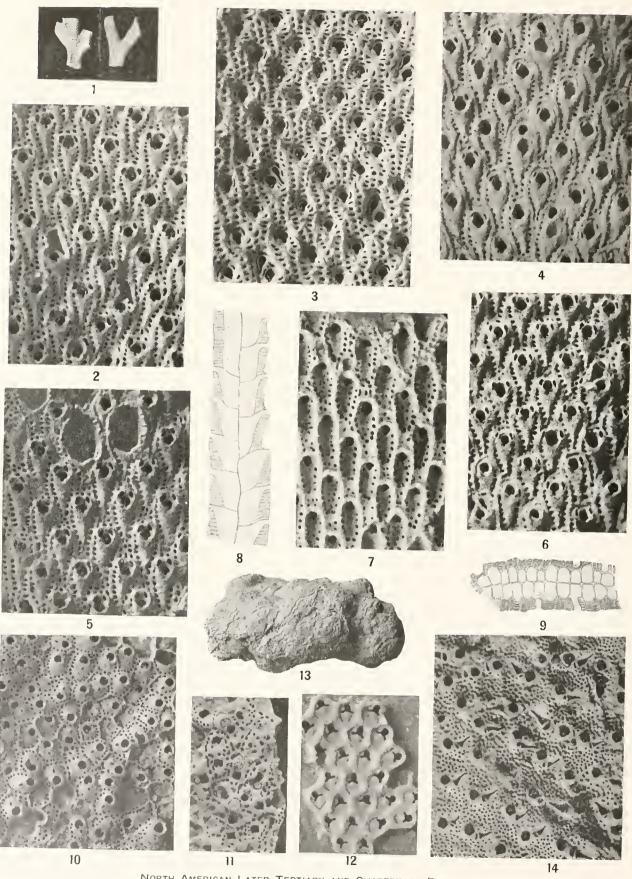
- 8. Portion of a zoarium, \times 20, showing two ancestrular areas with their surrounding zooecia (numbered 1, 2, 3, 4, 5).
 - 9. Ancestrular region, ×20, with the ancestrula covered by the pleurocyst of the adjacent zooecia
 - 10. Zooecia, \times 20, exhibiting the olocyst and illustrating that the pleurocyst is detachable.
 - 11. Ancestrular region, \times 20. The ancestrula is reduced to its apertura.
 - 12. Interior of the zooecia, × 20, exhibiting the olocyst.
 - 13. Worn zooecia, × 20, showing the dietellae.
- 14. Ancestrular region, \times 20, with surface worn away and illustrating the arrangement of the zooecia around the ancestrula.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SHE PAGE 274



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 275.

PLATE 31.

Figs. 1-9. Metrarabdotos auriculatum, new species. (p. 164.)

- 1. Two fragments of the narrow bilamellar zoarium, natural size.
- 2. Young convex zooecia, \times 20. Two zooecia bear the large supraoral avicularium.
- 3. The usual aspect of the zooecial surface, \times 20. The peristomice is deeply embedded.
- 4. Much calcified, convex zooecia, \times 20.
- 5. Zoarial surface, × 20, showing two broken ovicells.
- 6. Surface of a young zoarium, \times 20.
- 7. View of the interior of the zooecia, \times 20, illustrating the areolar pores, and the subjacent olocyst.
 - 8. Longitudinal thin section, \times 12.
- 9. Transverse thin section, \times 12, exhibiting the thick zooecial walls perforated by the areolar pores.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, and Monroe County (figs. 4-6), Florida.

Figs. 10-11. Smittina maleposita, new species. (p. 144.)

- 10. Surface of the incrusting zoarium, \times 20, showing the characteristic poorly oriented zooecia. Pliocene (Caloosahatchee marl); Shell Creek, De Soto County, Florida.
- 11. Zooecia, \times 20, several of which bear the large, globular, finely perforated ovicell. Pleistocene: Vero, Florida.

Fig. 12. Floridina parvicella, new species. (p. 57.)

The incrusting type specimen, \times 20.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Figs. 13,14. Schizopodrella aculeata, new species. (p. 104.)

- 13. The massive multiliamellar zoarium, natural size.
- 14. Zooecia, × 20, showing the wide tremopores and the large, long, pointed avicularium.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

PLATE 32.

Figs. 1-f. Holoporella bicornis, new species. (p. 178.)

1, 2. Two zoarial masses, natural size.

3. The usual aspect of the surface, < 20.

4. Another surface, × 20, illustrating occurrence of the deep zooecia.

Pliocene (Waccamaw marl): Waccamaw River, Horry County, South Carolina.

Fig. 5. Aimulosia radiata, new species. (p. 140.)

The incrusting zoarium, \times 20, illustrating the characteristic interareolar radiating costules.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida.

Figs. 6-10. Holoporella albirostris Smitt, 1872. (See also pl. 7, figs. 9-14.) (p. 174.)

6. The irregular, massive zoarium, natural size.

7. Usual aspect of the surface, \times 20, showing the zooccia altered by fossilization.

8. Portion of a zoarium, \times 20, showing numerous interzooccial avicularia.

9. Another zoarial surface, \times 20, in which the areolar pores are closed and the oral sinus is limited by a spiniform umbo.

10. Interior of zooccia, \times 20, illustrating zooccia around the ancestrula.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida.

Figs. 11-13. Schismopora brevincisa, new species. (p. 180.)

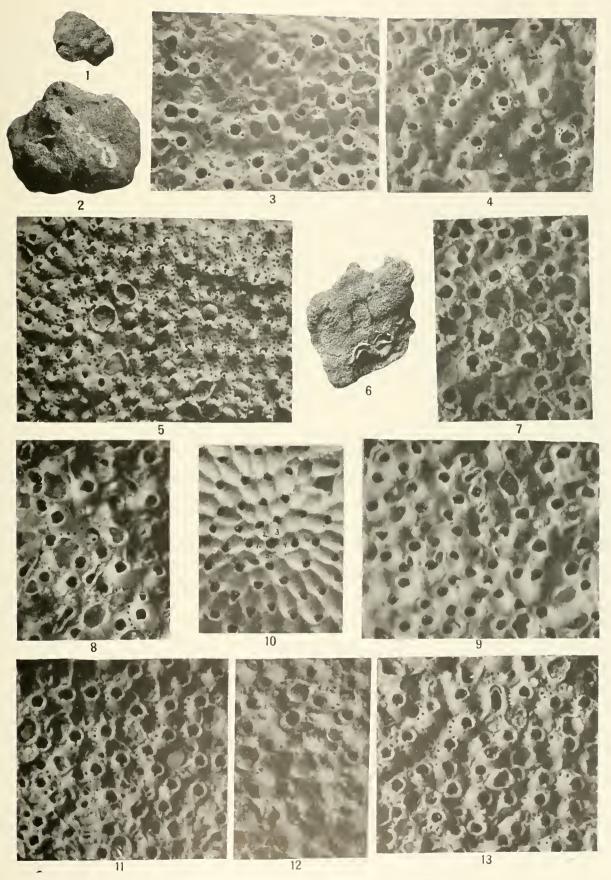
11. Surface of the incrusting zoarium, \times 20.

12. Another view, × 20, showing incomplete zooecia.

13. Zooecia, \times 20, illustrating the form of the aperture and the areolar pores.

Pliocene (Caloosahatchee marl): Shell Creek, De Soto County, Florida.

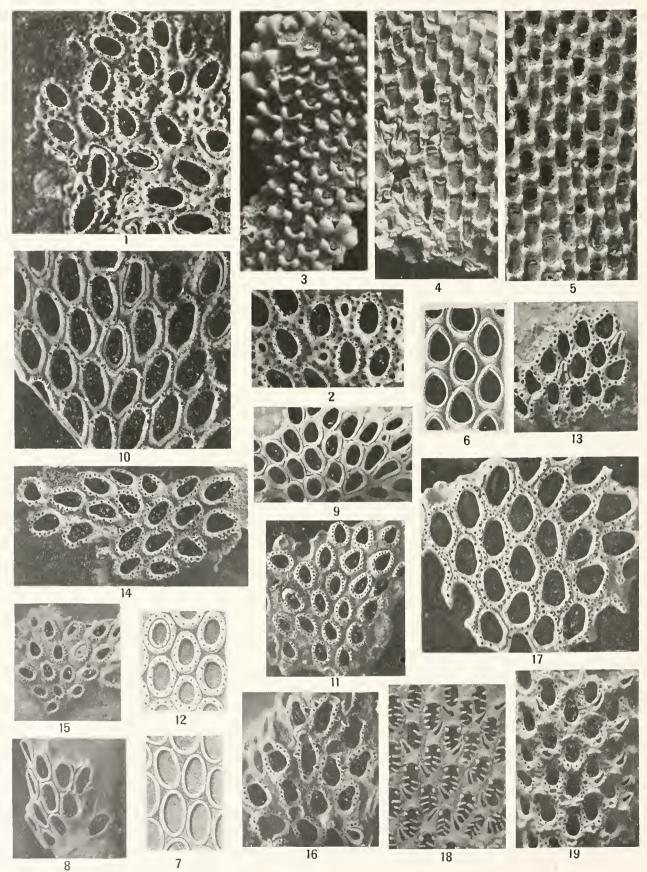
U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

U. S. NATIONAL MUSEUM

BULLETIN 125 PLATE 33



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

Figs. 1, 2. Mystriopora? arcolata, new species. (p. 19.)

- 1. The incrusting zoarium, × 20, showing the pyriform zooecia, the zooeciules, and the interzooecial areolae.
 - 2. Another example, × 20, preserving longer zooeciules.

Pleistocene: Long Wharf Canyon, Santa Monica, California.

Figs. 3-5. Membranipora tuberculata Bose, 1802. (p. 22.)

3. A fossil example of this widespread recent species, × 20, with very large tubercles.

4. Another example, × 20, with the tubercles less developed and the zooecia still retaining their chitinous parts.

Pleistocene: Santa Monica, California.

5. Surface of a recent specimen, < 20, showing identity with the fossil form.

Pacific Ocean: Santa Monica, California.

Fig. 6. Membraniporina californica Gabb and Horn, 1862. (p. 25.)

Original figure.

Pleistocene: Santa Barbara, California.

Figs. 7-9. Conopeum barbarensis Gabb and Horn, 1862. (p. 28.)

7. Gabb and Horn's figure of this species.

8. A small incrusting zoarium, < 20, showing the simplicity of structure.

9. The incrusting zoarium, \times 20, showing the distinct zooecia with thin mural rim and several avicularia.

Pleistocene: Santa Barbara, California.

Fig. 10. Hincksina quadrispinosa, new species. (p. 38.)

The incrusting zoarium, < 20, illustrating the large elongated zooecia with four distal spines. Two regenerated zooecia are also shown.

Pleistocene: Rustic Canyon, Santa Monica, California.

Fig. 11. Hincksina multispinata, new species. (p. 38.)

Zooecia, imes 20, of the incrusting zoarium, showing their small, elongated oval shape and the numerous hollow spines.

Pleistocene: Santa Barbara, California,

Fig. 12. Callopora multipora Gabb and Horn, 1862. (p. 44.)

Gabb and Horn's figure of this interesting species, which has not been rediscovered.

Pleistocene: Santa Barbara, California.

Fig. 13. Callopora crassospina, new species. (p. 41.)

The incrusting type specimen, \times 20. The long fusiform avicularium and the six hollow spines are to be noted.

Pleistocene: Santa Barbara, California.

Figs. 14-16. Cauloramphus triangularis, new species. (p. 48.)

14. The incrusting type specimen, < 20. The numerous hollow spines and the small triangular avicularium are to be noted.

An example showing the ancestrular zooecia, < 20.

16. An ovicelled specimen, \times 20.

Pleistocene: Santa Barbara, California.

Fig. 17. Cauloramphus porosus, new species. (p. 48.)

The incrusting type specimen, \times 20, illustrating the large distinct zooccia separated by numerous

Pleistocene: Santa Barbara, California.

Figs. 18, 19. Callopora horrida Hincks, 1880. (p. 43.)

18. Unilamellar, recent zoarium, / 20, with areal spines preserved.

Pacific coast near Los Angeles, California.

19. Zooecia of an incrusting fossil example, imes 20, illustrating the large avicularia and the areal

Pleistocene: Santa Barbara, California.

PLATE 34.

Figs. 1-3. Callopora circumclathrata Hineks, 1881. (p. 43.)

1. Well-preserved specimen, \times 20, with ovicells and avicularia.

2. Zoarial surface, \times 20, showing the zooccia, avicularia arcolar spaces, and the dietellae.

3. An ovicelled specimen, > 20, with large avicularia.

Pleistocene: Long Wharf Canyon, Santa Monica, California.

Fig. 4. Rhamphonotus multispinatus, new species. (p. 47.)

The incrusting type specimen, < 20, showing the two sizes of spines, a pair of large ones which constrict the opesium and numerous smaller ones.

Pleistocene: Santa Barbara, California.

Figs. 5-7. Cupularia robertsoniae, new species. (p. 82.)

5. The disk-shaped zoaria, natural size.

6. External inferior face, \vee 20.

7. Internal superior face, \times 20.

Pleistocene: Long Wharf Canyon, Santa Monica, California.

Fig. 8-10. Chaperia galeata Busk, 1853. (p. 52.)

8. A Pleistoccue example of this widespread recent and fossil species, \times 20.

Pleistocene: Santa Barbara, California.

9. Ovicelled zooccia showing the characteristic rectilinear distal border.

10. Another fragment, \times 20, with one complete ovicell and the spines and avicularia well shown. Pleistocene: Rustic Canyon, Santa Monica, California.

Figs. 11-14. Cellaria mandibulata Hincks, 1882. (p. 86.)

11. A segment, × 20, showing a large avicularian zooccium.

12. Another fragment, \times 20, with an avicularian zooecium. The round orifice of the inferior zooecium is the passage for the corneous joint bearing another segment.

13. An ordinary segment, \times 20.

14. Several fragmentary segments, natural size.

Pleistocene: Los Angeles, California.

Figs. 15-18. Cellaria fissurifera, new species. (p. 85.)

15. A worn segment, \times 20. The opening of the ovicell is very narrow.

16. Segment, > 20, bearing zooecium with round orifice through which the chitinous joint of another segment passed.

17. Segment, \times 20, with rhomboidal zooecia and bearing two avicularia. The orifice of the ovicell is a narrow elongated fissure.

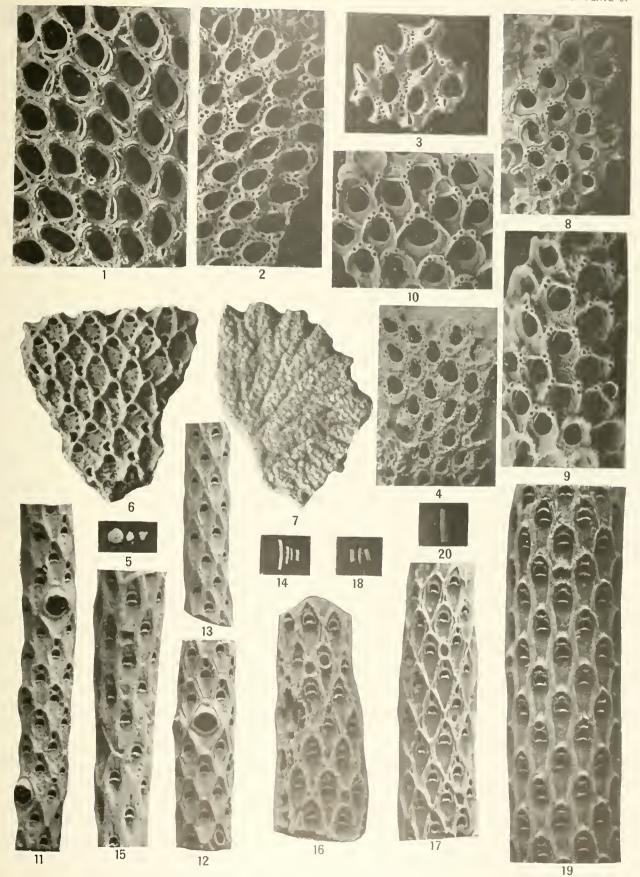
18. Several fragments, natural size.

Pleistocene: Rustic Canyon, Santa Monica, California.

Figs. 19, 20. Cellaria diffusa Robertson, 1905. (p. 86.)

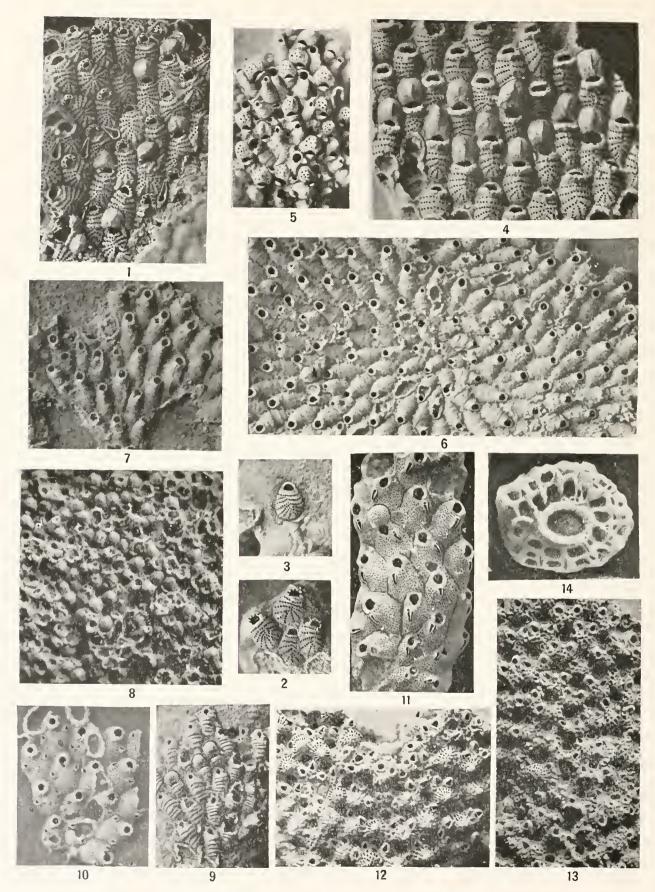
A segment natural size and enlarged, \times 20, illustrating the characters of this beautiful species. The orifice of the oxicell has the characteristic proximal tongue.

Pleistocene: Rustic Canyon, Santa Monica, California.



North American Later Tertiary and Quaternary Bryozoa.

FOR EXPLANATION OF PLATE SEE PAG 278.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 279.

PLATE 35.

Fig. 1. Puellina radiata forma scripta Reuss, 1847. (See also pl. 15, fig. 12.11 pp. 89.)

Well-preserved zoarium, \times 20, with elongated zooccia and well-developed avicularia and spines.

Pleistocene: Santa Barbara, California.

Figs. 2, 3. Puellina heermanni Gabb and Horn, 1862. (p. 89.)

2. Several zooecia of the incrnsting zoarium, × 20, showing the wide costules and the oral spines.

3. A single zooecium, \times 20, illustrating the rectilinear proximal border of the apertura.

Pleistocene: Santa Barbara, California.

Fig. 4. Metracolposa mucronata, new species. (p. 92.)

The incrusting type specimen, \times 20, showing the large keeled ovicell.

Pleistocene: Santa Barbara, California.

Figs. 5-8. Hippothoa hyalina Linnaeus, 1768. (p. 92.)

5. Portion of a zoarium, \times 20, illustrating the three kinds of zooecia.

Pleistocene: Santa Barbara, California.

6. A zoarium, \times 20, formed almost entirely of ordinary zooccia and showing the ancestrula.

7. A small incrusting patch, \times 20, with ordinary zooecia only.

8. A mature example, \times 20, in which the irregular arrangement of the male zooecia gives a celleporoid effect.

Pleistocene: Dead Mans Island, off San Pedro, California.

Fig. 9. Hippothoa hyalina, var. rugosa, new variety. (p. 94.)

Type example, \times 20, showing the characteristic frontal wrinkles.

Pleistocene: Santa Barbara, California.

Fig. 10. Schizolavella vulgaris Moll, 1803. (p. 108.)

The Pleistocene specimen referred to this species, \times 20.

Pleistocene: Santa Barbara, California.

Fig. 11. Schizomavella longirostrata Hincks, 1883. (p. 109.)

A fossil example referred to this recent species, \times 20. The large avicularium and the very small tremopores are shown.

Pleistocene: Santa Barbara, California.

Figs. 12-14. Trypematella papulifera, new species. (p. 135.)

12. Surface of the unilamellar zoarium, × 20, showing unovicelled zooecia.

13. A zoarinm, \times 20, bearing ovicell.

14. Transverse section of the hollow tubular zoarium, \times 20.

Pleistocene: Rustic Canyon, Santa Monica, California,

PLATE 36.

Fig. 1. Lepralia cribrosa! Maplestone, 1900. (p. 134.)

Surface of the specimen, \times 20, referred to this Australian Miocene species.

Pleistocene: Rustic Canyon, Santa Monica, California.

Figs. 2, 3. Fenestrulina malusi Savigny-Audouin, 1826. (p. 115.)

2. Photograph, × 20, showing small zooecia.

3. Zoarium, × 20, with large zooccia, some showing the ovicell.

Pleistocene: Santa Barbara, California.

Figs. 4, 5. Microporella ciliata Linnaeus, 1759. (See also pl. 20, figs. 1-6.) (p. 119.)

4. Zoarium, × 20, exhibiting zooccia with small avicularia.

5. Zoarium, imes 20, in which the zooecia have large avicularia.

Pleistocene: Santa Barbara, California.

Figs. 6, 7. Microporella umbonata Hincks, 1882. (p. 123.)

6. Marginal zooecia without ovicell, \times 20.

7. Ovicelled zooccia of the same specimen, \times 20, exhibiting also the frontal umbo.

Pleistocene: Santa Barbara, California.

Figs. 8-10. Microporella e difornica Hincks, 1883. (p. 123.)

8. Zooccia, \times 20, showing only a single but well-developed avicularium.

9. Zoarium with ovicell zooccia, × 20, with a single avicularium.

Pleistocene: Long Wharf Canyon, Santa Monica, California.

10. Normal and ovicelled zooecia of a large expansion, \times 20.

Pleistocene: Santa Barbara, California.

Figs. 11, 12. Microporella vibraculifera Hincks, 1883. (p. 124.)

11. A zoarium, \times 20, with ancestrular zooecia.

Pleistocene: Rustic Canyon, Santa Monica, California.

12. Ovicelled and normal zooecia, \times 20, illustrating the avicularia and the six hollow spines.

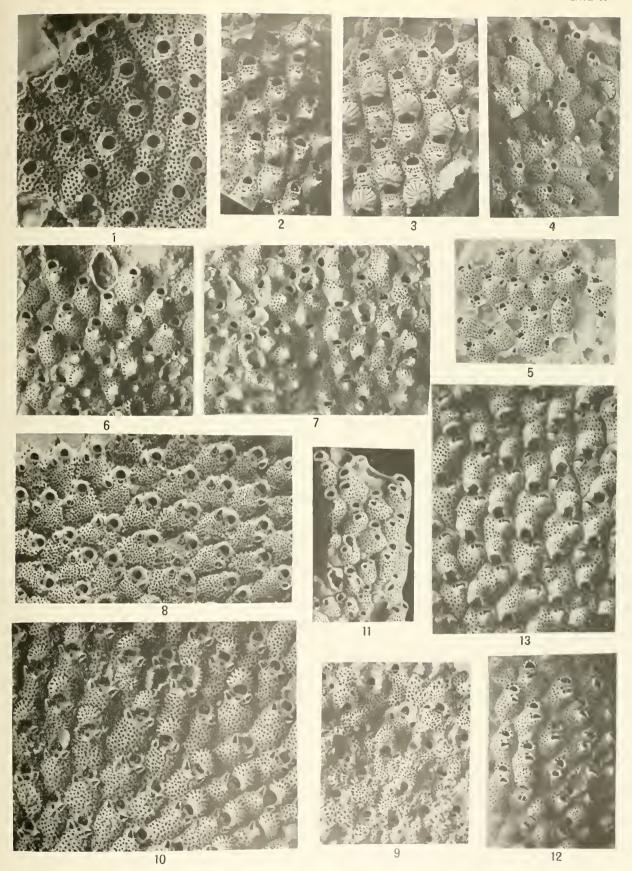
Pleistocene: Santa Barbara, California,

Fig. 13. Microporella customata Gabb and Horn, 1862. (p. 124.)

Zoarial surface, \times 20, with ovicelled and normal zooecia and with the avicularium oblique to the zooecial axis.

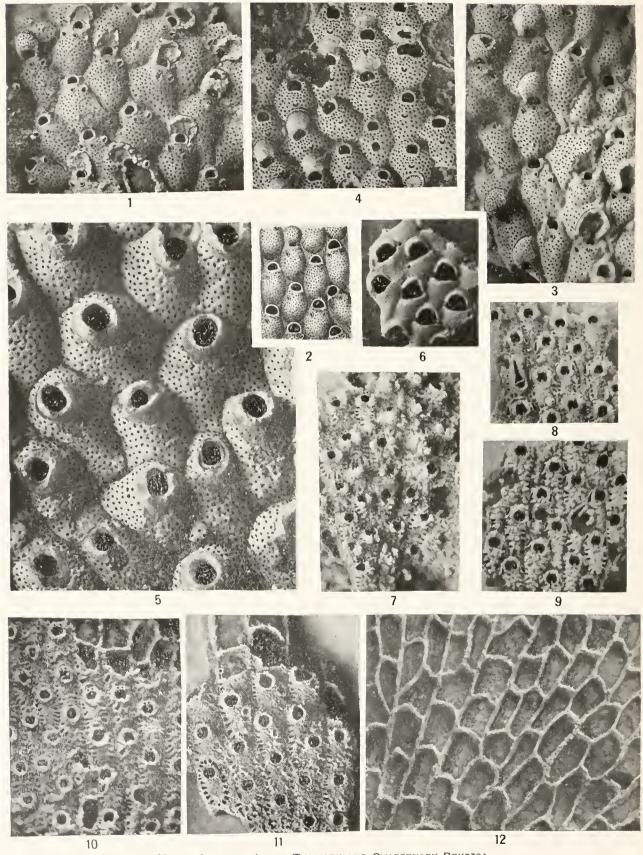
Pleistocene: Rustic Canyon, Santa Monica, California.

U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

BULLETIN 125 PLATE 37



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 281.

PLATE 37.

Figs. 1, 2. Microporella heermani Gabb and Horn, 1862. (p. 125.)

1. Ordinary and ovicelled zooecia, × 20. The large dimensions, granular frontal and small tremopores and avicularia are illustrated.

Pleistocene: Rustic Canyon, Santa Monica, California.

2. The original figure of the species.

Pleistocene: Santa Barbara, California.

Fig. 3. Microporella gibbera, new species. (p. 126.)
Surface of the unilamellar zoarium, × 20, showing the distinct, irregular, gibbose zooecia.

Pleistocene: Rustic Canyon, Santa Monica, California.

Fig. 4. Fenestrulina porosa, new species. (p. 117.)

Surface of the incrusting zoarium, \times 20.

Pleistocene: Long Wharf Canyon, Santa Monica, California.

Fig. 5. Cyclicopora gigantea, new species. (p. 139.)

Surface of the incrusting zoarium, \times 20, showing the extreme size of the zooecia.

Pleistocene: Tremochal Canyon, Santa Monica, California.

Fig. 6. Eurystomella bilabiata Hineks, 1884. (p. 142.)

 Λ fossil specimen, \times 20, referred to this interesting recent species.

Pleistocene: Dead Mans Island, off San Pedro, California.

Figs. 7-9, Smittina californicusis Robertson, 1908. (p. 146.)

- 7. Zooecia, imes 20, showing thin areolar costules and the peristome formed by two lateral tuberosities.
- 8. Another example, \times 20, with two small lateral avicularia and a large interzooecial avicularium developed.
- 9. Surface of zooecia, imes 20, with the two small avicularia, one of which has the beak directed upward and the other downward.

Pleistocene: Santa Barbara, California.

Figs. 10-12. Smittina grandicella, new species. (p. 145.)

- 10. The incrusting zoarium, imes 20, showing the large distinct zooecia and the distal pores in
 - 11. Another example, \times 20, not so well preserved.
- 12. A zoarium, \times 20, with the frontal removed, illustrating the sinuous arrangement of the walls, which are without septulae.

Pleistocene: Rustic Canyon, Santa Monica, California.

PLATE 38.

Figs. 1-7. Smittina discoidea, new species. (p. 145.)

- I. Fragments of the unilamellar zoarium, natural size.
- 2. Surface, × 20, showing the large interzooccial avicularium.
- 3. Zooccial surface, \times 20, with the large axicularium replaced by two small ones. Oxicelled zooccia are present.
 - 4. Lower side, \times 20.
 - 5. Another surface, \times 20, in which the large avicularium is unguiculate.
 - 6. Zooecia, × 20, with the large avicularia turned in different directions.
 - 7. Interior of zooccia, \times 20, showing that the lyrula is accompanied by two cardelles.

Pleistocene: Rustic Canyon, Santa Monica, California.

Fig. 8. Cystisella ariculifera, new species. (p. 152.)

Portion of the incrusting zoarium, \times 20, showing the small ancestrular zooecia and the large marginal ones.

Pleistocene: Santa Barbara, California.

Fig. 9. Smittina porifera Hineks, 1884. (p. 147.)

Zooecia of the incrusting zoarium, \(20\), showing the convex frontal perforated by pores.

Pleistocene: Santa Monica, California.

Figs. 10-15. Porella collifera Robertson, 1908. (p. 148.)

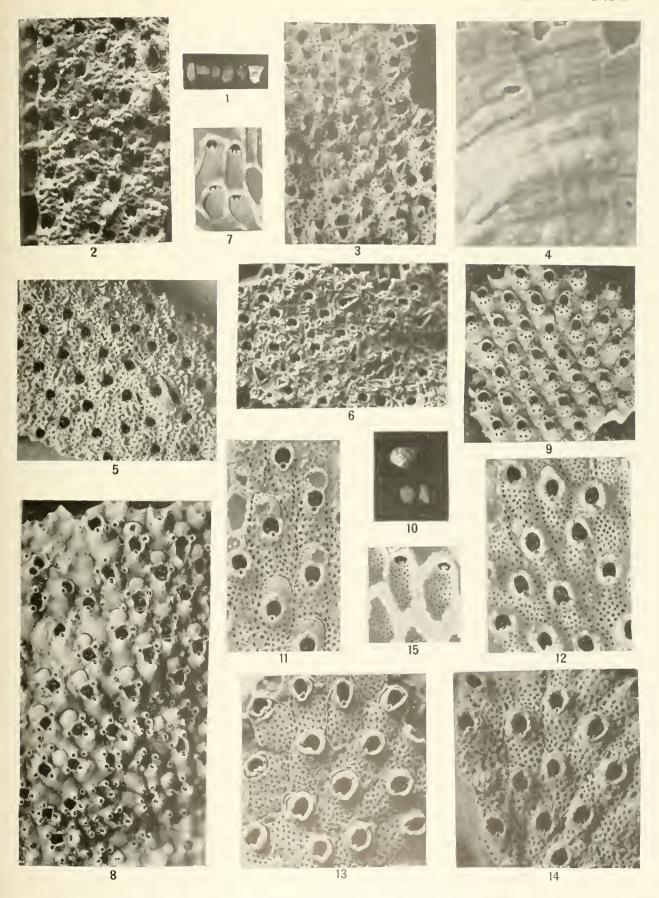
- 10. Zoaria, natural size.
- 11. Oyicelled zooecia, \times 20.
- 12. Surface, 14 20, with one zooecium bearing a small ovicell.
- 13. Zooecia, ≤ 20 , with a very salient peristome.

Pleistocene: Santa Monica, California.

- 14. Zooecia, × 20, with large tremopores.
- 15. Interior of zooecia, \times 20. The oral axicularium is supported by a denticulated lyrula.

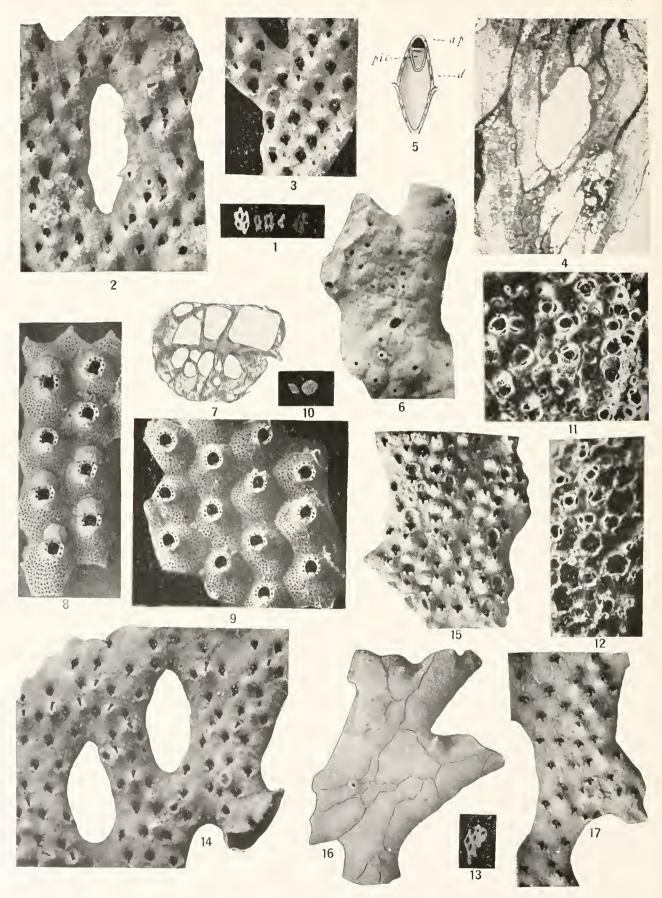
Pleistocene: Santa Barbara, California.

U. S. NATIONAL MUSEUM



North American Later Tertiary and Quaternary Bryozoa.

U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 283.

PLATE 39.

- Figs. 1-7. Philolopora pacifica Robertson 1908. (p. 154.)
 - 1. Zoarial fragments, natural size.
 - 2. Celluliferous side of fragment, \times 20, with the frontal axicularium well developed.
 - 3. Another fragment, \times 20, in which the frontal avicularium is absent or worn away.
 - 4. Tangential thin section of the dorsal, \times 25, illustrating the special calcification of the vibices.
 - 5. View of the interior of the zooecia, \times 20. ap, apertura; pic, peristomie: d, dietellae.
 - 6. Dorsal side of zoarium, \times 20, showing some radicular pores and an avicularium.
 - 7. Transverse thin section, × 25.

Pleistocene: Rustic Canyon, Santa Monica, California.

- Figs. 8, 9. Phylaetella spinosissima, var. major Hincks, 1884. (p. 170.)
 - 8. Zooecia of the incrusting zoarium, \times 20, showing the recumbent ovicell.
 - 9. A specimen, × 20, with zooccia exhibiting the spines, oral mucro, and the tremopores. Pleistocene: Long Wharf Canyon, Santa Monica, California.
- Figs. 10-12. Costazzia robertsoniae, new species. (p. 181.)
 - 10. Zoaria, natural size.
 - 11. Surface of the globular zoarium, \times 20, with ovicelled zooecia and interzooecial avicularia.
 - 12. Another surface, \times 20, showing incomplete zooecia.

Pleistocene: Rustic Canyon, Santa Monica, California.

- Figs. 13-17. Phidolopora labiata Gabb and Horn, 1862. (p. 154.)
 - 13. The reticulated zoarium, natural size.
 - 14. Celluliferous surface, ×20, showing zooecia with large salient avicularia.
 - 15. A fragment, \times 20, with ovicelled zooecia.
 - 16. Dorsal side of a fragment, \times 20, illustrating the irregular vibices.
 - 17. Another fragment, × 20, with the zooecia distinctly outlined by a salient thread.

Pleistocene: Santa Barbara, California.

PLATE 40.

- Figs. 1-4. Tubucellaria punctulata Gabb and Horn, 1862. (p. 170.)
 - I. Zoarial fragments, natural size,
 - 2. A ramified segment, \times 20,
 - 3. Another segment, \times 20, illustrating the zooecial surface.
 - 4. Longitudinal section, \times 20.

Pleistocene: Santa Barbara, California,

Figs. 5, 6. Tubicellaria punctulata, var. minor, new variety. (p. 170.)

A fragment natural size, and surface of the same, \times 20. The smaller size of the [zooecia and tremopores is evident.

Pleistocene: Santa Barbara, California.

Fig. 7. Lagenipora spinulosa Hincks, 1884. (p. 171₂)

An example, < 20, much changed by fossilization.

Pleistocene: Dead Mans Island, off San Pedro, California.

Figs, 8, 9. Holoporella umbonata, new species. (p. 178.)

8. Zooecial surface, \times 20, illustrating the arcolar pores.

9. Well-developed example with interareolar costules radiating from the umbo. Several ovicelled zooccia are present.

Pleistocene: Santa Barbara, California.

Figs. 10, 11. Schismopora abrupta, new species. (p. 180.)

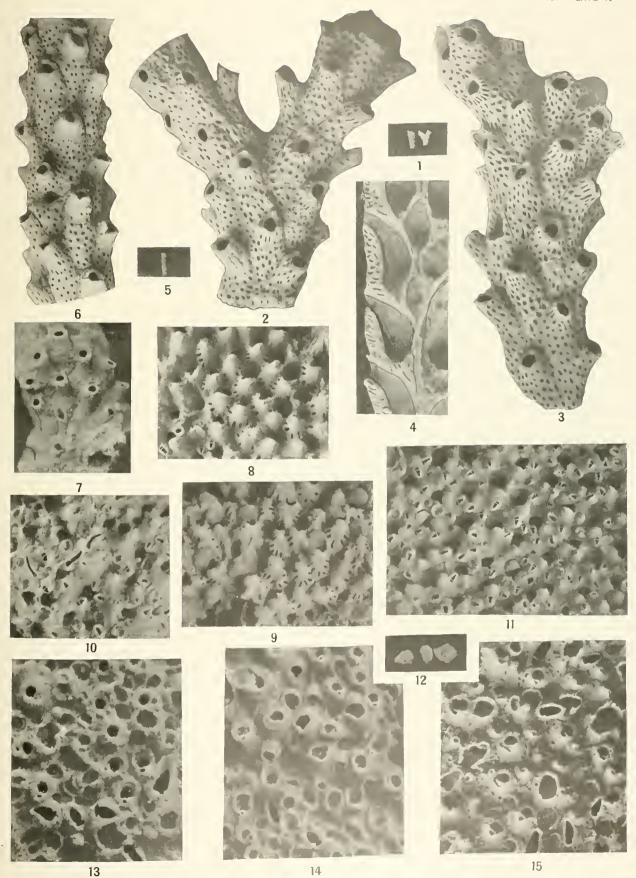
- 10. Surface of the incrusting zoarium, \times 20, with some of the usual small avicularia transformed into very long ones.
- 11. Another surface, × 20. The transverse, smooth, deeply embedded ovicell and the numerous small triangular avicularia are quite visible.

Pleistocene: Santa Barbara, California.

Figs. 12-15. Schismopora lanceolata, new species, (p. 181.)

- 12. The small lamellar zoarial masses, natural size.
- 13. Surface, -, 20, illustrating the triangular sinus of the apertura and the lanceolate interzooecial avicularia.
 - 14. Ovicelled zooecia, \times 20. The characteristic avicularia are also present.
 - 15. Portion of the zooecial surface, \times 20, showing the incomplete zooecia.

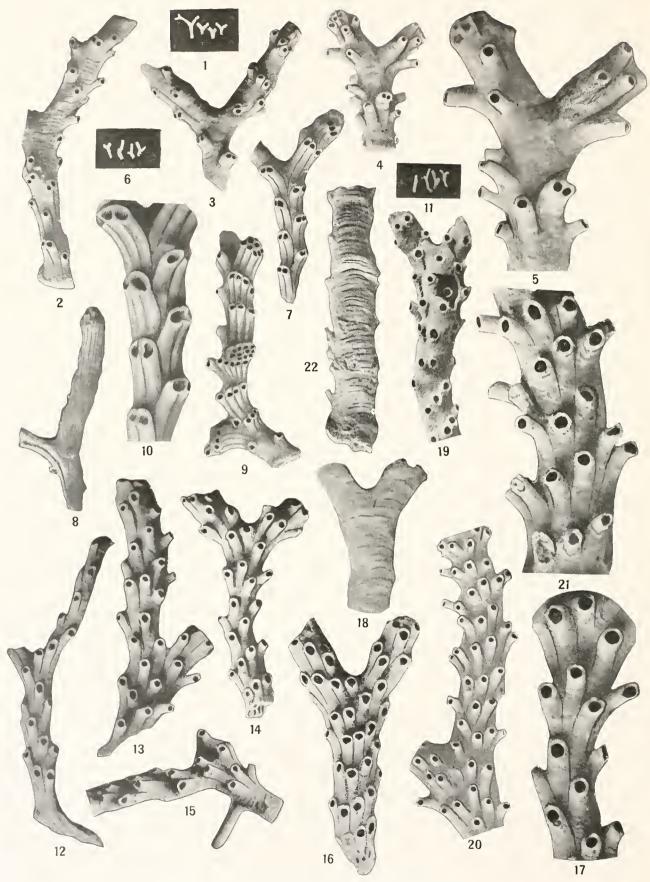
Pleistocene: Santa Monica, California.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

U. S. NATIONAL MUSEUM

BULLETIN 125 PLATE 41



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

PLATE 41.

Figs. 1-5. Entalophora fasciculifera, new species. (p. 195.)

1. Zoarial fragments, natural size.

2. A large example, × 12, with tubes little salient.

3. A fragment, \times 12, showing three fascicles of two zooecia each.

4, 5. An example with long peristomes, \times 12, and a portion of the same, \times 25.

Pleistocene: Tremochal Canyon, Santa Monica, California.

Figs. 6-10. Idmonea dispar, new species. (p. 198.)

6. Fragments, natural size.

7. Anterior (cellular) face, \times 12.

8. Posterior side showing longitudinal striations, \times 12.

9. Lateral view of a fragment, × 12, bearing a small branch.

10. A portion of figure 7, \times 25.

Pleistocene: Rustic Canyon, Santa Monica, California.

Figs. 11-19. Filisparsa clarki, new species. (p. 195.)

11. Fragments of the zoarium, natural size.

12-14. Three examples, \times 12, showing different aspects of the anterior face.

15. A specimen with a dorsal apophysis, \times 12.

16. An example, \times 12, in which the peristomes are grouped in transverse and oblique rows.

17. Portion of a zoarium, \times 25.

18. Posterior side, \times 12.

19. An ovicelled example, \times 12.

Pleistocene: Santa Barbara, California.

Figs. 20-22. Filisparsa clarki, var. parvula, new variety. (p. 196.)

20, 21. Anterior side of the zoarium, \times 12 and \times 25, showing the slightly smaller dimensions.

22. Posterior side, \times 12, illustrating transverse wrinkling.

Pleistocene: Santa Barbara, California.

PLATE 42.

Figs. 1-7. Crisia serrata Gabb and Horn, 1862. (p. 196.)

- 1. Zoarial segments, natural size.
- 2. Characteristic segment, \times 12.
- 3. Noncelluliferous side of a segment, \times 25, with two ramal bases.
- 4. Another segment, \times 25, with two ramal bases. Some of the apertures are closed by lamellae.
- 5. Another segment, \times 25.
- 6. A segment, \times 25, showing the characteristic median carina and but a single ramal base.

Pleistocene: Rustic Canyon, Santa Monica, California.

7. Portion of an ovicelled segment, \times 25.

Pleistocene: Santa Barbara, California.

Fig. 8. Crisia, species.

Several segments, \times 12, still united by their corneous joints, which have not been destroyed by fossilization.

Pleistocene: Rustic Canyon, Santa Monica, California.

Figs. 9-17. Tubuli pora fasciculifera Hincks, 1884. (p. 197.)

- 9-11. Three zoaria, \times 12, illustrating variations in form.
- 12. An example, \times 12, showing a small lateral ovicell.
- 13. Another ovicelled example, \times 12, illustrating the large oeciostome at the beginning of a fascicle.
 - 14, 15. Two unovicelled zoaria, >, 12, with short fascieles.
 - 16. Several zoaria, natural size.
 - 17. Fragment of a zoarinm, × 25.

Pleistocene: Santa Barbara, California.

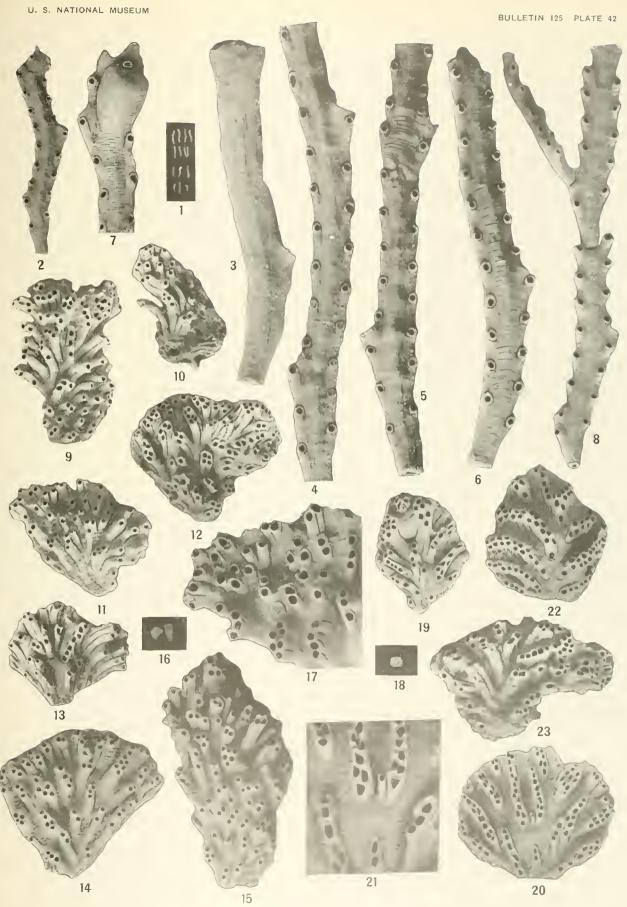
Figs. 18-23. Tubulipora tuba Gabb and Horn, 1862. (p. 198.)

- 18. Zoarium, natural size.
- 19. A young ovicelled specimen with short fascicles, \times 12.
- 20. An ovicelled example with a long capillary oeciostome, \times 12.
- 21. Surface of the same, \times 25, illustrating the ovicell and oeciostome.

Pleistocene: Rustic Canyon, Santa Monica, California.

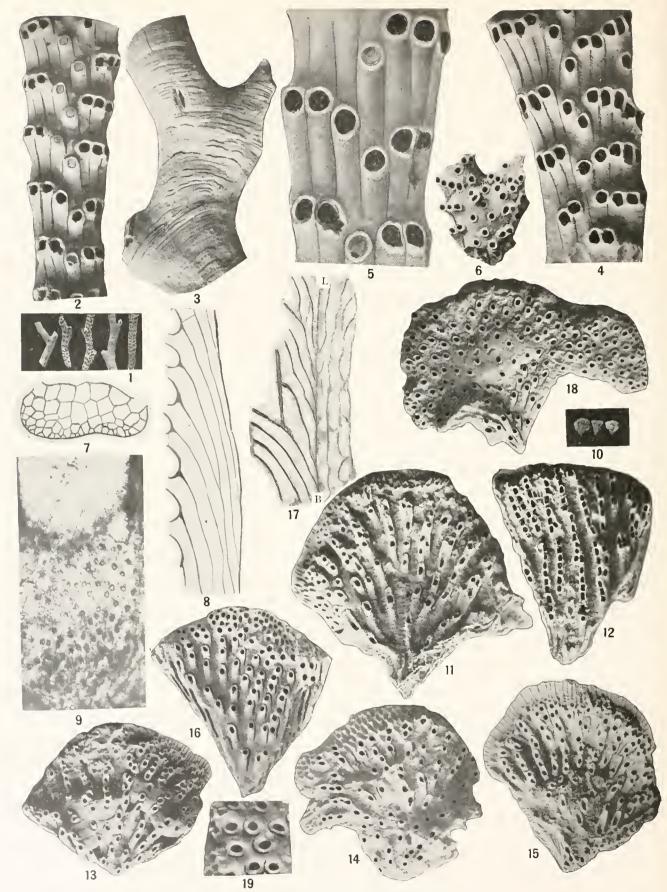
- 22. An idmoneiform ovicelled specimen, \times 12, with long fascicles.
- 23. An irregular flabellate specimen, \times 12.

Pleistocene: Santa Barbara, California.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 288.



NORTHIAMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 287.

PLATE 43.

Figs. 1-9. Idmonea californica D'Orbigny, 1852. (p. 199.)

- 1. Zoarial fragments, natural size.
- 2. Cellnliferous side, \times 12, showing the axial zooecia isolated and closed by a diaphragm.
- 3. Posterior side, \times 12, illustrating the transverse striations.
- 4. Another example, \times 12, in which the axial zooecia are open.
- 5. A portion of figure 2, \times 25.
- 6. An ovicelled specimen, \times 6.
- 7. Transverse thin section, \times 12. The tubes are polygonal.
- 8. Longitudinal thin section, \times 12.
- 9. Tangential thin section, \times 100. The mural perforations are no larger than in smaller species of the genus.

Pleistocene: Santa Barbara, California.

Figs. 10-17. Stathmepora flabellata, new species. (p. 201.)

- 10. Zoarial fragments, natural size.
- 11. A rather complete flabellate specimen, imes 12, with incomplete fascicles.
- 12. A fragment, × 12, showing bifurcated fascicles.
- 13. A specimen with large ovicell, \times 12.
- 14. An example with broken ovicell, \times 12, illustrating that the peristomes are here adjacent in the fascicles.
 - 15. An ovicelled example, imes 12, with a large basal lamella.

Pleistocene: Santa Barbara, California.

- 16. A flabellate example, \times 12, with a thick zone of growth.
- 17. Longitudinal section, \times 25. The tubes are cylindrical with triparietal germation. L. B.= basal lamella.

Pleistocene: Rustic Canyon, Santa Monica, California.

- Figs. 18, 19. Diaperoccia flabellata, new species. (p. 202.)
 - 18. The flabellate bilamellar zoarium, \times 12, with an ovicell developed.
 - 19. Portion of the same, \times 25.

Plesitocene: Rustic Canyon, Santa Monica, California.

PLATE 44.

Figs. 1-3. Lichenopora hispida Fleming, 1828. (p. 203.)

1. Several zoaria, natural size,

2. An example, \times 12, illustrating that the fascicles do not reach the zoarial margin.

3. An ovicelled example, × 12, with the ovicell not entirely covered by the cancelli and showing no occiopore.

Pleistocene: Santa Barbara, California.

Figs. 4-7. Lichenopora californica Conrad, 1855. (p. 203.)

4. Zoaria, natural size.

5. An ovicelled example, \times 12. The occiostome is located between two fascicles.

6. Another ovicelled example, \times 12, showing clearly the salient oeciostome and the ovicell covered with cancelli.

7. Inferior side of a discoid, free example, \times 12.

Pleistocene: Santa Barbara, California.

Figs. 8-9. Lichenopora verrucaria Fabricius, 1780. (p. 205.)

Celluliferous surfaces of two ovicelled examples, > 12, referred to this recent species.

Pleistocene: Rustic Canyon, Santa Monica, California.

Fig. 10. Lichenopora radiata Savigny-Andouin, 1826. (p. 204.)

An incomplete although typical zoarium of this wide spread recent species, \geq 12. The cancelli are larger than the zooccial oritices.

Pleistocene: Rustic Canyon, Santa Monica, California.

Figs. 11-21. Psilosolen capitiferar, Canu and Bassler, 1922. (p. 207.)

11. A longitudinal thin section, \times 25.

12. A longitudinal section showing the ovicell at the end of the branch, \times 12.

13. Longitudinal thin section through a bifurcated branch, \times 25. The tubes are cylindrical and ramify at all heights.

14, 15. Transverse thin sections, \times 25.

16. An example, \times 12, illustrating the irregular quincunx arrangement of the peristomes.

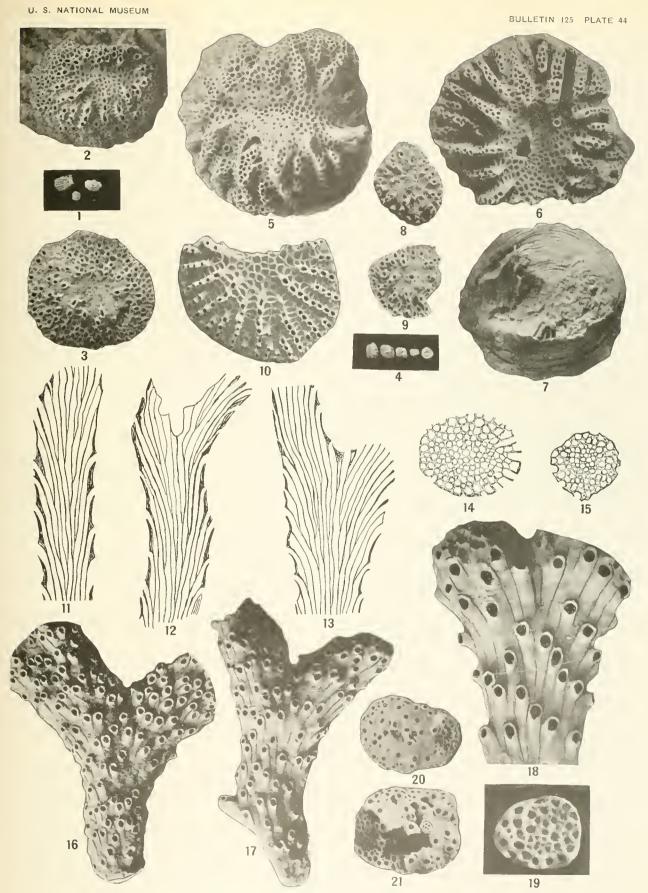
17. An example, \times 12, with the peristomes arranged in zones.

18. A portion of the same zoarial surface, + 25, illustrating that the tubes are visible and the peristome thin.

19. An ordinary transverse section, \times 25.

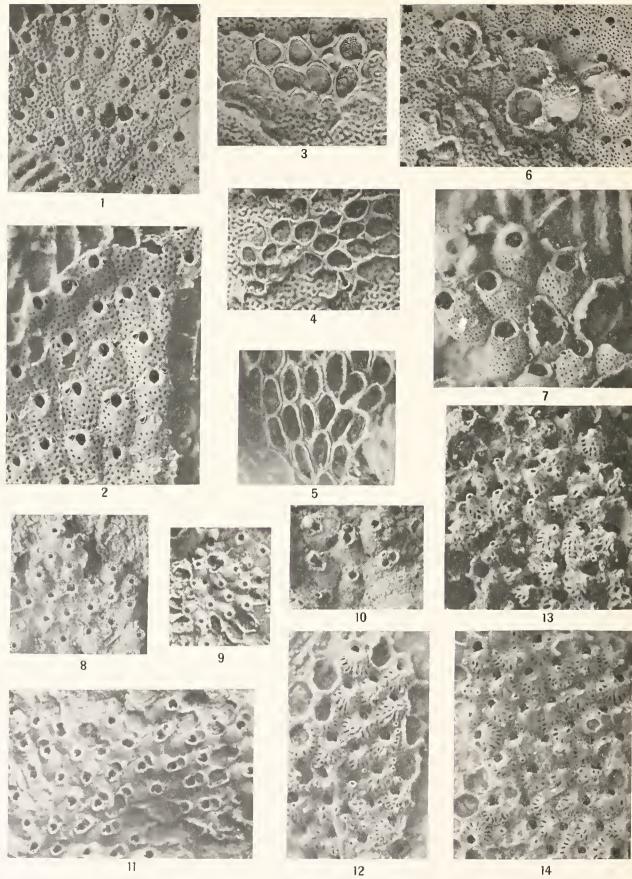
20, 21. Ovicelled specimens, \times 12, illustrating the position of the ovicell at the flattened end of the branch.

Pleistocene: Santa Barbara, California.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 288.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 289.

PLATE 45.

Figs. 1, 2. Schrzoporella magniporosa, new species. (p. 95.)

1. View of the incrusting zoarium, × 20, showing the hexagonal zooccia with large tremopores.

2. Ancestrular portion of a zoarium, \times 20. The aperture of the ancestrula is very large.

Pleistocene: Mount Hope, Panama Canal Zone.

Figs. 3, 4. Callopora guernei Jullien, 1903. (p. 42.)

3. Several marginal zooecia, × 20, of a zoarium incrusting a coral.

4. Zooecia, × 20, in the vicinity of the ancestrula and exhibiting an avicularium.

Pleistocene: Mount Hope, Panama Canal Zone.

Fig. 5. Callopora filum Jullien, 1903. (p. 42.)

Zooecia, \times 20.

Pleistocene: Mount Hope, Panama Canal Zone.

Fig. 6. Styloponia projecta, new species. (p. 103.)

Zooecia, \times 20, of the incrusting zoarium, exhibiting the ovicell and the small oral avicularium characteristic of the species.

Pleistocene: Mount Hope, Panama Canal Zone.

Fig. 7. Dakaria chevercuxi Jullien, 1903. (p. 97.)

Zooecia, \times 20.

Pleistocene: Mount Hope, Panama Canal Zone.

Figs. 8, 9. Hippoporina pusilla, new species. (p. 129.)

8. The incrusting zoarium, \times 20, with ovicelled and badly oriented zooecia.

9. Zoarium, × 20, showing the ancestrula and ancestrular zooecia.

Pleistocene: Mount Hope, Panama Canal Zone.

Fig. 10. Mastigophora pesanseris Smitt, 1873. (p. 172.)

Zooecia, \times 20.

Pleistocene: Mount Hope, Panama Canal Zone.

Fig. 11. Rhynchozoon levigatum, new species. (p. 157.)

Zooecia of the incrusting zoarium, \times 20. The salient globular ovicell and the spinule in the peristome are shown.

Pleistocene: Mount Hope, Panama Canal Zone.

Figs. 12, 13. Porella costulata, new species. (p. 150.)

12. Zooccia of the incrusting zoarium, \times 20, in the vicinity of the ancestrula.

13. Normal zooecia, \times 20, showing the large tremopores and prominent avicularian mucro.

Pleistocene: Mount Hope, Panama Canal Zone.

Fig. 14. Hippoporella costulata, new species. (p. 133.)

Portion of the incrusting zoarium, \times 20, with zooccia exhibiting arcolar pores and costules.

Pleistocene: Mount Hope, Panama Canal Zone.

PLATE 46.

Fig. 1. Holoporella turrita Smitt, 1873. (p. 179.)

The fossil example, \times 20, referred to this recent species.

Pleistocene: Mount Hope, Panama Canal Zone.

Fig. 2. Holoporella aviculifera, new species. (p. 179.)

A portion of the incrusting zoarium, \times 20. A small avicularium in the peristome is visible.

Pleistocene: Mount Hope, Panama Canal Zone.

Figs. 3-6. Cyclicopora multilamellosa, new species. (p. 138.)

3. Surface of the free orbicular zoarium, × 20, illustrating the form of the zooecia and ovicell.

4. Zooecia \times 20, with a large transverse avicularium.

5. Ordinary zooecia, \times 20.

6. Interior of zooecia, \times 20. The tremocyst is visible through the very thin olocyst.

Pleistocene Mount Hope, Panama Canal Zone.

Fig. 7. Holoporella mucronata, new species. (p. 179.)

The incrusting zoarium, \times 20, illustrating the large areolar pores and prominent oral mucro. Pleistocene: Mount Hope, Panama Canal Zone.

Figs. 8, 9. Acanthodesia savartii forma texturata Reuss, 1847. (See also pl. 5, figs. 1-5.) (p. 32.)

Surface of two small unilamellar examples, \times 20.

Pleistocene: Vero, Florida.

Fig. 10. Hemiseptella tuberosa, new species. (p. 71.)

The incrusting zoarium, × 20, showing the irregular inferior border of the opesium.

Pleistocene: Simmons Bluff, Yonges Island, Charleston County, South Carolina.

Figs. 11, 12. Membranipora osburni, new species. (p. 24.)

11. The incrusting zoarium, \times 20, illustrating the thin, rounded mural rim, the tubercles, and the small, concave proximal cryptocyst.

Pleistocene: Mount Hope, Panama Canal Zone.

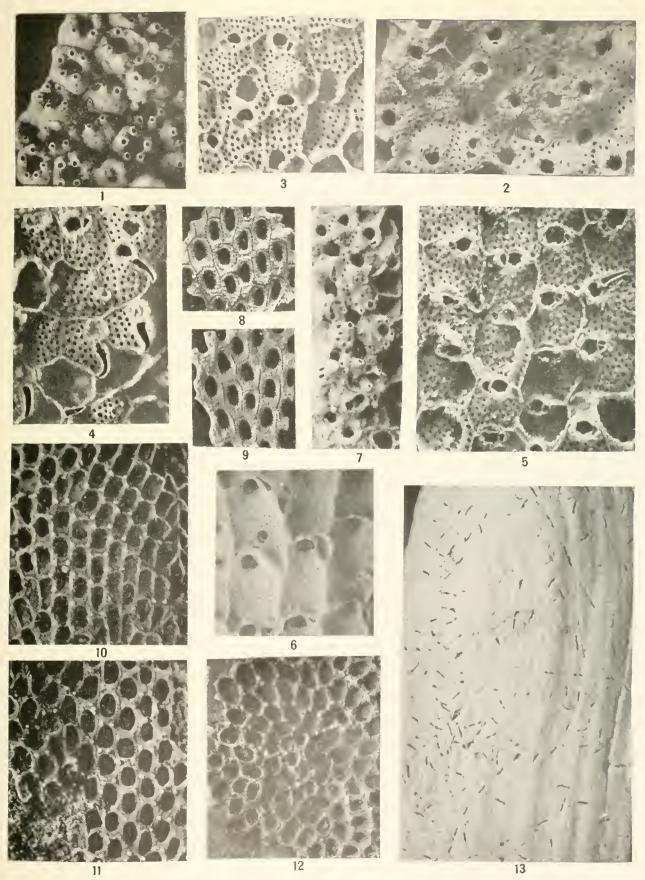
12. Ancestrular region of a zoarium, \times 20, referred to this species.

Miocene (Bowden marl): Bowden, Jamaica.

Fig. 13. Terebripora pacifica, new species. (p. 15.)

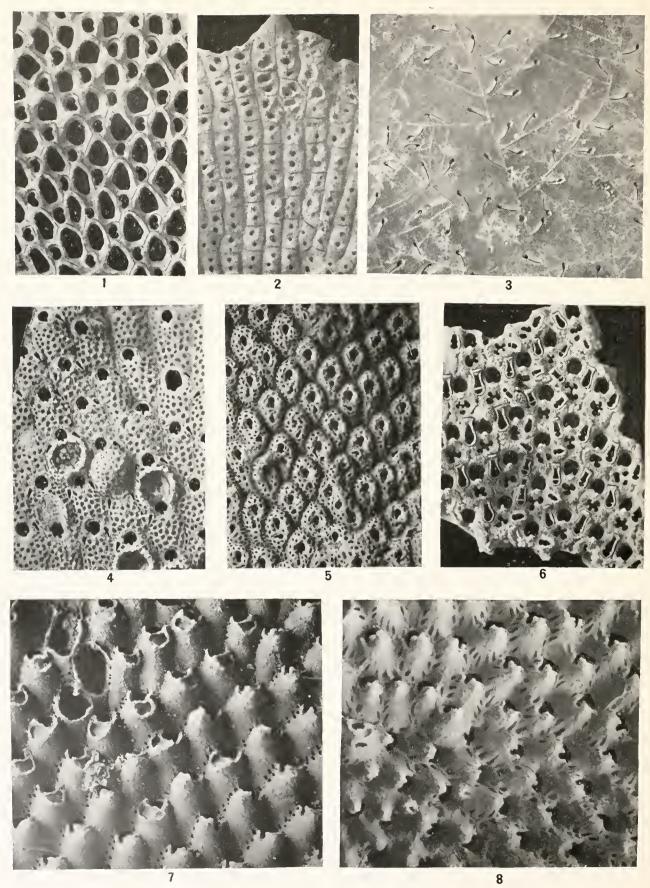
The type specimen, \times 10, showing the small pyriform zooecia separated by one to two times their length.

Pleistocene: Santa Barbara, California.



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

U. S. NATIONAL MUSEUM



NORTH AMERICAN LATER TERTIARY AND QUATERNARY BRYOZOA.

FOR EXPLANATION OF PLATE SEE PAGE 291.

PLATE 47.

Figs. 1, 2. Cupuladria biporosa, new species. (p. 29.)

1. Celluliferous surface of the orbicular zoarium, \times 20, showing the wide zooccia and the large auriculate vibracula.

2. Inner face of the same zoarium, \times 20, with the characteristics two pores to each zooecium. Lower Miocene (Bowden horizon): Santo Domingo.

Fig. 3. Spathipora longirima, new species. (p. 16.)

A portion of the type specimen, \times 20, illustrating the delicate canalicules with the attached zooecia, the apertures of which bear a long rimule.

Pliocene (Waccamaw marl): Waccamaw River, South Carolina.

Fig. 4. Stylopoma magniporosa, new species. (p. 103.)

Surface of the bilamellar zoarium \times 20. The large tremopores, the avicularia, and the ovicell hiding the aperture are apparent.

Lower Miocene (Bowden horizon): Santo Domingo.

Fig. 5. Laminopora miocenica, new species. (p. 160.)

Zooecial surface, \times 20, with the parietal dietellae, tremopores, elongate oval zooecia, and long rimule visible.

Lower Miocene (Bowden horizon): Santo Domingo.

Fig. 6. Tremogasterina truncatorostris, new species. (p. 244.)

The bilamellar zoarium, \times 20, with the truncated axicularium and the large trifoliate fronta pore well preserved.

Lower Miocene (Bowden horizon): Santo Domingo.

Figs. 7, 8. Rhynchozoon grandicella, new species. (p. 156.)

7. Marginal zooecia of the incrusting zoarium, \times 20, showing the pleurocyst frontal with small areolar pores and the prominent avicularian chamber.

8. Ancestrular zooccia, \times 20, with the costules of the frontal better developed.

Pleistocene: Rustic Canyon, Santa Monica, California.

alarupta, Schismopora.		Page.	Page,
acarolima, Chaperin.	abrupta, Schismopora,		
Acenidaca		50	
Immoss		30	Aspidostoma giganteum. 65
Disposition		30	Aspidostomidae 64
Savart 1. 1. 1. 1. 1. 1. 1. 1		34	
Savari	rectangularia.	34	reptans 191
bioliata		30,31	auricularia, Selenaria
Manufactura 192		33	auriculatum, Metrarabdotos 164
reytl	delicatula	33	avellana, Tretocycloccia 206
Texturata		32	aviculifera, Cystisella 152
Lexturata	revti	31	Holoporella
typica		32	baccata, Hippodiplosia 131
aeuleata, Aumulosia. 139 Batopora 158, 1884 Schizopodrella 144 Adeona. 158 rosula. 189 heckéli 155 stolickai 188 hporoa. 155 barbarensis, Conopeum. 288 riolacra 150 barbarensis, Conopeum. 288 riolacra 150 beanianta, Retepera 153 Adeonellopsis. 161 beanianta, Retepera 153 Adeonella. 162 beanianta, Retepera 153 Adeonella. 163 bella, Lepralia. 147 Actea. 19 Beanianta, Retepera 153 Adeonella. 164 beanianta, Retepera 153 Adeonella. 165 beanianta, Retepera 153 Adeonella. 167 beanianta, Retepera 153 Adeonella. 168 beanianta, Retepera 153 Adeonella. 169 Beanianta, Retepera 153 Adeonella. 169 Beanianta, Retepera 153 Adeonella. 169 Beanianta, Retepera 153 Adeonella. 160 Beanianta, Retepera 153 Adeonella. 161 bidentata, Fedora (Cupularia) 160 Beanianta, Retepera 161 bidentata, Fedora (Cupularia) 161 Biflustra delicatula. 163 Aumulosia. 163 bidentata, Fedora (Cupularia) 163 Alderina. 164 bidentata, Fedora (Cupularia) 163 Alderina. 165 bidentata, Fedora (Cupularia) 163 Alderina. 165 bidentata, Fedora (Cupularia) 163 Alderina. 165 bidentata, Fedora (Cupularia) 163 Alderina. 168 Amphilbestrum. 169 bidentata, Fedora (Cupularia) 163 Alderina. 169 bidentata, Fedora (Cupularia) 163 Alderina. 169 bidentata, Fedora (Cupularia) 163 Alderina. 160 bidentata, Fedora (Cupularia) 163 Alderina. 161 bidentata, Fedora (Cupularia) 163 Alderina. 162 bidentata, Pedora (Cupularia) 163 Alderina. 163 Anasca. 164 bidentata, Chaperia 164 Anasca. 165		31	
Schizopotrella	The state of the s	139	Batopora
Mecha 158	,	104	
heckeli		158	rosula 188
Darbacea 158		158	stoliczkai 188
violacea 159 Membranipora 28 Adeonellopsis 161 beaniana, Retepora 133 coccinella 161 bella, Lepralia 147 umbilicata 158 Porella 147 Adeonidae 158 Smittia 147 Actea 19 Berenicea flabellum? 194 anguina? 19 biaperta, Eschara 190 gellus, Amphilitestrum 47 Lepralia 190 Ramphonotus 49 Schizoporella 99 Aimulosia 139 Stephanosella 99 aculeata 139 bicornis, Holoporella 178 brevis 140 bidientata, Fedora (Cupularia) 191 radiata 140 bifiliata, Carchiae 33 aitrensis, Inversiula (Microporella) 122 swarti 33 aitrensis, Inversiula (Microporella) 174 Microporella 122 biscopora 174 Microporella 123 biscopora 174		158	barbarensis, Conopeum
Adeonellopsis 161	*	159	
161		161	
umbilicata 158 Porella 147 Adeonidae 158 Smittia 1147 Actea 19 Berenicea flabellum? 194 anguina? 49 biaperta, Eschara 100 agellus, A mphiblestrum 47 Lepralia 100 Ramphonotus 49 Schizoporella 99 Aimulosia 139 bicornis, Holoporella 178 brevis 140 bidientata, Fedora (Cupularia) 191 radiata 140 Biflustra delicatula 33 airensis, Inversiula (Microporella) 129 suarti 33 airensis, Inversiula (Microporella) 129 suarti 33 airensis, Inversiula (Microporella) 129 suarti 33 albiospora 174 bifloata, Acanthodesia savarti 23 albiospora 174 Microporella 121 atbispina, Chaperia (Membranipora) 35 bigiberra, Hippotiplosia 131 Alderina 41 Microporella 141		161	bella, Lepralia 147
Adeonidae 158 Smittia 117 Actea 49 Berenicea flabellum? 194 anguina? 49 biaperta, Eschara 100 agellus, Amphiblestrum 47 Lepralia 100 Ramphonotus 49 Schizoporella 99 Aimulosia 139 Stephanosella 99 aculeata 130 bicornis, Holoporella 178 bervis 140 bidentata, Fedora (Cupularia) 191 radiata 140 Bifustra delicatula 33 aitrostris, Crelic pora 174 bifoliata, Acanthodesia savarti 23 albirostris, Celle pora 174 Membranipora 33 Holopoprella 174 Microporella 121 albispina, Chaperia (Membranipora) 52 bigierra, Hippodiposia 131 Alderina 39 Lepralia 141 Cesticella 39 Lepralia 142 Amphiblestrum 41 bilamellata, Chaperia 51,52 annulus		158	Porella
Actea		158	
anguina?		49	Delegates maneralism
agellus, Amphihlestrum. 47 Lepralia. 100 Ramphonotus. 49 Schizoporella. 90 Nimulosia. 139 Stephanosella. 99 brevis. 140 bidentata, Fedora (Cupularia) 191 brevis. 140 bidentata, Fedora (Cupularia) 191 radiata. 140 bidentata, Fedora (Cupularia) 191 radiata. 140 bidentata, Fedora (Cupularia) 191 alirensis, Inversiula (Microporella). 122 swarti. 33 alirensis, Inversiula (Microporella). 123 swarti. 33 albispina, Cellepora. 174 Microporella. 121 dibispina, Chaperia (Membranipora) 52 bigiberra, Hippodiplosia 131 Alderina. 39 bilabalta, Eurystomella 141, 142 eesticella 39 Lepralia. 141, 142 eesticella 44 bilamellata, Chaperia 51, 52 argeltus. 45 biporosa, Cupuladria 29 ecsticella 45		49	ona provide a social contraction of the soci
Ramphonotus 49 Schizoporella 99 Aimulosia 139 Stephanosella 99 aculeata 139 bicornis, Holoporella 178 brevis 140 bidentata, Fedora (Cupularia) 191 radiata 140 Biflustra delicatula 33 sirenis, Inversiula (Microporella) 129 swarti 32 albirostris, Cellepora 174 bifoliata, Acanthodesia savarti 23 Biscopora 174 Membranipora 33 Holoporella 174 Membranipora 33 Albicaria 39 bigiberra, Hippodiplosia 131 Alderina 39 bilabiata, Eurystomella 111 acesticella 39 Lepralia 142 Amphiblestrum 44 bilamellata, Chaperia 51,52 agunulus 52 biporosa, Cupuladria 29 constrictum 41 bitrevina, Thalamoporella 62 deformis 160 160 resistatum 52		47	25t pratta
Aimulosia 139 Stephanosella 99 aculeata 139 bicornis, Holoporella 178 brevis 140 bidentata, Fedora (Cupularia) 191 radiata 140 Biflustra delicatula 33 airensis, Inversiula (Microporella) 129 savarti 32 albirostris, Cellepora 174 bifoliata, Acanthodesia savarti 23 Biscopora 174 Membranipora 33 Holoporella 174 Microporella 131 Alderina 39 Lepralia 141 Cesticella 39 Lepralia 141 cesticella 39 Lepralia 142 Amphiblestrum 41 bilamellata, Chaperia 51,52 annulus 52 biporosa, Cupuladria 29 constrictum 41 birerio, Lepralia 10 cristalam 52 birevicalda, Pyripora 19 amplectens, Heterooccium 162 brevicalda, Pyripora 19 anglica, Herpetopo		49	Denta portera
aculeata. 139 bicornis, Holoporella. 178 brevis. 140 bidentata, Fedora (Cupularia) 191 radiata. 140 Biflustra delicatula. 33 savarti 32 alirensis, Inversiula (Microporella). 129 bifoliata, Acanthodesia savarti 23 Albicopora 174 bifoliata, Acanthodesia savarti 23 Discopora 174 Microporella 121 albispina, Chaperia (Membranipora) 32 bigiberra, Hippodiplosia 131 Alderina. 39 bilabitata, Eurystomella. 141 Amphiblestrum. 44 bilamellata, Chaperia 142 Amphiblestrum. 45 bijperforata, Thalamnoporella 62 annulus. 52 bijperforata, Thalamnoporella 62 annulus. 54 bijperforata, Thalamnoporella 62 constrictum 45 botterii, Lepralia. 160 cristatum 52 Bracebridgia 160 cristatum 65 brevicaula, Pyripora 19 Anarthropora 162 Anarthropora 163 Anasca 17 brevinetisa, Schismopora 180 angulatum, Rhynehozoon 153 angulatum, Rhynehozoon 153 Anmulus, Amphiblistrum 52 Chaperia 164 Chaperia 165 Chaperia 165 Arthropoma 161 Arthropoma 162 Arthropoma 163 Arthropoma 164 Arthropoma 165 Arthropoma 165 Arthropoma 166 Arthropoma 167 Arthropoma 168 Arthropoma 169 Arthropoma 169 Arthropoma 160 Cristata 160 Chaperia 160 Chap		139	Ett pilatiobella
Drevis.		139	bieornis, Holoporella
radiata. 140 Biflustra delicatula. 33 3avarti 32 3albirostris, Celle pora. 174 bifoliata, Acanthodesia savarti 23 23 23 23 24 25 25 25 25 25 25 25		140	indentary i capta (dipartition and)
uirensis, Inversiula (Microporella) 129 savarti 32 albirostris, Cellepora 174 bifoliata, Acanthodesia savarti 23 Discopora 174 Membranipora 33 Holoporella 174 Microporella 121 albispina, Chaperia (Membranipora) 52 bigiberra, Hippodiplosia 131 Alderina 39 Lepralia 141 Cesticella 39 Lepralia 142 Amphiblestrum 44 bilamellata, Chaperia 51, 52 agellus 47 biperforata, Thalamoporella 62 annulus 52 biporosa, Cupuladria 29 constrictum 41 biterii, Lepralia 108 cristalam 52 biporosa, Cupuladria 108 <tr< td=""><td></td><td>140</td><td>Title Gold of technique of the contract of the</td></tr<>		140	Title Gold of technique of the contract of the
albirostris, Cellepora. 174 bifoliata, Acanthodesia savarti. 23 Discopora. 174 Membranipora. 33 Holoporella. 124 Microporella. 121 albispina, Chaperia (Membranipora). 52 bigiberra, Hippodiplosia. 131 Alderina. 39 bilabiata, Eurystomella. 141 Cesticella. 39 Lepralia. 142 Amphiblestrum. 44 bilamellata, Chaperia. 51, 52 agrellus. 47 biperforata, Thalamoporella. 62 annulus. 52 biporosa, Cupuladria. 29 constrictum. 44 botterii, Lepralia. 108 cristatam. 52 bracebridgia. 160 cristatam. 52 bracebridgia. 160 tenniparietis. 45 deformis. 160 amplectens, Heterooecium. 162 brevicauda, Pyripora. 19 Anasca. 17 brevicauda, Pyripora. 18 angica, Herpetopora. 15 brevicoilis, Lagenipora (?). <td></td> <td>129</td> <td>O'EUGI CO</td>		129	O'EUGI CO
Discopora		174	Difformatia, Academore Sta Savarett
Holoporella	, ,	174	Me morani por a
albispina, Chaperia (Membranipora) 52 bigiberra, Hippodiplosia 151 Alderina 39 bigiberra, Hippodiplosia 151 Alderina 39 Lepralia 142 Amphiblestrum 44 bilamellata, Chaperia 51, 52 agellus 47 biperforata, Thalamoporella 62 annulus 52 biporosa, Cupuladria 29 constrictum 52 biporosa, Cupuladria 29 constrictum 52 biporosa, Cupuladria 160 cristatum 52 biperforata, Thalamoporella 160 cristatum 52 biperforata, Thalamoporella 160 cristatum 52 biperforata, Thalamoporella 160 deformis 160 deformis 160 deformis 160 tenuiparietis 45 brevicauda, Pyripora 19 Anarthropora 161 brevincisa, Schismopora 150 anglica, Herpetopora 15 brevincisa, Schismopora 150 anglica, Herpetopora 15 brevincisa, Schismopora 150 anguina?, Aetea 49 brevis, Aimulosia 161 anguina?, Aetea 153 Smittina (?) 160 annulus, Amphiblistrum 52 californica, Idmonea 190 annulus, Amphiblistrum 52 Lepralia 123 Ammoranipora 161 Membraniporina 203 Amembranipora 161 Membraniporina 203 areolata, Mystriopora (?) 19 Microporella ciliata 123 Arthropoma 97 Microporella ciliata 203 Arthropoma 203 Chaperia 203 cornuta 203 Microporella ciliata 203 cornuta 203 Micr		174	ATTET OF OTEMA
Alderina. 39 bilabiata, Eurystomella 1142 cesticella 39 Lepralia 142 Amphiblestrum. 44 bilamellata, Chaperia 51, 52 agellus. 47 biperforata, Thalamoporella 62 annulus. 52 biporosa, Cupuladria 29 constrictum. 44 botterii, Lepralia 108 cristatum. 52 Bracebridgia 160 tenniparietis. 45 deformis 160 tenniparietis. 18 brevicauda, Pyripora 19 Anarthropora. 162 brevicauda, Pyripora 19 Anasca. 17 brevicauda, Pyripora 18 anglica, Herpetopora. 16 brevicollis, Lagenipora (?) 171 Anasca. 17 brevinesias, Schismopora. 180 anglica, Herpetopora. 49 brevirostris, Hippaliosina 105, 166 angulatum, Rhynchozoon. 153 Smittina (?) 116 annulus, Amphiblestrum. 52 californica, Idmonea		52	Digiperra, Implomptosa
cesticella 39 Leprala 142 Amphiblestrum 44 bilamellata, Chaperia 51, 32 agellus 47 biperforata, Thalamoporella 62 annulus 52 biporosa, Cupuladria 29 constrictum 44 botterii, Lepralia 108 cristatum 52 Bracebridgia 160 tenuiparietis 45 deformis 160 tenuiparietis 18 brevicauda, Pyripora 19 Anarthropora 162 brevicollis, Lagenipora (?) 171 Anarthropora 162 brevinceisa, Schismopora 180 anglica, Herpetopora 15 brevirostris, Hippallosina 165 166 angulatum, Rhynchozoon 153 Smittina (?) 116 166 annulus, Amphibitestrum 52 californica, Idmonea 123 Chaperia 53 Lepralia 123 Membranipora 52 Lichenopora 203 areolata, Mystriopora (?) 19 Microporella ciliata </td <td></td> <td>39</td> <td></td>		39	
Amphiblestrum. 44 bilamellata, Chaperia 3, 22 agellus 47 biperforata, Thalamoporella 62 annulus. 52 biperforas, Cupuladria 29 constrictum. 44 botterii, Lepralia 10 cristatum. 52 Bracebridgia 160 tenuiparietis 45 deformis 160 amplectens, Heterooccium 162 brevicauda, Pyripora 17 Anartropora. 162 brevicollis, Lagenipora (?) 171 Anasca 17 brevincisa, Schismopora 180 anglica, Herpetopora 18 brevirostris, Hippaliosina 165 anguina?, Aetea 49 brevis, Aimulosia 146 angulatum, Rhynchozoon 153 Smittina (?) 146 annulus, Amphiblestrum 52 californica, Idmonea 123 Membranipora 52 Licencopora 203 areolata, Mystriopora (?) 19 Microporella 23 Arthropoma 97 Microporella ciliata <td< td=""><td></td><td>39</td><td></td></td<>		39	
agellus 47 biperiorata, Thanamopotena. 29 annulus. 52 biperiorata, Thanamopotena. 29 constrictum. 44 botterii, Lepralia. 108 cristatum. 52 Bracebridgia. 160 tenuiparietis. 45 brevicauda, Pyripora. 19 amplectens, Heterooccium. 162 brevicollis, Lagenipora (?). 171 Anarthropora. 17 brevincisa, Schismopora. 180 Anasca. 17 brevincisa, Schismopora. 180 anglica, Herpetopora. 18 brevirostris, Hippaliosina. 165 anguina?, Aetea. 49 brevis, Aimulosia. 194 annulus, Amphibibestrum. 52 smittina (?). 116 Annulus, Amphibibestrum. 52 Lichenopora. 203 Membranipora. 52 Lichenopora. 203 areolata, Laminopora. 161 Membraniporina. 23 areolata, Mystriopora (?). 19 Microporella ciliata. 123 Arthropoma. 97		44	bilamellata, Chaperia
annulus. 52 biporesa, Cupuladria. 29 constrictum. 44 botterii, Lepralia. 108 cristatum. 52 Bracebridgia. 160 tenuiparietis. 45 deformis. 160 amplectens, Heterooecium. 18 brevicauda, Pyripora. 19 Anarthropora. 162 brevicollis, Lagenipora (?). 171 Anasca. 17 brevincisa, Schismopora. 180 anglica, Herpetopora. 18 brevirostris, Hippaliosina. 165, 166 angulatum, Rhynehozoon. 133 Smittina (?). 140 angulatum, Rhynehozoon. 153 Smittina (?). 16 annulus, Amphiblestrum. 52 californica, Idmonea. 199 Chaperia. 53 Lepralia. 123 Membranipora. 263 Membraniporina. 23 areolata, Mystriopora (?). 19 Microporella. 123 Arthropoma. 97 Microporella ciliata 123 Arthropoma. 97 Viicavea. <t< td=""><td></td><td>47</td><td>Diperiorata, Thatamoporcha.</td></t<>		47	Diperiorata, Thatamoporcha.
constrictum. 44 botterii, Lepralia. 108 cristatum. 52 Bracebridgia. 166 tenuiparietis. 45 deformis. 160 amplectens, Heterooecium. 18 brevicanda, Pyripora. 19 Anarthropora. 162 brevicollis, Lagenipora (?). 171 Anasca. 17 brevineisa, Schismopora. 180 anglica, Herpetopora. 18 brevirostris, Hippaliosina 105 lee anglica, Herpetopora. 49 brevis, Aimulosia. 140 angulatum, Rhynchozoon. 153 Smittina (?). 16 annulus, Amphibitestrum. 52 californica, Idmonea 199 Chaperia. 53 Lepralia. 123 Membranipora. 263 Membraniporina 263 arbuscula, Laminopora. 161 Membraniporina 23 arcolata, Mystriopora (?). 19 Microporella ciliata 123 Arthropoma. 97 Microporella ciliata 123 cornuta. 97 Microporella ciliata 203		52	bijorosa, cupulauria
cristatum. 52 Bracebridgia. 160 tenuiparietis 45 deformis. 160 amplectens, Heterooccium 18 brevicauda, Pyripora. 19 Anarthropora. 162 brevicollis, Lagenipora (?). 171 Anasca. 17 brevincisa, Schismopora. 180 anglica, Herpetopora. 49 brevirostris, Hippaliosina 165, 166 anguina?, Actea. 49 brevirostris, Aimulosia 140 angulatum, Rhynchozoon. 153 Smittina (?). 146 annulus, Amphiblestrum 52 californica, Idmonea 193 Chaperia. 53 Lepralia 123 Membranipora 52 Lichenopora 203 areolata, Laminopora 161 Membraniporina 23 areolata, Mystriopora (?). 19 Microporella ciliata 123 Arthropoma 97 Microporella ciliata 23 cornuta 203 193		44	outer a, Depraise.
tenuiparietis 45 deforms 100 amplectens, Heterooecium 18 brevicauda, Pyripora 19 Anarthropora 162 breviceollis, Lagenipora (?) 171 Anasca 17 brevincisa, Schismopora 180 Angica, Herpetopora 15 brevirostris, Hippaliosina 165, 166 anglica, Herpetopora 15 brevis, Aimulosia 194 angulatum, Rhynchozoon 153 Smittina (?) 116 annulus, Amphiblistrum 52 californica, Idmonea 193 Chaperia 53 Lepralia 123 Membranipora 52 Lichenopora 203 arbuscula, Laminopora 161 Membraniporina 223 arcolata, Mystriopora (?) 19 Microporella 123 Arthropoma 97 Microporella ciliata 123 cornuta 202 19		52	Diacebridgia
amplectens, Heterooecium 18 brevicauda, Pyripora 19 Anarthropora 162 brevicauda, Pyripora 17 Anarthropora 17 brevicolisa, Schismopora 180 Anasca 17 brevirostris, Hippaliosina 165, 166 anglica, Herpetopora 49 brevirostris, Hippaliosina 140 angulatum, Rhynchozoon 153 Smittina (?) 116 annulus, Amphiblestrum 52 californica, Idmonea 193 Chaperia 53 Lepralia 123 Membranipora 203 Membraniporina 223 arbuscula, Laminopora 161 Membraniporina 223 areolata, Mystriopora (?) 19 Microporella 123 Arthropoma 97 Microporella ciliata 123 cornuta 97 Unicavea 203		4.5	deloi uns
Anarthropora. 162 brevicous, Lageripota (*) 180 Anasea 17 brevirostris, Schismopora. 180 anglica, Herpetopora. 18 brevirostris, Hippaliosina. 165, 166 angulatum, Rhynehozoon. 153 Smittina (?). 146 annulus, Amphiblestrum. 52 californica, Idmonea. 199 Chaperia. 53 Lepralia. 123 Membranipora. 52 Lichenopora. 203 arbuscula, Laminopora. 161 Membraniporina. 23 areolata, Mystriopora (?). 19 Microporella. 123 Arthropoma. 97 Microporella ciliata 123 cornuta. 202 19 Microporella ciliata 203		18	offericanda, i yripora
Anasea 17 brevineisa, Senismopora. 16 anglica, Herpetopora 49 brevirostris, Hippaliosina 165, 166 anguina?, Actea 49 brevis, Aimulosia 140 angulatum, Rhynchozoon 153 Smittina (?) 116 annulus, Amphibiestrum 52 californica, Idmonea 123 Chaperia 53 Lepralia 223 Membranipora 52 Lichenopora 203 arbuscula, Laminopora 161 Membraniporina 23 areolata, Mystriopora (?) 19 Microporella 123 Arthropoma 97 Microporella ciliata 123 cornuta 97 Unicavea 203		162	brevieours, Lagenrpora (*)
anglica, Herpetopora. 18 brevirostris, Hippanosna. 184 anguina?, Aetea. 49 brevirostris, Hippanosna. 186 anguina?, Aetea. 153 Smittina (?). 116 annulus, Amphiblestrum. 52 californica, Idmonea. 199 Chaperia. 53 Lepralia. 123 Membranipora. 161 Membraniporina. 23 areolata, Mystriopora (?). 19 Microporella. 123 Arthropoma. 97 Microporella ciliata. 123 cornuta. 203 Unicavea. 203		17	Dievincisa, achishtopora.
anguina?, Aetea. 49 brevis, Alliniosis. 116 angulatum, Rhynchozoon. 153 Smittina (?). 116 annulus, Amphiblestrum. 52 californica, Idmonea. 123 Chaperia. 53 Lepralia. 203 Membranipora. 161 Membraniporina. 203 areolata, Laminopora. 19 Microporella. 123 Arthropoma. 97 Microporella ciliata. 123 cornuta. 97 Unicavea. 203		18	
153 Smittina (:). 193 201 202 202 202 202 202 203 203 204 205 205 205 205 205 20		49	
annulus, Amphiblestrum 52 caniornea, fundosa 123 Chaperia. 53 Lepralia. 123 Membranipora. 52 Lichenopora. 203 arbuscula, Laminopora. 161 Membraniporina. 23 areolata, Mystriopora (?) 19 Microporella. 123 Arthropoma. 97 Microporella ciliata 123 cornuta. 97 Unicavea 202		153	Silliting (1)
Chaperia. 53 Leprala. Membranipora. 52 Lichenopora. 203 arbuscula, Laminopora. 161 Membraniporina. 23 areolata, Mystriopora (?). 19 Microporella. 123 Arthropoma. 97 Microporella ciliata. 123 cornuta. 97 Unicavea. 203		52	Californica, Tamones
Membranipora. 52 Lienenopora. 2co arbuscula, Laminopora. 161 Membraniporina. 2co areolata, Mystriopora (?). 19 Microporella. 123 Arthropoma. 97 Microporella ciliata. 123 cornuta. 97 Unicavea. 203		53	Lepi atta
arbuscula, Laminopora. 161 Membramporma. areolata, Mystriopora (?). 19 Microporella. 123 Arthropoma. 97 Microporella ciliata. 123 cornuta. 97 Unicavea. 203		.52	Lichenopota
areolata, Mystriopora (?) 19 Microporeua 123 Arthropoma 97 Microporella ciliata 123 cornuta 97 Univavea 203		161	Memoramporma
Arthropoma 97 Microportia titula 203 cornuta 97 Universa 203		19	311ct oporcha
cornuta		97	A telephoteta tettata
009		97	C Recurrent
	12184 23—Bull, 125 24		293

	Page.		Fage.
raliforniensis, Cellepora.	11,7	5 cesticella, Alderina.	98
? Membrantpora	_ 43	3 Chaperia	50,51
Smittia	140	acanthina.	50
Smittina	144	all ispina.	25
Callopora	40	annulus	53
circumelathrata	. 43		52
erassospina .	- 41		51
	4(51,52
dumerilii			
lata .	10		51
filum	= 42		
guernei	42		52
horrida	43		52
lanccolata	41		52
nultipora	4.1		ů.
parvirostris ==	41	tropica	52
speciosa	44	Chaperiidae	= #
tenell r	24	charlesworthi, Melicerita .	8.
Calloporina	113, 128	Cheilopora sincera	163
decorata.	112, 129		168
diadema = =	. 125		163, 169
renipuneta.	128		17
scandens	. 129		97
	81		87
Calpensia			
impressa	68, 8		
Calpensiidae	67, 68		115
calpensis, Membranipora.	83		F12, 115, 118, 119
caminosa, Chaperia = =	5-		119
Membranipora	5		121
canariensis, Cupuladria	28, 29, 8	2 circumclathrata, Callopora	6
Membranipora =	. 29	8 Membranipora	. 42
capensis, Chaperia (Amphiblestrum).	. 55	2 clarki, Filisparsa	193
capitiferax, Psilosolen	200	parvula, Filisparsa	196
carolinensis, Puellina radiata	90	clavula, Hippaliosina (Lepralia)	167
catenularia, Pyripora	- 1		187
Cauloramphus	4		161
porosus	4		145
triangularis	1		162
	*		68,69
Cellaria			26
diffusa	4		25
fissurifera	`	·	21
mandibulata			
sinuosa			20
tenella	5.		27
Cellariidae	×		20
Cellepora	183	2 constructum, Amphiblestrum.	-11
albirostris	17-	4 contigua, Lepraliella	155
californiensis.	11.	5 contorta, Laminopora	161
ciliata	119	9 convoluta, Palmicellaria	150
cribrosa	18	3 coriacea, Micropora	58 59
heckeli	153		97
hyalina	9:		100
intermata.	100		84
maculata.	183		4
malusii	11.	L.	68, 8-
manalia	- 17		181
massali«	18		181
minuta			Si
rarccosta	9		13:
script:1,	8		
spongites	10		150
subtorquata.	13		91
tesselata	12		191
umbilicata.	15		4
rerruculati	1.5	-	63
Celleporella hyalina	1)		63
Celleporidae	17		17:
Ceriopora	19	3 poissoni crinispina	17
virginiana.	19	3 'Cribrilina	87
cervicornis, Chaperia	51,5	2 cuspidata .	85

	Page.		Burn
Criorilina ligulata.	88	diadema, Calloporma (Lepralia	Page.
punctata	87	Diaperoccia	202
radiata innominata.	90	flabellata	202
Cribrilinidae	87	milneana	202
cribrosa, Cellepora	183	Diaperoeciidae	201
Lepralia	134	Diastopora flabellum	194
erinispina, Crepidaeantha poissoni.	174	Diastoporidae	193
(`risia	196	diffusa, Cellaria	56
pacifica	196	Diplodidymia	= 67,69
serrata	196	complicata.	68,69
Crisiidae =	196	Diporula	125, 126
Crisina	200	hastigera	125
striatopora	200	verrueosa	112, 125
cristata, Chaperia (Ampliblestrum	51	Discoflustrellaria doma .	77
crustacea, Myriozoella	185, 186	discoidea, Smittina	= = 145
Ctenostomata	15 163	Discopora albirostris	174
cuculata, Watersiporaeucullata, Spathipora	163	hispidu	203
	97	Discoporetta denticuluta	80
cumulata, Schizoporella Cupuladria	28,75	dispar, Idmonea disparilis, Reptescharellina	198 58
biporosa	26	divaricata, Microporella	118
canarieusis	28	doma, Cupularia	77
Cupularia cunariensis	29, 82	Discoflustrellaria	77
denticulata	77	doverensis, Retepora	152
denticulata	79	Schizopodrella.	107
doma	77	dumerilii, Callopora	40
guineensis	29	Flustra	40
haidingeri	77,7%	Membranipora	_ 40
johnsoni	77	dutertrei, Mustigophora	172
lowei	80	echinata, Holoporella (?)	177
multispinata	76, 75	edax, Leiosella	142
reussiana	77, 78	edwardsi, Fedora	191
robertsoniae	82	Electra	17
umbellata	75, 76, 80	monostachys	17
timpenata			
curtum, Rhynchozoon	. 156	pilosa	
curtum, Rhynchozoon cuspidata, Cribrdina	. 156 88	pilosa Electrinidae	18 17
enrtum, Rhynchozoon euspidata, Cribrdina Cyclicopora	. 156 88 138	pilosa Electrinidaeelegans, Palmicellaria	18 17 149
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicoporagigantea	. 156 - 88 - 138 - 139	pilosa Electrinidae elegans, Palmicellaria Steganoporella.	18 17 149 63
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi	. 156 88 138 139 138	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora	18 17 149 63 127
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa	. 156 88 138 139 138	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhipsopora (labellaris	18 17 149 663 127 112,127
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa	. 156 88 138 139 138 138	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus	18 17
enrtum, Rhynchozoon euspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata	. 156 88 138 139 138 138 135	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus elongata, Microporina	18 17 149 663 127 112,127
enrtum, Rhynchozoon euspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera	. 156 88 138 139 138 138 135 135	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella	18 17 149 63 127 112,127 84 68
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spimifera tenuiparietis.	. 156 88 138 139 138 138 135 135 136	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora	18 17 149 63 127 112,127 84 68
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis.	. 156 88 138 139 138 138 135 135	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella	18 17 149 63 127 112,127 84 68 58
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenniparietis. Cycloperiella rubra	. 156 88 138 139 138 135 135 136 136 136	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora eusifera, Smitta Entalophora fasejeuhfera	18 17 149 63 127 112,127 84 68 58 - 15 145 - 195
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spimfera tenuiparietis. Cycloperiella rubra	. 156 88 138 139 138 135 135 136 136 137 137 137	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhipsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora cusifera, Smittia Entalophora faseiculifera punctulata	18 17 18 18 18 127 112,127 84 68 58 - 15 145 - 195 195
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenniparietis. Cycloperiella rubra	. 156 88 138 139 138 135 135 136 136 137 137	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora easifera, Smittia Entalophora fasciculifera punctulata Erma	18 17 149 63 127 112,127 84 68 58 15 145 195 195 170 87
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spimfera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra).	. 156 88 138 139 138 135 135 136 136 137 137 193 52 152	pilosa Electrinidae Steganoporella Elhipsopora (labellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora casifera, Smittia Entalophora fasciculfera punctulata patagonica	18 17 149 63 127 112,127 84 68 58 15 145 195 195 17 87
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella	. 156 88 138 139 138 135 136 136 137 137 193 52 152	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora cusifera, Smittia Entalophora fasejeuhfera punctulata Erina patagonica Eschara bia perta	18 17 149 63 127 112,127 84 68 58 - 15 145 - 195 170 87 85
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella a viculifeta	. 156 88 138 139 138 135 135 136 136 137 137 137 193 52 152 152	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora ensifera, Smittia Entalophora faseiculifera punctulata Erina patagonica Eschara bia perta impressa	18 17 149 63 127 112,127 84 68 58 - 15 145 - 195 170 87 85 100 83
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella a viculifeta dactylus, Fedora (Discoflustrellaria).	. 156 88 138 139 138 135 135 136 136 137 137 137 193 52 152 152 191 97	pilosa Electrinidae clegans, Palmicellaria Steganoporella. Elhipsopora flabellaris cllipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora casifera, Smittia Entalophora fascienhfera punchulata Erina patagonica E schara bia perta impressa nobilis	18 17 149 63 127 112,127 84 68 58 15 145 195 195 197 87 85 100 83 83
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spimifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella aviculifeta dactylus, Fedora (Discoflustrollaria) Dakaria	. 156 88 138 139 138 135 135 136 136 137 137 193 52 152 152 191 97 97	pilosa Electrinidae Steganoporella Steganoporella Elhpsopora Glabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora casifera., Smittia Entalophora fasciculifera punctulata Erma patagonica Eschara biaperta impressa nobilis s pongites	18 17 149 63 127 112,127 84 68 58 -15 145 -195 195 -100 83 83 102
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella dactylus, Fedora (Discoflustrellaria). Dakaria chevreuxi grandis parviporosa	. 156 88 138 139 138 135 135 136 136 137 137 193 52 152 191 97 97	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora eusifera, Smitta Entalophora faseiculifera punctulata Erina patagonica. Eschara bia perta impressa nobilis s pongites eulgaris	18 17 149 63 127 112,127 84 68 58 - 15 145 - 195 170 87 85 100 83 83 102 108
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa. Cyclocolposa perforata spimifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella aviculifeta dactylus, Fedora (Discoflustrollaria). Dakaria chevreum grandis parvipiorosa torquata	. 156 88 138 139 138 135 135 136 136 137 137 137 199 52 152 152 191 97 97	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora ensifera, Smittia Entalophora fascientifera punctulata Erina patagonica E schara bia perta impressa nobilis s pongites enlgaris Escharella landsboroti.	18 17 149 63 127 112,127 84 68 58 -15 145 -195 195 -100 83 83 102
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella avienlifeta dactylus, Fedora (Discoflustrellaria). Dakaria chevreuxi grandis parviporosa torquata decorata, Calloporina	. 156 88 138 139 138 135 135 136 136 137 137 137 137 152 152 152 191 97 97 97	pilosa Electrinidae elegans, Palmicellaria Steganoporella . Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora ensifera, Smittia Entalophora fascienhfera patagonica Eschara biaperta impressa mobilis s pongites eugaris Escharella landsborovi. rostrigera	18 17 18 17 18 18 18 17 18 18 18 19 112 112 112 112 115 115 115 116 117 117 117 117 117 117 117 117 117
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spimicra tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella avienlifeta dactylus, Fedora (Discoflustrellaria). Dakaria clievreuxi grandis parviporosa torquata decorata, Calloporina deformis, Bracebridgia	. 156 88 138 139 138 135 135 136 136 137 137 137 193 52 152 152 191 97 97 98 98 112, 128	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora easifera, Smittia Entalophora fasciculifera patagonica. Eschara bia perta impressa nobilis spongites enlgaris Escharella landsboroit. restrigera ierruculata	18 17 149 63 127 112,127 84 68 58 15 145 195 195 170 87 85 100 83 83 102 102 104 147
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora	. 156 88 138 139 138 135 136 136 137 137 193 52 152 152 152 152 152 152 152 152 152	pilosa Electrinidae elegans, Palmicellaria Steganoporella . Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora ensifera, Smittia Entalophora fascienhfera patagonica Eschara biaperta impressa mobilis s pongites eugaris Escharella landsborovi. rostrigera	18 17 149 63 127 112,127 84 68 58 15 145 195 195 100 83 102 108 147 167
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa. Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella avienlifeta dactylus, Fedora (Discoflustrellaria) Dakaria clievreuxi grandis parviporosa. torquata decorata, Calloporina. deformis, Bracebridgia. delicattissima, Siphonoporella delicatula, Acanthodesia savarti	. 156 88 138 139 138 135 135 136 136 137 137 137 193 52 152 152 191 97 97 98 98 112, 128	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora easifera, Smittia Entalophora faseiculifera punctulata. Erina patagonica. E schara bia perta impressa nobilis s pongites culgaris Escharella landsborou rostrigera ierruculata Escharellidae	18 17 149 163 127 112,127 84 68 58 15 145 195 195 170 87 85 100 83 83 102 104 147 167 157 95 177 95
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spimicra tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella aviculifeta dactylus, Fedora (Discoflustrellaria). Dukaria chevreuxi grandis parviporosa torquata decorata, Calloporina. deformis, Bracebridgia delicatissima, Siphonoporella delicatula, Acanthodesia savarti Biflustra	. 156 88 138 139 138 135 136 136 137 137 137 199 52 152 152 152 152 152 152 152 152 152	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora easifera, Smittia Entalophora fasciculifera putagonica. Eschara biaperta impressa nobilis spougites eulgaris Escharella landsboroit. rostrigera ierruculata Eschareliidae Eschariua pesanserts torquata tumiduli	18 17 149 63 127 112,127 84 68 58 15 145 195 195 100 83 102 108 147 167 157 95 172 98
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella avienlifeta dactylus, Fedora (Discoflustrellana). Dukaria chevreum grandis parviporosa torquata decorata, Calloporina deformis, Bracebridgia delicatula, Acanthodesia savarti. Biflustra deutata, Membranipora	. 156 88 138 139 138 135 135 136 136 137 137 137 193 52 152 152 191 97 97 97 98 112,128 160 63 33	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella Terebripora ensifera, Smitta Entalophora faseienhfera punctulata Erina patagonica Eschara biaperta impressa nobilis spongites endgaris Escharella landsboroit. rostrigera ierruculata Escharellidae Eschariua pesanseris torquata tunidula iufgaris	18 17 149 63 127 112,127 84 68 58 - 15 145 - 195 170 87 85 100 83 83 100 147 167 157 195 172 98 98 98
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella avienlifeta dactylus, Fedora (Discoflustrellaria). Discoflustrellaria grandis parviporosa torquata decorata, Calloporina deformis, Bracebridgia	. 156 88 138 139 138 138 135 135 136 137 137 137 193 52 152 152 152 152 151 97 97 97 98 112,128 160 63 33 33 52	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora ensifera, Smittia Entalophora patagonica Erina patagonica Eschara bia perta impressa nobilis spongites enlgaris Escharelli landsboroit. rostrigera rerruculata Escharellidae. Eschariua pesanserts torquata tumidula tumidula eligaris Escharoldes terruculata.	18 17 18 18 18 18 18 18 18 19 112 112 112 112 112 115 115 115 110 117 117 117 117 117 117 117 117 117
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa. Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella aviculifeta dactylus, Fedora (Discoflustrellaria) Dakaria clievreuxi grandis parvipiorosa. torquata deorata, Calloporina. deformis, Bracebridgia. delicattissima, Siphonoporella delicatula, Acanthodesia savarti. Biflustra denticulata, Membrani pora denticulata, Cupularia Discoporella	. 156 88 138 139 138 138 135 135 136 136 137 137 137 193 52 152 191 97 97 97 98 112,128 160 63 33 33 52 79 80 73	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora ensifera, Smittia Entalophora faseienhfera punctulata Erma patagonica Eschara biaperta impressa mobilis s pongites eulgaris Escharella landsborott. rostrigera terruculata Escharila pesanseris torquata tunidula tunidula tulgaris Escharoides (ceruculata. Escharoides (ceruculata. Eugmoma	18 17 149 63 127 112,127 84 68 58 -15 145 195 195 170 83 83 102 108 147 167 157 95 172 98 98 98 108
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spinifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella avienlifeta dactylus, Fedora (Discoflustrellaria). Discoflustrellaria grandis parviporosa torquata decorata, Calloporina deformis, Bracebridgia	. 156 88 138 139 138 138 135 135 136 136 137 137 193 52 152 152 191 97 97 97 97 98 112,128 160 63 33 35 52 79 80 73	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Ellipsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora Entalophora fasciculifera punctulata Erma patagonica Eschara bia perta impressa mobilis s pongites culgaris Escharellidae Escharida landsboroit. rostrigera ierruculata Escharida escharisticulata Eschariolae Eschariolae Eschariolae Eschariolae Escharoides (cruculata yermiformis	18 17 149 63 127 112,127 84 68 58 -15 145 195 195 170 87 85 100 83 83 83 102 108 147 167 157 95 172 98 98 108 157 85
curtum, Rhynchozoon cuspidata, Cribrdina Cyclicopora gigantea mansfieldi multilamellosa Cyclocolposa perforata spimifera tenuiparietis. Cycloperiella rubra Cyclostomata cylindricea, Chaperia (Electra). Cystisella aviculifeta dactylus, Fedora (Discoflustrellaria). Diskaria grandis parviporosa torquata decorata, Calloporina. deformis, Bracebridgia delicatissima, Siphonoporella delicatula, Acanthodesia savarti Biflustra deutata, Membranipora denticulata, Cupularia Discoporella Hemiseptella	. 156 88 138 139 138 138 135 135 136 136 137 137 137 193 52 152 191 97 97 97 98 112,128 160 63 33 33 52 79 80 73	pilosa Electrinidae elegans, Palmicellaria Steganoporella. Elhpsopora flabellaris ellipticus, Corynostylus elongata, Microporina Rectonychocella. Terebripora ensifera, Smittia Entalophora faseienhfera punctulata Erma patagonica Eschara biaperta impressa mobilis s pongites eulgaris Escharella landsborott. rostrigera terruculata Escharila pesanseris torquata tunidula tunidula tulgaris Escharoides (ceruculata. Escharoides (ceruculata. Eugmoma	18 17 149 63 127 112,127 84 68 58 -15 145 195 195 170 83 83 102 108 147 167 157 95 172 98 98 98 108

	Pag	ge.		Page.
Enritina eurita		65	Flustrellaria texturata	32
Eurystomella		142	Foraminella	66
bilabiata	141,		lepida	65
foraminigera		141	foraminigera, Eurystomella	141
Eurystomellidae		141	fossulifera, Membranipora	20 56
eustomata, Microporella = -		124	fusifera, Floridinagaleata, Chaperia	
Reptoporina		124	Membranipora.	
excelsa, Fedora (Kionidella).		191	Gemelliporella	
excentrica, Orbitulipora	187,		asper	
exigua, Fenestrulina (Microporcda).		113	punetata	
exilis, Manzonella		61	vorax	
expansa, Idmonea (?)		200	Gephyrophora	
falcata, Crateropora		65	germana, Membranipora	
Farcimia oculata		65	Conopeum	
tenella		55	gibbera, Microporella	
fasciculifera, Entalophora		195	gibbosa, Hippoporina. gigantea, Cianotremella.	
Tubulipora	186.	197	Cyclicopora (?).	
Fedorabidentata		191	giganteum, Aspidostoma	
erassilabris		191	glomerata, Theonoa	
daetylus		191	grande, Membrendoecium	
· · · · · · · · · · · · · · · · · · ·		191	grandicella, Hemiseptella	
excelsa		191	Rhynchozoou	
minutissima		191	Smittina	
obliquescriata		191	grandis, Dakaria.	
persimplex		191	granulata, Thalamoporella	
protecta		191	granulosa, Hemiseptella	. 74
simplex		191	Mastigophora	
smithi		191	Rhamphostomelia.	
feegeensis, Hippopolima.		163	guernei, Callopora	
Fenestrulina		113	Membranipora	. 42
exigud		113	guineensis, Cupularia	. 29
malusi	.12, 115,		haddoni, Cheiloporina	163, 169
parvipora	113,		haidingeri, Cupularia	
porosa proxima	112		Lunulites	
filmargo, Hemiseptella			hastigera, Diporula	
Filisparsa		195	heckeli, Adcona	
clarki		195	Cellepora	
parvula		196	Microporella.	
filum, Callopora		42	Hemiscptella	
Membranipora		42	denticulata	
fissa, Schizotheca		153	filimargo	
tissurifera, Cellaria	-	85	fistnla	
Microporella		121	grandicella	
tistula, Hemiseptella	٠	73	granulosa	
Membrani pora	1.101	73 ,127	labiata	
flabellata, Diaperoccia	112	202	lata	
Membranipora		20	michaelsem	
Stathmepora		201	minor	
flabelligera, Microporella		118	planulata	
flabellum? Berenicea		194	rectangulata	
Diastopora		194	steganoporoides	
Floridina		56	tennis	
fusifera		56	tuberosa	. 7
minima		57	hemispherica, Holoporella	
parvicella		57	hermanni, Microporella	
pyripora		56	Puellina	
regularis floridina, Schizopodrella		57	Reptescharella	
Flustra Dumerilii		106 40	Reptescharellina,	12
savarti		31	Heterooccium amplectens.	
tehuelca		22	Heteropora tortilis.	
tuberculati		22	Heteroporidae	
***	127	,128	hexagona, Microporella	. 12
marginata		.127	hexagonum, Rhagasostoma	6

	Page,		Page.
hians, Odontionella (Membranipora)	66	ldmonea planula.	199
Hincksina	35	imperati, Schizellozoon	153
multispinata,	38	impressa, Calpensia.	68, 83
	38	Eschara	, 83
quadrispinosa		Micropora.	83
Hippaliosina	165, 166		149
brevirostris	165,166	inermis, Palmicellaria.	_ 119
clavula,	165	inflata, Microporella	
depressa	165	informata, Celle pora =	102
laxipora	165	Reptocelleporaria	102
rostrigera	163, 166, 167	infratelum, Hippomenella	132
sandbergeri	. 165, 166	innominata, Cribrilina radiata.	90
Hippellozoon novaezelandiae	. 153	Le pralia	90
Hippodiplosia	. 131	Puellina	90
baccata	131	inversa, Inversiula	112
bigiberra	131	inversa (Microporella).	129
	132	Inversiula	113, 129
Hippomenella		airensis	. 129
infratelum	132	inversa	112, 129
Hippopodina feegeensis	163		129
Hippopodinidae	162, 163	nutrix	129
Hippoporella	132	quadricornis	
costulata	133	isabelleana, Hippothoa	103
papulifera	133	Stylopoma.	103
spinosa	132	johnsoni, Cupularia	77
Hippoporina	129	labiata, Hemiseptella =	68, 70, 72, 89
	130	Phidolopora	. 153, 154
gibbosa	131	labiatus, Corynostylus.	. 68, 84
lata	129	Labiopora	, 67
pusilla		crenulata	65
vestita	_ 130		67
Hippothoa	92	miocenica	191
hyalina	92	Lacerna	99
rugosa	94	mucronata	79 70
isabellcana	103	lacinia, Hemiseptella	
mucronatu	99	lacroixii, Conopeum	_ 26
pesanscris	172	Membranipora	22
s pongites	102	lacrymosum, Metrarabdotos.	164
	_ 92	Lagenipora	171
Hippothoidae	203	brevicollis	171
hispida, Discopora	203	spinnlosa	171
Lichenopora		Laminopora	160, 161
Holoporella	174	arbuscula	. 161
albirostris	_ 174	contorta	161
aviculifera	179	miocenica	160
bicornis	178	Janceolata, Callopora	41
echinata	177	Schismopora.	181
hemispherica	_ 176	-	147
massalis	177	landsborovi, Escharellu	40
mucronata	179	lata, Callopora dumerilu .	70
orbifera	= 177	Hemiseptella	131
parvula	175	Hippoporina	151
rostrifera	175	laticella, Rhamphostomella	96
	176	latisinuata, Schizoporella	
subturrita	179	laxipora, Hippaliosina	163
turrita	178	Leiosella	142
umbonata	61	edax	_ 142
holostoma, Woodipora		lenticularis, Orbitulipota.	187, 189
horrida, Callopora	43	lepida, Foraminella	Gã
Membranipora	43	Lepralia	133
Tremogasterina	153, 168	1 11	147
hyadesi, Microporella	118	1.5 months	100
hyalina, Celle pora	92		143
Celle porclla	92		10
Hippothoa	92		12
	92		_ 13-
Le pralia	94	1	9:
rugosa, Hippothea	92	T. There	
Schizoporella	198	the control of the control of	91
Idmonea	199		18
californica	195	21	×
dispar		C. Comm	13
expansa	200		*
- ila avug	202	Print but a	

		Page.		Page.
Lepralia reversa.		148	Membranipora circu metathrata	4:
trispinosa		143	cul prusis	24
turrita		179		
unicornis			deutata	51
		105	dumerilii	40
riolacea		158	filu m	40
vulgaris		108	fistula	77
Lepraliella contigua		153	flabellata	20
levigatunn, Rhynchozoon (?)		157	fossulifera	21
Lichenopora		203		
californica		203	galinta	52
			germaua	21
hispida		203	guernei	42
radiata		204	horrida	43
verrucana		205	łacinia	70
Lichenoporidae		203	lacroixii	22
ligulata, Cribrilina		88	magnilabris	63
jimosa, Acanthodesia (Membrampora)		30		
			membranacea	18
longicanda, Spathipora		16	monostachys	17
longirima, Spathipora		16	nitidula	27
longirostrata, Schizomavella		109	oblougula	34
lorea, Palmicellaria (Eschara)		149	osburni	24
lowei, Cupuluria		80		
Luunlites denticulata		79	parvula	37
			reyfi	31
haidingeri		77	savarti	31
u mbellata		80	speciosa	41
mavulata, Cellepora		182	spiculata	21
Lepralia		182	tchuclea	22
Selenaria		59	fuberculata	22
and the first of t		63		
			tuberimargo == = =	23
Steganoporella		63	vaughani	23
magniporosa, Schizoporella		95	Membraniporidra	39
Stylopoma		103	parca	39
major, Mucronella spinosissima		170	Membraniporina	24
Phylactella spinosissima		170		
Malacostega		11	baccata	25
			ealifornica	25
maleposita, Smittina		144	tenella	24
malusi Cellepora		115	vincularina	26
Fenestrulina	112,11	5,116	Marilla Andrews	36
Microporella		115		
Mamillopora		191	grande	36
tuberosa		192	parvicapitatum	36
			mesleri, Proboscina	193
mandibulata, Cellaria		86	Metracolposa	92
mansfieldi, Cyclicopora?		138	ninci onata	92
Manzonella		60		
exilis		61	Metrarabdotos	162
marginata, Flustramorpha	111	2,127	aurieulatum	= 164
Microporella	***	118	colligatum	162
		107	lacrymosum	164
Schizopodrella			moniliferum	163
mursupiam porifera, Porella		147		
marylundica, Lepralia		87	michaelseni, Hemiseptella	70,73
massalis, Cellepora		177	Micropora	
Holoporella		177	corracea	. 55 59
Mastigophora		172	im pressa	\$3
			·	
dutertici		172	Microporella	
granulosa		172		121
pesanseris		172	californica	123
Mecynoeciidae		195	ciliata 112, 113	5, 118, 119
Melicerita		87	coronata	
charlesworthi		85	divarieata	
Melobesia rudiata				
		204	eustomata	
membranacea, Membranipora		18	fissurifera	
Membranipora		20	flabellaris	115
annulus		52	flabelligera	. 118
burbarensis		28	gibbera	
bifoliata		33	hastigera	
californiensis		43		
сатінова		54	heermannt =	125
canariensis		25	hexagona	129

	Page.		Page,
Microporella hyadesi	115	oblonga, Verminaria	68
inflata	119	oblongula, Acanthodesia.	. 31
malusi	115	Membrani pora	
marginata.	118	occidentalis, Tubulipora.	198
personata	118	oculata, Farcimia	55
praeciliata .	119	Nellia	55
rndis	118	occultata, Odontionella	, 65,66
rugosa	118	Odontionella	64,66
tessellata	122		66
umbonata	123	hians	
		oceulata	65,66
verrucosa	118	Ogivalina	37
vibraculifera	124	mutabilis	
violacca	158	parvnla	37
Microporellae	112, 113	Oncousoeciidae	195
Microporina	. 3	Opesiulidae	56
elongata	68	ophidiana, Smittia	
milneana, Diaperoccia	202	Smittina .	144
Idmonea	202	onulenta, Temachia.	173
minima, Floridina	57	orbifera, Holoporella Orbitulipora	177
minor, Hemiseptella (Membranipora)	70	Orbitulinora	186 187 189
minuta, Cellepora	182	excentrica.	187, 189
Stylopoma	101	C CCIMITICA.	
		lentienlaris.	
minutissima, Fedora (Lepralia)	191	petiolus Orbituliporidae	187,189
miocenica, Labiopora	67		186
Laminopora	160	osburni, Membranipora	
moekleri, Mystriopora	18	otophorum, Pachykvaspedon	108
monilifera, Acanthodesia savarti	32	ovale, Conopeum	. 26
moniliferium, Metraralidotos	163	Pachykraspedon otophorum	108
Triphyllozoon	153	pacifica, Crisia	196
monostachys, Electra	17	Phidolopora	154
Membrani pora	. 17	Retepora	154
montifera, Lepralia	. 134	Terebripora	. 15
mueronata, Hippothoa		Palmicellaria	149
Holoporella	179	convoluta	150
Lacerna			150
	92	costulata	149
Metracolposa		elegans	
Mucronella spinosissima var major	170	(Eschara) lorea.	149
Multicrescis tortilis	206	inermis	149
multilamellosa, Cyclicopora	138	punetata	150
multipora, Callopora(?)	. 41	skenei	149
Siphonella (Flustrellaria)	41	tenuis	
Multiporina umbilicata	138	palulosa, Chaperia	52
multiradiata, Batopora	189	papulifera, Hippoporella	
multispinata, Cupularia	76,75	m . 11	107
Hineksina	38	parca, Membraniporidra	39
Ramphonotus	47	parvicapitatum, Membrendoecium.	
	0.00	parvicella, Floridina	
mutabilis, Ogivalina	. 105		
Schizopodrella		Steganopotella	
Myriozoella		Terebripora	15
crustaeea	185, 186	parvipora, Fenestrulina (Mieroporella).	116
Myriozoidae	185		
Myriozoum	184, 186	parviporosa, Dakaria	
coarctatum	15	parvirostris, Callopora	41
subgracile	. 185	parvispina, Chaperia	54
truneatum	184	parvula, Holoporella	175
Mystriopora	19	Membrani pora	37
(?)arcolata	_ 19	Ogivalina	37
möckleri	18	patagonica, Erina	\5ā
	55	perforata, Cyclocolposa.	. 135
Nellia	5.5	Peristomellae	
oculata	55	persimplex, Fedora (Stichoporma)	191
tenella	27		
nitidula, Membranipora		personata, Microporella	172
Conopeum?	27	pesanseris, Escharina	172
nobilis, Eschara	83	Hippothon	172
novaezelandiae, Hippellozoon.	153	Mastigophora.	. 172
nutrix, Inversiula	129	Schizoporella	
obliquescripta Fedora (Kionidella)	191	petiolus, Orbitulipora	= = 187, 189

300 INDEX.

	F	age.		Page.
Phidolopora =		154	Reptescharella carolinensis.	. 90
labiata.	15-	4, 153	herrmanni.	×5
pacifica .		154	Reptescharellina cornuta	100
Phylactella		170	dis parilis.	. 5
spinosissima major		170	heermanni	
Phylactellidae		170	Reptocelleporaria informata	
pilosa, Electra		18 199	Reptoporina customata	
planulat, Hemiseptella		74	Reteporabeaniana	
poissoni, crinispina, Crepidacantha.		174		
Porella		147	pacifica	
bella		147	Reteporidae	152
collifera		148	reticulata, Smittina	1.1.
marsupium poritera		147	reussiana, Cupularia	
reversa		148	renssi, Stichoporina	1.90
porifera, Porella marsupium.		147	reversa, Lepralia(?)	. 14
Smittina		147	Porella	14
porosa, Adeona		158	reyti, Acanthodesia savarti	
Fenestrulina		117	Mcmbranipora	
porosus, Cauloramphus		45	Rhagasostoma hexagonum	- fi2
praeciliata, Microporella		119	Rhamphostomella	
Proboseina		193	granulosa	
mesleri		193	laticella	
protecta, Fedora (Stichoporina).		191	Rhynchozoon	
proxima, Fenestrulina Psilosolen	113	3, 116	angulatumeurtum	
capitiferax		207 207	grandicella	$\frac{150}{156}$
Puellina		207	levigatum	157
crassilahiata		91	vaughani	155
herrmanni		89	verruculatum	
innominata		90	robertsoniae, Costazzia	
radiata carolinensis.		90	Cupularia	
rarecosta		90	rostrifera, Holoporella	175
seripta		59	rostrigera, Escharella	167
punctata, Cribrilina		87	Hippaliosina	66, 167
Gemelliporella		111	rosula, Batopora	
Lepralia		57	rozieri, Thalamonorella	61
Palmicellaria		150	rubra, Cycloperiella	137
punctulata, Entalophora		170	rudis, Microporella	118
minor, Tubucellaria.		170	rugosa, Hippothoa hyalina	
Tubucellaria		170	Microporella	
pusilla, Hippoporina Schizopodrella		129 106	sandbergeri, Hippaliosina	
Vibracellina		35	savarti, Acanthodesia	
Pyripora		19	Riflustra	
brevieauda		19	bifoliata, Acauthodesia	
catenularia		18	delicatula, Acanthodesia	
pyripora, Floridina		56	Flustra	
quadricormis, Inversiula (Microporella).		129	Membrani pora	
quadrispinosa, Hineksina		38	monilifera, Aeanthodesia	32
radiata, Aimulosia		140	reyti, Aeanthodesia	
Lichenopora		204	texturata, Acauthodesia	
Melobesia		204	typica, Acanthodesia	
rarecosta Puellina		90	scandens, Calloporina (Microporella)	
scripta, Puellina		89	Sehismopora	
Ramphonotusagellus		46	abrupta	
asperus		47 46	breviucisa lanccolata	
multispinatus.		47	Schizellozoon imperati	
rarecosta, Cellepora		90	Schizolavella	105
rectangularia, Aeanthodesia		34	vulgaris	
rectangulata, Hemiseptella.		74	Schizomavella	
Reetonychocella		5.5	longirostrata	
elongata.		38	Sehizopodrella	
regularis, Floridina		57	aculeata	
renipuneta, Calloporina (Microporella)		128	doverensis	107
Reptadeonella violacea		159	floridina	
reptans, Atelesopora		194	marginata	107

INDEX. 301

	Page.		Page	e.
Schizopodrella mutabilis.	_ 105	Steganoporella	(62
pusilla	_ 106	$\epsilon l \epsilon gans{}$		63
unicornis	. 105	magnilabris		63
Schizoporella bia perta		parvicella		62
cumulata		Steganoporellidae		62
floridina		steganoporoides, Hemiseptella.	70,	
hyalina		Stephanosella		99
latisinuata		biaperta		99
magniporosa			186, 1	90
pesanseris		tuberosa		92
spongites		Stomachetosellidae.		42
subquadrata		striatopora, Crisina?		200
Schizoporellae	95	Stylopoma		01
Schizorthosecos	186, 190	isabelleana		03
Schizotheca fissa		magniporosa		103
seripta, Cellepora		minuta		101
Puellina radiata		spongites	1	102
Selenaria		subgracile, Myriozoum	1	183
aurieularia		subquadrata, Schizoporella	1	105
Semitubigera tuba	10.7	subtorquata, Cellepra		9%
errata, Crisia		subturrita, Holoporella	3	176
simplex, Fedora (Stichoporina)		Tegminula]	151
siucera Cheilopora.		tchuclea, Flustra		22
sinefilum, Terebripora		Membranipara		22
sinuosa Cellaria		Temachia		173
Siphonella (Flustrellaria) multipora		opulenta		173
Siphonoporella		tenclla, Callopora		24
delicatissima	63	Cellaria		55
skeuei, Palmicellaria (Millepora)		Farcimia		55
smithi, Fedora (Mamillopora)		Membraniporina		24
Smittia bella	147	Nellia		55
californiensis	146	tenuiparietis, Amphiblestrum		45
ensifera	145	Cyclocolposa		133
ophidiana	145	tenuis, Hemiseptella (Membranipora)		70
trispinosa	143	Palmicellaria		149
Smittina	143	Terebripora		15 15
brevis	146	elongata		15
ealifornieusis	146	pacifica		15
discoidea		parvicellasinefilum		15
grandicella	145	Terebriporidae		15
maleposita	144	tesselata, Cellepora		122
ophidiana	144	Microporella		122
porifera	147	texturata, Acanthodesia savarti		32
reticulata	144	Flustrellaria		32
trispinosa	. 143	Thalamoporella		61
Smittinidae	143	biperforata		62
Spathipora	16	granulata		61
cucullata	16	rozieri		61
longicanda		Thalamonorellidae		60
longirima	4	Theonoa		201
speciosa, Callopora(?)		glomerata		201
speciosa, Membranipora	186 190	Theonoidae		201
Sphaerophora spiculata, Membranipora	21	torquata, Dakaria		il.
spiculata, Membranipora.	136	Escharina		37
spinifera, Cyclocolposa (?)spinosa, Hippoporella	132	tortilis, Heteropora		206
spinosissima major, Mucronella		Multicrescis		206
spinosissima major, Micronella	170	Tretocycloecia		167
spinulosa, Lagempora	171	Tremogasterina	1.53	
spongites, Cellepora	102	horrida	. 100	206
Eschara	102	Tretocycloecia		206
Hippothoa	102	tortilia		206
Schizonorella	102	and the state of t		206
Stylonoma	102	1 -1 Cauloromphus		48
Stathmenora	201	and the second of the second o		153
flabellata	201	Tubulana mana		

	Page.		Page.
trispinosa, Lepralia.	143	umbonata, Microporella	123
Smittia	143	Unicavea californica	203
Smittina.	143	unicornis, Lepralia	105
tropica, Chaperia	52	Schizopodrella	105
truncatum, Myriozoum	184	vaughani, Membranipora	23
Trypematella	134	Rhynchozoon	. 155
papulifera	135	venusta, Trypostega	, 95
Trypostega	95	vermiformis, Euginoma	
venusta	95	Verminaria	. 83
tuba, Semitubigera	198	oblonga	. 68
Tubulipora	198	verruearia, Lichenopora	. 205
tuberenlata, Flustia	22	verrucosa, Diporula	112,125
Membranipora		Microporella (Diporula)	
tuberimargo, Membranipora (?)		verruculata, Celleporu	. 157
tuberosa, Hemiseptella		Eschare!la	
Mamillopora		Escharoides	
Sticho por ina		verruculatum, Rhynchozoon	
Tubucellaria		vestīta, Hippoporina (?)	
punetulata	170	Vibracellina	. 35
var minor		pusiHa	
Tubuccllariidae	170	simplex	
Tubulipora	170	vibraeulifera, Microporella	
fasciculifera	197	Microporella ciliata	. 124
occidentalis	198	vincularina, Membraniporina	. 26
tuba	198	violacea, Adeona	
Tubuliporidae	197	Lepralia	. 158
tumitula, Escharina	95	Microporella	. 158
turrita, Holoporella	179	Kepladeonella	. 159
Le pralia	179	virginiana, Ceriopora	. 193
typica, Acanthodesia savarti	31	vorax, Gemelliporella	. 111
umbellata, Cupularia	, 76, 80	vulgaris, Escharina	. 108
Lunulites	80	Lepralia	
umbilicata, Adeonellopsis	158	Schizolavella	108, 109
Cellepora		Watersipora cuculata	. 163
Multiporina	158	Woodipora	
ambonata, Heloporella		holostoma	. 61

ADDITIONAL COPIES

OF THIS PUBLICATION MAY BE PROCURED FROM
THE SUPERINTENDENT OF DOCUMENTS
GOVERNMENT PRINTING OFF AE
WASHINGTON, D. C.
AT
75 CENTS PER COPY

PURCHASER AGREES NOT TO RESELL OR DISTRIBUTE THIS COPY FOR PROFIT.—PUB. RES. 57, APPROVED MAY 11, 1922 $\hfill \bigtriangledown$















