

Research



Cite this article: Rodríguez-Malagón MA, Camprasse ECM, Angel LP, Arnould JPY. 2020 Geographical, temporal and individual factors influencing foraging behaviour and consistency in Australasian gannets. *R. Soc. Open Sci.* **7**: 181423. <http://dx.doi.org/10.1098/rsos.181423>

Received: 28 August 2018

Accepted: 1 May 2020

Subject Category:

Ecology, conservation, and global change biology

Subject Areas:

behaviour/ecology

Keywords:

foraging behaviour, behavioural consistency, GPS tracking, foraging ecology, Australasian gannet, accelerometry

Author for correspondence:

John P. Y. Arnould

e-mail: john.arnould@deakin.edu.au

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.4979942>.

Geographical, temporal and individual factors influencing foraging behaviour and consistency in Australasian gannets

Marlenne A. Rodríguez-Malagón, Elodie

C. M. Camprasse, Lauren P. Angel

and John P. Y. Arnould

School of Life and Environmental Sciences (Burwood Campus), Deakin University, Geelong, Victoria 3220, Australia

Foraging is a behaviour that can be influenced by multiple factors and is highly plastic. Recent studies have shown consistency in individual foraging behaviour has serious ecological and evolutionary implications within species and populations. Such information is crucial to understand how species select habitats, and how such selection might allow them to adapt to the environmental changes they face. Five foraging metrics (maximum distance from the colony, bearing from the colony to the most distal point, tortuosity index, total number of dives and mean vectorial dynamic body acceleration) were obtained using GPS tracking and accelerometry data in adult Australasian gannets (*Morus serrator*) from two colonies in southeastern Australia. Individuals were instrumented over two breeding seasons to obtain data to assess factors influencing foraging behaviour and behavioural consistency over multiple timescales (consecutive trips, breeding stages and years) and habitats (pelagic, mixed pelagic and inshore, and inshore). Colony, breeding stage and year were the factors which had the greatest influence on foraging behaviour, followed by sex. Behavioural consistency, measured as the contribution of the individual to the observed variance, was low to moderate for all foraging metrics (0.0–27.05%), with the higher values occurring over shorter timescales. In addition, behavioural consistency was driven by spatio-temporal factors rather than intrinsic characteristics. Behavioural consistency was higher in individuals foraging in inshore than pelagic habitats or mixed pelagic/inshore strategy, supporting suggestions that consistency is favoured in stable environments.

1. Introduction

Foraging is a primary activity of animals which can be highly influenced by intrinsic factors such as age, sex or genotype [1–3], extrinsic factors such as geographical location, local weather or predation risk [4–6], and by reproductive constraints such as breeding stage or brood size [7,8]. However, such factors do not necessarily affect all animals in the same way, with one or multiple influences potentially acting in different directions at a particular time [9]. Furthermore, as foraging is generally a high energy expenditure activity, there are strong incentives for animals to develop foraging strategies that minimize their energy costs [10]. Such strategies can vary in associated foraging time and effort in particular habitats, associated choice of specific search methods and/or of food types consumed [11]. If a particular foraging strategy provides greater rewards, it is likely that this strategy will be repeated over time, favouring the development of behavioural consistency [12]. It is known that behavioural consistency in foraging activities leads to the evolutionary development of foraging specialization within animal populations, but the information on the persistence of this phenomenon over different timescales and habitats is limited [13].

Foraging specialization refers to the use of a specific proportion of the full range of available resources (or foraging strategies) by a subset of a population, resulting in inter-individual niche variation [14]. This phenomenon has been demonstrated in a wide variety of taxa [15]. Information on the factors influencing behavioural consistency (e.g. extrinsic versus intrinsic factors) and on the link between habitat selection (e.g. pelagic versus benthic foraging) and behavioural consistency is lacking; however, foraging specializations are thought to arise in stable environments in which resources are predictable and diverse, enabling individuals to develop behavioural differences to reduce niche overlap with conspecifics and, thus, minimize competition [16]. Such behavioural consistency may, therefore, have significant ecological consequences at the individual level but also on the development of offspring during the breeding season [17]. Consequently, knowledge of foraging temporal and spatial variation in specializations is important to fully understand their ecological implications within species [18,19].

The marine environment is complex and dynamic, and the foraging ecology of marine life is highly influenced by environmental variables [20]. At a global scale, oceans display clear patterns of water circulation and climate [21]. At local scales, physical features such as bathymetry, tidal regimes and nutrient fluxes determine the structure of marine and coastal ecosystems and influence the behaviour of marine fauna [22]. Marine environments comprise different ecosystem types and biomass levels which can lead to the development of a wide range of foraging techniques (higher ecological opportunity) [16,23] even within the same species and populations [24]. Behavioural consistency in foraging activities have been found within different animal groups in the marine environment [25,26], and it is expected to occur more commonly for top-order marine predators who are regulated by bottom-up processes and experience high levels of resource competition [27,28].

Marine birds are important top-order predators [29,30]. They are long-lived animals and, during the breeding season, adopt a central place foraging strategy which can lead to high levels of resource competition [31]. These attributes have been shown to favour the development of behavioural consistency within this group and, combined with other factors such as age, sex or breeding status, influence the development of individual behavioural differences [32]. However, the degree to which species and populations exhibit individual behavioural consistency in foraging activities can vary [33]. Studies suggest that intra- and inter-population differences may be related to temporal changes in resource availability [32,34,35], but the mechanisms influencing individual foraging consistency across populations or habitats are poorly understood [36]. Such information is crucial to enable predictions about marine predators' habitat selection and the responses of natural populations to changing environments [18,37].

The Australasian gannet (*Morus serrator*) is an important marine predator in southeastern Australia and New Zealand [38,39], with an estimated annual consumption of 228.2 tons of schooling pelagic fish (e.g. Australian sardine *Sardinops sagax*, barracouta *Thyrstites atun* and blue mackerel *Scomber australasicus*) in Australian waters alone [40]. This region is one of the fastest warming oceanic areas and significant changes to ocean currents are predicted to occur [41,42]. Such changes are likely to alter the distribution and abundance of marine species [43,44]. Indeed, expansions and shifts in fish and invertebrate species ranges have already been documented in southeastern Australia [45]. Therefore, knowledge of the factors influencing foraging activity and behavioural consistency in Australasian gannets is necessary to understand how their populations may adapt to changes in the supply of marine resources.

Like other members of the Sulidae family, the Australasian gannet is considered a generalist forager and has been shown to be adaptable in its feeding habits [40,46,47]. It displays reverse sexual

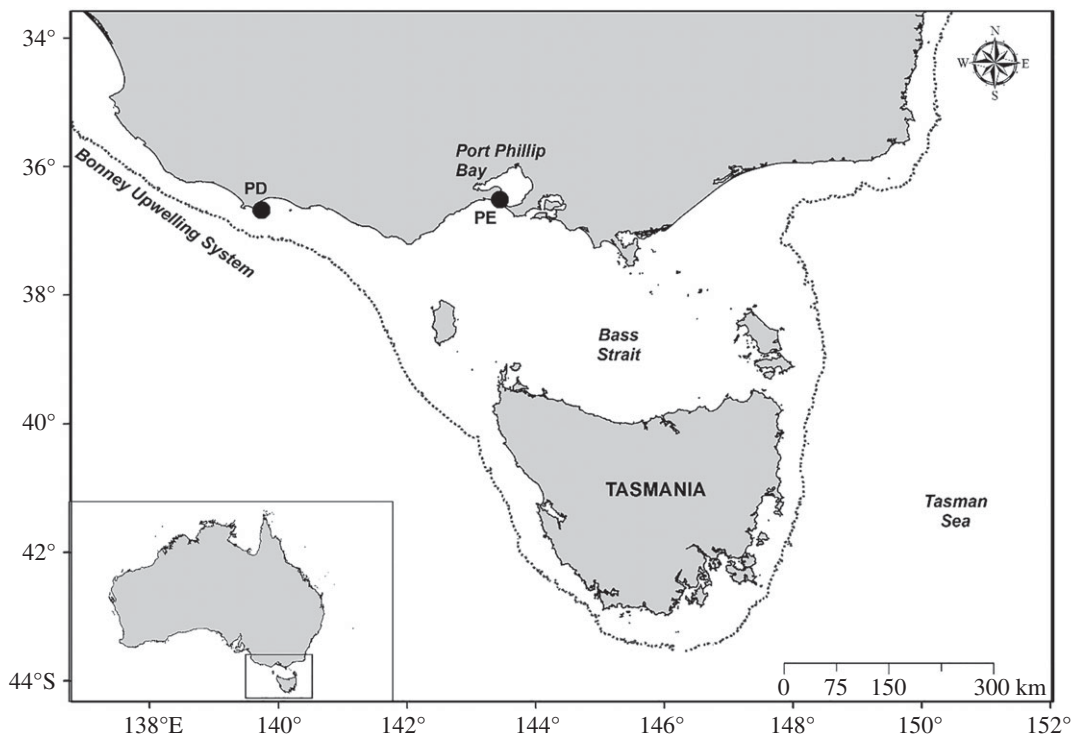


Figure 1. Location of study sites: Point Danger (left) and Pope's Eye (right). The 200 m bathymetric contour is given to indicate the edge of the continental shelf.

dimorphism (females larger than males) and recent studies suggest individuals exhibit sex-related differences in habitat use [48–50] and prey selection [47]. Furthermore, inter-colony differences in diving behaviour and habitat selection have been documented [51,52]. However, little is known of the factors influencing foraging behaviour and behavioural consistency in this species. The aims of the present study, therefore, were to determine, in Australasian gannets: (i) the influence of extrinsic and intrinsic factors on foraging behaviour, (ii) the degree of behavioural consistency in foraging activities, (iii) the persistence of behavioural consistency through time and space, and (iv) the influence of extrinsic and intrinsic factors on behavioural consistency.

2. Material and methods

2.1. Study sites and animal handling

The study was conducted at two Australasian gannet breeding colonies in northern Bass Strait, southeastern Australia, which experience divergent oceanographic conditions and may present differences in resource availability or habitat accessibility (figure 1). Point Danger (PD; 38°23'36.09" S, 141°38'55.94" E) is located at the western edge of Bass Strait near the seasonally active (Austral summer) Bonney Upwelling, an important source of primary productivity for the Bass Strait region [22,53]. Individuals from this colony range up to 238 km northwest and southeast, remaining over the narrow (approx. 40 km wide) continental shelf to forage on schooling fish and cephalopods [22,52]. Pope's Eye (PE; 38°16'35.88" S, 144°41'56.21" E) is located at the entrance of the Port Phillip Bay (PPB) on an artificial structure. Birds from PE forage within the shallow (average depth less than 13.6 m) [54] waters of PPB primarily on benthic/demersal fish, outside of PPB within northern Bass Strait on schooling fish and cephalopods, or in both habitats [49,52].

Data were collected during the 2014/15 and 2015/16 breeding seasons (October–March) in each of three breeding stages: incubation; early chick-rearing (chick age 0–50 days); and late chick-rearing (chick age greater than 50 days) [55]. Individuals were captured by hand or with the aid of a noose-pole [56] at the nest and weighed in a cloth bag with a suspension scale (± 25 g, Salter Australia Pty Ltd, Australia). A GPS data logger (programmed to record location every 2 min; I-gotU GT-600, Mobile Action Technologies Inc., Taiwan, ± 10 m error), and a tri-axis accelerometer data logger

(sampling rate of the individual 25 Hz; X8M-3mini, Gulf Coast Data Concepts LLC, USA), encapsulated together in heat shrink plastic (total package 53.7 g, <3% body mass), were then attached to the central tail feathers of the individual using water-proof tape (Tesa® 4651, Beiersdorf AG, Germany). The instrumentation was done in a consistent manner, with the GPS logger at the top, towards the head of the animals, and the accelerometer at the bottom towards the tail. Individuals were then returned to the nest and resumed natural behaviours within 10 min of capture.

After 10–12 days, individuals were recaptured as previously described and the data loggers were removed by peeling the tape from the feathers, and body mass was recorded. Morphometric measurements of culmen length and bill depth, and tarsus length and ulna length, were taken using Vernier callipers (± 0.1 mm) and metal ruler (± 1 mm), respectively. A blood sample (0.1 ml) was then obtained by venipuncture of a tarsal vein for genetic sexing (DNA Solutions, Wantirna South, Victoria, Australia) before the bird was returned to the nest. Where possible, the same individuals were sampled in multiple breeding stages and across years.

2.2. Data processing and statistical analysis

Unless stated otherwise, all data processing and statistical analyses were conducted in R v. 3.3.2 [57]. Deployment data were checked by visual inspection, and split into individual foraging trips using the return of the birds to the colonies' coordinates as endpoints for each trip. Trips were then filtered using a speed filter to remove erroneous locations [58] in the *trip* package [59], applying a maximum average speed of 55 km h^{-1} suggested for northern gannets (*M. bassanus*) [34]. Subsequently, for each foraging trip, maximum distance from the colony (km), total distance travelled (km) and bearing ($0\text{--}360^\circ$, from the colony to the most distal point) were calculated using the *adehabitatHR* package [60]. A tortuosity index, a measure of an animal's searching behaviour, was also estimated by dividing the maximum distance reached from the colony with the total distance travelled during the trip [61,62].

At-sea behaviours throughout the foraging trip were determined from the tri-axis accelerometer data loggers. Data were initially inspected visually to assign foraging behaviours (plunge diving and surface foraging) in IGOR Pro (v. 6.37, WaveMetrics, USA) [63], based on the acceleration profiles suggested for other species of gannets and boobies [64–66]. The *Ethographer* package was then used to identify these behaviours by performing a *k*-means, unsupervised cluster analyses of 1 s windows of continuous wavelet spectra computed from the time series. Later, each identified cluster was assigned a specific behaviour based on the previous visual identification [67]. From these data, the total number of dives (plunge diving and surface foraging) was estimated for each foraging trip. In addition, the accelerometry data were used to calculate the average vectorial dynamic body acceleration (VeDBA) throughout each foraging trip. This average was used as a proxy of the energy expenditure and allowed comparisons of the rate of energy expended across foraging trips [68–70].

A body condition index (BCI) was calculated for each bird at each deployment, as a proxy for total body fat (%) content, using body mass (kg), wing ulna (mm) and tarsus (mm) measurements [48]. As Australasian gannets are sexually dimorphic, body size indices were calculated to investigate the effect of size on foraging behaviour independently of sex. A body size index (BSI) and wing length index (WLI) were calculated using the deviation of each individual's body mass (kg) and wing length (mm) from the means for their respective sex.

To determine the factors influencing the foraging behaviour of instrumented individuals, linear mixed effects models were created using the *nlme* package [71]. The foraging metrics (maximum distance from the colony, average bearing and tortuosity index), the total number of dives and mean VeDBA, were used separately as response variables. Fixed factors such as colony (PD, PE), year (2014/15, 2015/16), breeding stage (incubation, early chick-rearing, late chick-rearing) and sex were used as explanatory variables in combination with the BCI, BSI and WLI. For these models, the full dataset was used (i.e. data obtained from all individuals in both sites and 2 years of sampling) using sampling size of three or more foraging trips per deployment. In this study, the influence of the factors colony, breeding stages and year were considered to reflect the differences in resource availability and environmental variation, respectively. Consequently, specific environmental variables were not analysed.

Where appropriate, variables were cube-root-transformed to fit model assumptions of constant variance and normal distribution of residuals [72]. Model assumptions were checked by plotting residuals and using quantile–quantile plots. Collinearity among all the explanatory variables was checked before conducting each model using pairplots, boxplots and the variance inflation factor (cut-off value used = 2) [73]. The initial models were then fitted with restricted maximum likelihood (REML) and models with and without the random structure (nest identity, due to the use of breeding partners

from the same nests, and individual identity) were compared using the *anova* function. Variance structure for the explanatory variables was included when the residuals inspection suggested it was necessary. The best-fixed structure was found using the *dredge* function of the *MuMIn* package based on the AICc values [74], using models refitted with maximum likelihood (ML). Where multiple models had $\Delta\text{AICc} \leq 4$ and no single model had an AICc weight above 0.90, model averaging was used to calculate the relative importance of each explanatory variable using the *MuMIn* package [74,75]. This multi-model statistical approach was selected as it allows to identify strong associations between multiple explanatory variables, while AICc values compare multiple models all at once incorporating model selection uncertainty and enabling inferences that are unconditional on a specific model [76,77].

To quantify the magnitude of individual behavioural consistency in each foraging metric, variance component analyses were conducted using the models containing the parameters defined as influential after model averaging. The *ape* package [77] was used to calculate the variance, standard deviation and proportion of total variance occurring at the individual level, as well as the residual variation. The variance explained by the individual is considered an estimate of the individual specialization within a population [15,19].

To investigate the factors influencing individual variation, a second set of models using another measure of consistency, the coefficients of variation of the foraging metrics (standard deviation in the case of the bearing as it is a circular variable), calculated per deployment and used as response variable, was then developed [78]. The same set of explanatory variables and modelling approach described above were used.

As multiple logger deployments were performed on most individuals (mean \pm s.e.: 1.9 ± 0.1 deployments per bird), the full dataset allowed comparisons at different timescales to be made: trip-to-trip (T-to-T, data from consecutive trips obtained within the same deployment), breeding stage-to-breeding stage (S-to-S, data obtained from different breeding stages within the same year), and year-to-year (Y-to-Y, data obtained from the same breeding stage in two different years). This partition enabled the assessment of the timescales over which individual behavioural consistency is maintained. The full dataset was then partitioned to match each timescale tested, using in each case three or more foraging trips per deployment.

For the T-to-T comparison, the dataset was also subdivided into colonies (PD and PE) to quantify the individual consistency level at each site. The PD data were analysed in its entirety, reflecting the relatively uniform foraging habitat used by these individuals, whereas the PE data were split according to the predominant habitat individuals foraged in. Individuals that spent greater than 70% of trips during a deployment in PPB or Bass Strait were classified as PE-inshore and PE-pelagic, respectively, while individuals that did not were defined as PE-mixed. This partition allowed a comparison of individual behavioural consistency associated with different habitat selection. Models from these further separations were made using the same set of response and explanatory variables and followed all considerations described previously. Unless otherwise stated, results are presented as mean \pm s.e.

3. Results

3.1. Factors influencing foraging behaviour

The GPS and accelerometry data loggers were deployed on 142 breeding birds (260 deployments) from which 3–50 foraging trips were obtained (18.1 ± 0.9 trips per individual). From the GPS data loggers, a total of 2493 foraging trips were recorded but, due to battery life restrictions, accelerometry data were recorded for only 1284 trips. Consequently, the sample size used to obtain the different foraging metrics varied depending on the device used. A summary of the calculated foraging metrics is presented in table 1.

The top-ranked statistical models explaining the factors influencing the foraging behaviour of Australasian gannets were determined using model averaging, as the combined weight of the top set of models was low ($\omega_i < 0.9$, electronic supplementary material, tables S1 and S2). After model averaging, the best explanatory variables for maximum distances from the colony were colony, breeding stage, year and sex. Namely, individuals from PD, individuals during incubation, individuals during 2015/16 and females reached greater distances from the colony. For bearing and tortuosity index, colony, breeding stage and year were the most influential variables. Specifically, individuals from PD, individuals during late chick-rearing and individuals during 2015/16 showed higher bearings and tortuosity indices. Lastly, mean VeDBA and number of dives per foraging trip were both

Table 1. Means \pm s.e. of the foraging trip parameters collected from instrumented breeding Australasian gannets (*Morus serrator*) during two years 2014/15 and 2015/16 at the Point Danger (PD) and Pope's Eye (PE) colonies in Victoria, Australia. Instrumentation in birds lasted 10–12 consecutive days. Data represent the dataset acquired summarized by sex, breeding stages (incubation = INC, early chick-rearing = ECR, late chick-rearing = LCR), year and colony-habitat. Samples sizes of the metrics estimated from the GPS (maximum distances from the colony, bearing and tortuosity index; n_1) and accelerometer data loggers (mean vectorial dynamic body acceleration and number of dives, n_2) are shown.

		n_1	distances from colony (km)	bearing (°)	tortuosity index	n_2	mean VeDBA (g)	Number of dives
sex	♂	1424	60.41 \pm 1.76	192.5 \pm 2.0	0.29 \pm 0.01	666	0.63 \pm 0.21	345.7 \pm 11.8
	♀	1056	77.22 \pm 1.98	201.4 \pm 2.0	0.30 \pm 0.01	618	0.57 \pm 0.17	390.6 \pm 12.8
breeding stage	INC	452	95.47 \pm 4.08	186.2 \pm 4.1	0.26 \pm 0.01	205	0.54 \pm 0.16	584.0 \pm 28.4
	ECR	1143	65.58 \pm 1.78	193.9 \pm 2.0	0.30 \pm 0.01	619	0.61 \pm 0.19	313.6 \pm 9.5
year	LCR	885	55.88 \pm 1.90	204.6 \pm 2.3	0.30 \pm 0.01	460	0.61 \pm 0.20	342.8 \pm 14.6
	2014	1224	64.18 \pm 2.09	187.8 \pm 2.2	0.27 \pm 0.01	390	0.62 \pm 0.21	398.1 \pm 19.4
colony-habitat	2015	1256	70.86 \pm 1.64	204.8 \pm 1.9	0.31 \pm 0.01	894	0.59 \pm 0.19	353.7 \pm 9.2
	PD–pelagic	1162	93.31 \pm 2.32	234.1 \pm 1.9	0.32 \pm 0.01	686	0.59 \pm 0.18	369.8 \pm 12.5
PE–mixed	PE–pelagic	699	56.77 \pm 1.61	181.0 \pm 1.6	0.29 \pm 0.01	375	0.58 \pm 0.17	369.1 \pm 14.8
	PE–inshore	264	20.63 \pm 1.68	131.1 \pm 3.6	0.22 \pm 0.01	78	0.74 \pm 0.29	273.4 \pm 29.6
	PE–mixed	355	39.46 \pm 1.75	150.8 \pm 4.1	0.26 \pm 0.01	145	0.62 \pm 0.22	401.4 \pm 26.2

influenced the most by breeding stage and sex. Individuals during early chick-rearing and males had higher VeDBA values, and individuals during incubation and females displayed a higher number of dives (table 2).

3.2. Influence of timescales and habitats on foraging behaviour consistency

Variance component analyses were performed to determine the proportion of variance explained by the individual for each of five foraging metrics. As 86% ($n=226$) of the deployments were conducted simultaneously on breeding partners from the same nests, nest identity was tested during the modelling as a random component. The addition of this random component did not significantly improve models in all cases ($p>0.05$ in all cases) and was, therefore, unnecessary. Conversely, the individual random component was significant ($p<0.05$) in all but two of the sets of models developed. For short-term comparisons (T-to-T comparisons), the variance associated with the individual component ranged from low to moderate (11.1–27.1%) and overall decreased as the timescale comparison increased to mid-term (S-to-S: 9.5 to 22.9%) and long-term (Y-to-Y: 0.0 to 28.6%, table 3).

For PE, 127 of the deployments were categorized according to the predominant habitat in which each bird foraged, with 70 classified as PE-pelagic, 33 as PE-mixed and 24 as PE-inshore (see examples in figure 2). The proportion of females for each classification was 69%, 27% and 4%, respectively, with males being more abundant at PE-inshore and PE-mixed. The proportion of variance explained by the individual between habitats overall ranged from low to moderate values (3.2–50.4%). Consistency values (maximum distances from the colony, tortuosity indices, number of dives) were higher for both pelagic habitats (PD-pelagic, PE-pelagic) compared to the mixed strategy (PE-mixed). Except for one variable (VeDBA), consistency values were higher for the inshore strategy (PE-inshore) (table 4).

Using the coefficients of variation (or the standard deviations) of foraging metrics within deployments as a measure of individual consistency, the factors influencing individual variation were investigated. The five foraging metrics examined (maximum distance from the colony, bearing from the colony to the most distal point, tortuosity index, mean VeDBA and total number of dives) required model averaging due to the lack of a single best model from the candidate set of models. After model averaging, the most influential factors on individual variation identified were year, colony and breeding stage for the T-to-T comparison level, with individuals sampled during year 2014/15, at PD and in late chick-rearing stage having the highest consistency (electronic supplementary material, table S3).

4. Discussion

Determining the factors influencing foraging behaviour in marine predators and the persistence of behavioural consistency through time is crucial to understand habitat selection and how populations can adapt to fast environmental changes [78]. In the present study, spatio-temporal factors (colony, stage, year) influenced the foraging behaviour metrics obtained in breeding Australasian gannets the most, while individual characteristics (BCI, BSI and WLI) did not, with the exception of sex. The proportion of variation explained by the individual showed higher values over shorter (T-to-T) than longer (S-to-S and Y-to-Y) timescales, consistent with previous studies investigating the persistence of behavioural consistency in seabirds [78,79] and the repeatability of behaviours in several taxa [80]. Individual consistency in foraging behaviour was found to be higher in inshore compared to pelagic habitats and mixed use of both habitats, supporting suggestions that consistency is favoured in stable environments with predictable resources [79,81]. Lastly, measures of individual variation (CVs and SDs) were explained by spatio-temporal factors rather than individual characteristics; this supports the idea that consistency is linked to the strategies displayed by individuals depending on habitat selection and prey availability dictated by environmental variables rather than intrinsic factors.

4.1. Factors influencing foraging behaviour

The use of metrics in foraging ecology research to describe the behaviour and estimate energetic expenditure of animals, particularly marine birds and mammals, and to provide an indication of the foraging strategies and habitats used, is common practice [82]. Colony, year and breeding stage were the most influential factors on the foraging metrics analysed in the present study (i.e. maximum distance from the colony, bearing, tortuosity index, mean VeDBA and number of dives), followed by sex. Indices of body condition and body size (BCI, BSI, WLI) did not influence these metrics.

Table 2. Most parsimonious models after model averaging, and their corresponding estimated regression parameters, for five foraging metrics obtained from instrumented breeding Australasian gannets (*Morus serrator*).

foraging metric	model fixed effects	fixed effect	estimate	s.e.	d.f.	t-value	p-value
distance from colony (km)	colony + stage + year + sex	(intercept)	4.25	0.09	2340	47.58	<0.0001
		colony (PE)	-0.79	0.09	134	-8.65	<0.0001
		stage (INC)	0.51	0.06	2340	8.18	<0.0001
		stage (LCR)	-0.21	0.06	2340	-3.48	0.0005
		sex (male)	-0.32	0.09	134	-3.54	0.0005
		year (2015)	0.13	0.04	2340	3.08	0.002
bearing (°)	colony + stage + year	(intercept)	226.72	4.71	2353	48.15	<0.0001
		colony (PE)	-71.88	6.22	135	-11.55	<0.0001
		stage (INC)	-16.91	3.98	2353	-4.24	<0.0001
		stage (LCR)	11.73	3.81	2353	3.07	0.002
		year (2015)	14.82	2.97	2353	4.97	<0.0001
		(intercept)	0.31	0.005	2353	66.70	<0.0001
tortuosity index	colony + stage + year	colony (PE)	-0.05	0.006	135	-8.18	<0.0001
		stage (INC)	-0.05	0.005	2353	-10.07	<0.0001
		stage (LCR)	0.01	0.005	2353	-0.55	0.58
		year (2015)	0.04	0.004	2353	11.20	<0.0001
		(intercept)	0.83	0.01	1175	140.90	<0.0001
		stage (INC)	-0.04	0.01	1175	-5.55	<0.0001
mean VeDBA	stage + sex	stage (LCR)	-0.01	0.01	1175	-0.07	0.95
		sex (male)	0.03	0.01	108	3.70	0.0003
		(intercept)	6.62	0.14	1127	47.76	<0.0001
		stage (INC)	1.48	0.16	1127	9.12	<0.0001
		stage (LCR)	0.19	0.14	1127	1.40	0.16
		sex (male)	-0.36	0.17	106	-2.04	0.04

Table 3. Variance component analysis of instrumented breeding Australasian gannets (*Morus serrator*). Short- (trip-to-trip), medium- (stage-to-stage) and long-term (year-to-year) comparisons are shown. Sample sizes (number of trips/number of individuals) are presented for each final model. The significant fixed components of the models for which the coefficients of variation were used as response variable are shown as the factors influencing the individual variation in each case.

foraging trip parameter	timescale	σ^2	Σ	σ^2 (%)	n	influences on individual variation
distances from colony (km)*	T-to-T	0.21	0.46	27.05	2480/137	colony, year
	S-to-S	0.14	0.37	13.00	1166/56	colony, breeding stage
	Y-to-Y	0.166	0.40	16.64	1069/53	sex
bearing (°)	T-to-T	1090.48	33.02	26.32	2490/137	breeding stage, year
	S-to-S	944.32	30.72	22.90	1184/57	breeding stage
	Y-to-Y	1195.88	34.58	28.63	1069/53	none
tortuosity index	T-to-T	0.0006	0.026	11.07	2490/137	colony, sex, year, WLI
	S-to-S	0.0005	0.024	9.45	1184/57	colony, BSI, sex
	Y-to-Y	0.0005	0.023	8.70	1069/53	WLI, year, sex
mean VeDBA (g)*	T-to-T	0.001	0.005	16.91	1237/108	stage, sex
	S-to-S	0.001	0.005	14.66	641/51	none
	Y-to-Y	—	—	0.00	190/15	NA
number of dives*	T-to-T	0.57	0.75	18.89	1237/108	colony, breeding stage, year
	S-to-S	0.55	0.74	16.25	641/51	none
	Y-to-Y	—	—	0.00	190/15	NA

*Cube-root-transformed variables.

Geographical variation has previously been reported in the foraging behaviour of gannets [51,52,83,84] and other marine predators, reflecting spatial differences in resource availability or habitat accessibility [32,85,86]. The results of the present study are consistent with these findings and reveal the substantial differences in oceanographic regimes and habitats available to individuals from the PD and PE gannet colonies [49,52]. In particular, the individuals from PD, which forage within the Bonney Upwelling system, had longer foraging trips and higher tortuosity index than individuals sampled at PE, consistent with previous findings [52].

Year of sampling was found to influence the foraging behaviour of breeding Australasian gannets with individuals travelling less, having a lower tortuosity index, higher energy expenditure rate and diving more often during the 2014/15 compared to the 2015/16 breeding season. Breeding success (proportion of chicks fledged) was lower in 2014/15 (25% versus 50% at PE, and 48% versus 79% at PD, respectively; Rodríguez-Malagón 2014–2016, unpublished data). This suggests both sites experienced similar environmental variation influencing both foraging behaviour and reproductive success in a similar way. Previous studies at PE have reported an increased foraging effort in years of low local marine productivity [87], and inter-annual variation in foraging behaviour in response to environmental perturbations have been observed in other gannet species [83,88,89]. Indeed, primary productivity (as measured by chlorophyll-a concentration) was substantially higher in 2014/15 than in 2015/16 [90], coinciding with a strong El Niño-Southern Oscillation event with sea surface temperatures above average (bom.gov.au) in the later year.

Differences in foraging metrics were also evident between the different stages of the breeding season. Individuals conducted longer foraging trips, had a higher tortuosity index, lower energy consumption rate and dived more during incubation compared to the later breeding stages. Similar observations have been made in Australasian gannets [40,87] and other seabirds, and are thought to reflect a shift from self-feeding during incubation behaviour to chick-provisioning [32]. However, other studies have related changes in foraging behaviour between incubation and chick-rearing to be in response to temporal variation in prey availability due to environmental changes around colonies throughout the breeding period [91–93].

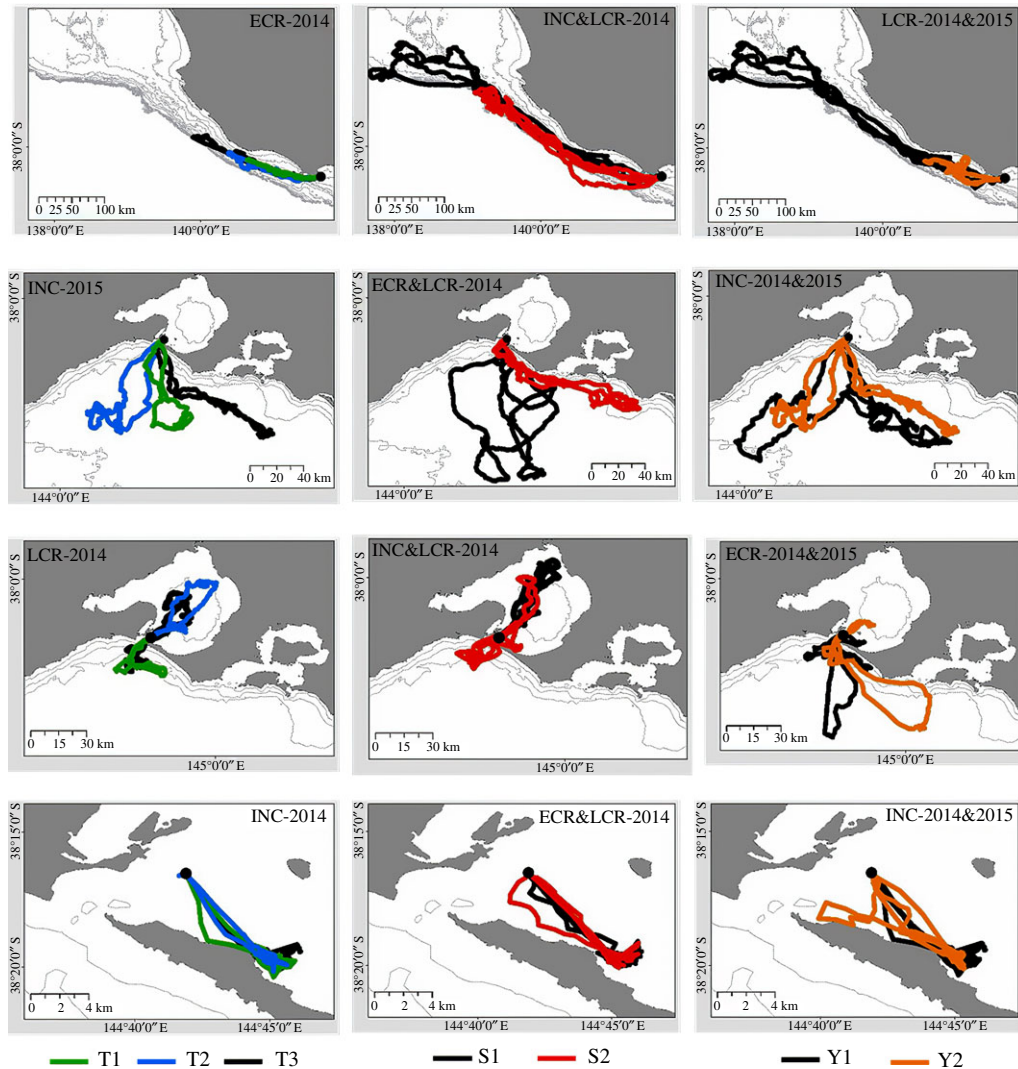


Figure 2. Examples of the time-scale comparisons investigated: T-to-T (trip-to-trip, left column); S-to-S (stage-to-stage, middle column); and Y-to-Y (year-to-year, right column). Each row represents an instrumented bird: first row, male from Point Danger-pelagic; second row, female from Pope's Eye-pelagic; third row, male from Pope's Eye-mixed; and fourth row, male from Pope's Eye-inshore. Breeding stages include incubation (INC), early chick-rearing (ECR) and late chick-rearing (LCR).

Maximum distance from the colony and the number of dives were shown to be influenced by sex, with females exhibiting higher values than males. Previous research at the two study colonies has shown sex differences in core foraging areas with only 4.2% and 18.4% of overlap at PD and PE, respectively [52]. Australasian gannets display reverse sexual dimorphism, with females being significantly heavier and larger than males [48]. This is consistent with observations in other Sulidae species in which males forage closer inshore than females, the larger sex [1,94,95]. In species with sexual size dimorphism, trophic or spatial segregation can function to reduce intra-specific competition, particularly during periods of intense resource competition [32,89]. Despite the greater foraging range and higher dive rate, females in the present study had lower mean VeDBA. This suggests females may be more efficient in some aspects of their foraging behaviour. Indeed, females from the study colonies have been previously been reported to spend a greater proportion of their foraging trips in gliding rather than flapping flight [52,87].

4.2. Influence of timescales and habitats on foraging behaviour consistency

The level of behavioural consistency displayed by individuals is thought to be related to the foraging strategy they adopt and influence how adaptable they can be when faced with rapid environmental changes [78,96]. Individuals in the present study displayed overall low to moderate levels of

Table 4. Variance component analysis of instrumented Australasian gannets (*Morus serrator*). Short time-scale comparison (trip-to-trip) results are shown of the models split by colony and habitat. Sample sizes (number of trips/number of individuals) are presented for each final model. The significant fixed components of the models for which the coefficients of variation were used as response variable are shown as the factors influencing the individual variation in each case.

foraging trip parameter				
colony-habitat	σ^2	Σ	σ^2 (%)	<i>n</i>
distances from colony (km)*				
PD-pelagic	0.24	0.49	15.93	1170/76
PE-pelagic	0.019	0.13	3.17	704/41
PE-mixed	0.227	0.47	20.22	355/25
PE-inshore	0.28	0.53	50.37	264/16
bearing (°)				
PD-pelagic	1171.45	34.22	26.32	1170/76
PE-pelagic	443.88	21.06	23.48	704/41
PE-mixed	1032.96	32.13	14.74	355/25
PE-inshore	2200.57	46.91	49.18	264/16
tortuosity index				
PD-pelagic	0.0005	0.02	8.92	1170/76
PE-pelagic	0.0004	0.02	8.53	704/41
PE-mixed	0.0015	0.038	10.84	355/25
PE-inshore	0.0010	0.031	12.21	264/16
mean VeDBA (g)*				
PD-pelagic	0.001	0.03	14.81	668/54
PE-pelagic	0.001	0.03	15.48	353/36
PE-mixed	0.001	0.02	9.73	149/18
PE-inshore	0.01	0.04	12.56	76/10
number of dives*				
PD-pelagic	0.24	0.49	8.00	668/54
PE-pelagic	0.49	0.70	20.17	353/36
PE-mixed	0.98	0.99	26.03	149/18
PE-inshore	0.96	0.98	41.62	76/10

*Cube-root-transformed variables.

behavioural consistency in foraging metrics. As time between sampling increased, behavioural consistency decreased.

Over the short-term (T-to-T), breeding Australasian gannets in the present study displayed moderate levels of consistency, specifically in distances from the colony and bearing to most distal point, which suggest some degree of foraging side fidelity during consecutive trips, and potentially the exploitation of the same resource patches, similar to what has been found in northern gannets [97] and other seabirds [32,79,96]. By contrast, tortuosity index, mean VeDBA and number of dives were shown to be less consistent, suggesting that while individuals tended to revisit patches, they could adapt to current local prey availability and environmental conditions, rather than be limited by individual morphology or foraging/diving abilities.

Consistent with other studies in marine top-order predators [78,79], behavioural consistency in the present study decreased over time; it was higher for consecutive trips, compared to between breeding stages, and between years. Behavioural consistency in foraging implies individuals learn, remember and select specific resources and foraging strategies [98]. It requires predictability in the abundance and location of the exploited resources so that the strategies can be maintained in the population [36], which is less likely to be maintained over time because of environmental variability. Our results

support the theory that behavioural consistency can only persist as long as stability in environmental conditions prevails [99]. Importantly, as seabird foraging conditions are highly susceptible to fluctuations in the environment [20,100,101], it may be advantageous for individuals to maintain a certain level of behavioural plasticity to respond to such change [97]; behavioural consistency can lead marine predators to encounter ecological traps in degraded environments, and limit the adaptability of individuals to environmental changes [97,102,103].

In the present study, differences in consistency were seen when breeding Australasian gannets selected different foraging habitats. Birds which foraged in pelagic environments at both colonies (PD-pelagic and PE-pelagic) were less consistent than birds which used both pelagic and inshore environments (PE-mixed), which in turn were less consistent than foraging in inshore environments (PE-inshore). Individual consistency is thought to be promoted in stable environments [81,104,105]. Birds in the PE-inshore category foraged in PPB, a shallow environment with an important coverage of seagrasses and sandy bottoms that represent important habitats for marine invertebrates and fish in southeastern Australia [106]. Such benthic environments are indeed considered refuges for fish and marine invertebrate communities, as they provide predictable stable habitats and nutrients [96,107]. Thus, they provide predictable resources for marine predators, but also bathymetric features that can be used as cues for resource availability and aids for navigation which can be memorized [32,49]. This theory is supported by a high level of behavioural consistency displayed by other benthic foraging seabirds [79,108,109]. By contrast, the less consistent individuals in the present study foraged in pelagic habitats (PD-pelagic and PE-pelagic) and were likely to exploit schooling fish [49], a temporally and spatially variable prey resource, influenced by various oceanographic processes subject to intra- and inter-annual variations affecting prey species in the region [110].

The differences in consistency observed between individuals foraging in inshore and pelagic environments is unlikely to be due to the geometry, size or complexity of the available habitat. Despite the narrowness of the northwest/southeast axis of continental shelf habitat frequented by PD-pelagic animals, these individuals displayed less consistency in bearing than those foraging in the more circular PPB area. Similarly, despite the arc of available headings to potential foraging areas being similar for the PE-pelagic and PE-inshore birds, the later were more consistent in their bearings from the colony. In addition, while the area of Bass Strait used by PE-pelagic birds was approximately two to three times the area of PBB, PE-inshore birds represented less than 20% of the sampled population, such that the latter are likely to have had greater *per capita* available habitat yet displayed more consistency in their foraging behaviour. Furthermore, despite PBB representing a smaller absolute area, it is characterized by greater habitat diversity (seagrass beds, rock reefs, shallow sand-banks and deeper channels) compared to the open water habitat of Bass Strait.

Finally, some individuals from PE in the present study adopted a strategy of consistently foraging in both pelagic (Bass Strait) and inshore (PPB) habitats, either within the same or successive foraging trips, suggesting a degree of behavioural plasticity. Similar findings have been reported for gentoo penguins (*Pygoscelis papua*) in which some individuals switched between pelagic and benthic strategies on successive foraging trips [111]. While it is not known whether this mixed foraging strategy has specific benefits, it has been suggested that spatial and temporal environmental variation and resource competition can promote different adaptive responses in individuals, giving rise to different levels of plasticity [112].

5. Conclusion

In summary, the present study found foraging behaviour in Australasian gannets to be influenced primarily by colony, breeding stage and year, reflecting the spatial and temporal variation in resources around breeding colonies and, to a lesser degree, by sex. Overall, low to moderate levels of behavioural consistency were observed, decreasing with increasing timescales between sampling, but higher in inshore environments, where individuals displayed more benthic foraging strategies associated with more stable and predictable environments. These findings could have important implications for population dynamics as individuals may not be uniformly affected by environmental variability. Southeastern Australia is one of the fastest warming marine areas in the world and the anticipated oceanographic changes are likely to affect the distribution, abundance and diversity of prey species [41,42]. Inter-individual differences in foraging behaviour and behavioural plasticity in Australasian gannets, therefore, could affect how the population responds to changing environmental conditions [113]. Future studies should investigate the links between specific environmental conditions

and behavioural consistency further, and the benefits conferred by strategies adopted by individuals and breeding pairs on reproductive success.

Ethics. All procedures were conducted under the approval of the Deakin University Animal Ethics Committee (B20–2013), the Department of Environment, Land, Water and Planning (DELWP, Wildlife Research Permit No. 10006878, File No. FF383295) of the Victorian Government and the Australian Bird and Bat Banding Scheme (ABBBS, R Class Banding Licence No. 3168). All applicable institutional and/or national guidelines for the care and use of animals were followed.

Data accessibility. See the electronic supplementary material to obtain data used in this paper.

Authors' contributions. M.A.R.-M. and J.P.Y.A. conceived and designed the study. M.A.R.-M. collected the data. M.A.R.-M., E.C.M.C. and L.P.A. performed and contributed to the data analysis. M.A.R.-M. and J.P.Y.A. wrote the paper. All authors gave final approval for publication.

Competing interests. The authors of this manuscript declare no competing interests.

Funding. This research was partly funded by the Holsworth Wildlife Research Endowment and Birdlife Australia.

Acknowledgements. We thank the generous support from the Victorian Marine Science Consortium (especially to Roderick Watson, Elizabeth McGrath and Yvonne Gilbert), Parks Victoria and the Point Danger Committee of Management (Ewen Lovell and Phillip King) for facilitating the field activities required for this study. The assistance of numerous volunteers who participated in field activities is gratefully acknowledged, as is the advice on statistical analyses provided by Dr Peter Biro from Deakin University.

References

- Lewis S, Benvenuti S, Dall-Antonia L, Griffiths R, Money L, Sherratt TN, Wanless S, Hamer KC. 2002 Sex-specific foraging behaviour in a monomorphic seabird. *Proc. R. Soc. Lond. B* **269**, 1687–1693. (doi:10.1098/rspb.2002.2083)
- Pankiw T, Tarpay DR, Page RE. 2002 Genotype and rearing environment affect honeybee perception and foraging behaviour. *Anim. Behav.* **64**, 663–672. (doi:10.1006/anbe.2002.3096)
- McGraw WS, Vick AE, Daegling DJ. 2011 Sex and age differences in the diet and ingestive behaviors of sooty mangabeys (*Cercocebus atys*) in the Tai Forest, Ivory Coast. *Am. J. Phys. Anthropol.* **144**, 140–153. (doi:10.1002/ajpa.21402)
- Holmes WG. 1984 Predation risk and foraging behavior of the hoary marmot in Alaska. *Behav. Ecol. Sociobiol.* **15**, 293–301. (doi:10.1007/bf00292992)
- Staniland IJ, Reid K, Boyd IL. 2004 Comparing individual and spatial influences on foraging behaviour in Antarctic fur seals *Arctocephalus gazella*. *Mar. Ecol. Prog. Ser.* **275**, 263–274. (doi:10.3354/meps275263)
- Peat J, Goulson D. 2005 Effects of experience and weather on foraging rate and pollen versus nectar collection in the bumblebee, *Bombus terrestris*. *Behav. Ecol. Sociobiol.* **58**, 152–156. (doi:10.1007/s00265-005-0916-8)
- Soanes LM, Arnould JPY, Dodd SG, Milligan G, Green JA. 2014 Factors affecting the foraging behaviour of the European shag: implications for seabird tracking studies. *Mar. Biol.* **161**, 1335–1348. (doi:10.1007/s00227-014-2422-x)
- Lewis S, Phillips RA, Burthe SJ, Wanless S, Daunt F. 2015 Contrasting responses of male and female foraging effort to year-round wind conditions. *J. Anim. Ecol.* **84**, 1490–1496. (doi:10.1111/1365-2656.12419)
- Galef BG, Giraldeau L-A. 2001 Social influences on foraging in vertebrates: causal mechanisms and adaptive functions. *Anim. Behav.* **61**, 3–15. (doi:10.1006/anbe.2000.1557)
- Schoener TW. 1971 Theory of feeding strategies. *Annu. Rev. Ecol. Syst.* **2**, 369–404. (doi:10.1146/annurev.es.02.110171.002101)
- Cohen D. 1993 The equilibrium distribution of optimal search and sampling effort of foraging animals in patchy environments. In *Adaptation in stochastic environments* (eds J Yoshimura, CW Clark), pp. 173–191. Berlin, Germany: Springer.
- Reader SM. 2015 Causes of individual differences in animal exploration and search. *Top. Cogn. Sci.* **7**, 451–468. (doi:10.1111/tops.12148)
- Dall SR, Bell AM, Bolnick DI, Ratnieks FL. 2012 An evolutionary ecology of individual differences. *Ecol. Lett.* **15**, 1189–1198. (doi:10.1111/j.1461-248.2012.01846.x)
- Sargeant BL. 2007 Individual foraging specialization: niche width versus niche overlap. *Oikos* **116**, 1431–1437. (doi:10.1111/j.2007.0030-1299.15833.x)
- Bolnick DI, Svanback R, Fordyce JA, Yang LH, Davis JM, Hulsey CD, Forister ML. 2003 The ecology of individuals: incidence and implications of individual specialization. *Am. Nat.* **161**, 1–28. (doi:10.1086/343878)
- Araújo MS, Bolnick DI, Layman CA. 2011 The ecological causes of individual specialisation. *Ecol. Lett.* **14**, 948–958. (doi:10.1111/j.1461-0248.2011.01662.x)
- Provencher JF, Elliott KH, Gaston AJ, Braune BM. 2013 Networks of prey specialization in an Arctic monomorphic seabird. *J. Avian Biol.* **44**, 551–560. (doi:10.1111/j.1600-048X.2013.05717.x)
- Bolnick DI *et al.* 2011 Why intraspecific trait variation matters in community ecology. *Trends Ecol. Evol.* **26**, 183–192. (doi:10.1016/j.tree.2011.01.009)
- Dingemans NJ, Dochtermann NA. 2013 Quantifying individual variation in behaviour: mixed-effect modelling approaches. *J. Anim. Ecol.* **82**, 39–54. (doi:10.1111/1365-2656.12013)
- Ballance LT, Pitman RL, Fiedler PC. 2006 Oceanographic influences on seabirds and cetaceans of the eastern tropical Pacific: a review. *Prog. Oceanogr.* **69**, 360–390. (doi:10.1016/j.pocean.2006.03.013)
- Macdonald AM, Wunsch C. 1996 An estimate of global ocean circulation and heat fluxes. *Nature* **382**, 436–439. (doi:10.1038/382436a0)
- Butler A, Althaus F, Furlani D, Ridgway K. 2002 *Assessment of the conservation values of the Bonney upwelling area: a component of the Commonwealth Marine Conservation Assessment Program 2002–2004*. Report to Environment Australia. Clayton, Australia: CSIRO Marine Research.
- Yurkowski DJ, Ferguson S, Choy ES, Loseto LL, Brown TM, Muir DCG, Semeniuk CAD, Fisk AT. 2016 Latitudinal variation in ecological opportunity and intraspecific competition indicates differences in niche variability and diet specialization of Arctic marine predators. *Ecol. Evol.* **6**, 1666–1678. (doi:10.1002/ece3.1980)
- Paiva VH, Geraldes P, Ramirez I, Meirinho A, Garthe S, Ramos JA. 2010 Foraging plasticity in a pelagic seabird species along a marine productivity gradient. *Mar. Ecol. Prog. Ser.* **398**, 259–274. (doi:10.3354/meps08319)
- Cummings M, Mollaghan D. 2006 Repeatability and consistency of female preference behaviours in a northern swordtail, *Xiphophorus nigrensis*. *Anim. Behav.* **72**, 217–224. (doi:10.1016/j.anbehav.2006.01.009)
- Matich P, Heithaus MR, Layman CA. 2011 Contrasting patterns of individual specialization and trophic coupling in two marine apex predators. *J. Anim. Ecol.* **80**, 294–305. (doi:10.1111/j.1365-2656.2010.01753.x)
- Estes J, Riedman M, Staedler M, Tinker M, Lyon B. 2003 Individual variation in prey selection by sea otters: patterns, causes and implications. *J. Anim. Ecol.* **72**, 144–155. (doi:10.1046/j.1365-2656.2003.00690.x)

28. Baylis A, Orben R, Arnould J, Peters K, Knox T, Costa D, Staniland I. 2015 Diving deeper into individual foraging specializations of a large marine predator, the southern sea lion. *Oecologia* **179**, 1053–1065. (doi:10.1007/s00442-015-3421-4)
29. Shealer DA. 2001 Foraging behavior and food of seabirds. In *Biology of marine birds* (eds E Schreiber, J Burger), pp. 137–177. Boca Raton, FL: CRC Press.
30. Smith JL, Muler CPH, Ellis JC. 2011 Seabirds as ecosystems engineers: Nutrient inputs and physical disturbance. In *Seabird islands: ecology, invasion and restoration* (eds CPH Mulder, WB Anderson, DR Towns, PJ Bellingham), pp. 27–55. Oxford, UK: Oxford University Press.
31. Lewis S, Sherratt T, Hamer K, Wanless S. 2001 Evidence of intra-specific competition for food in a pelagic seabird. *Nature* **412**, 816–819. (doi:10.1038/35090566)
32. Phillips RA, Lewis S, González-Solís J, Daunt F. 2017 Causes and consequences of individual variability and specialization in foraging and migration strategies of seabirds. *Mar. Ecol. Prog. Ser.* **578**, 117–150. (doi:10.3354/meps12217)
33. Ceia F, Ramos J. 2015 Individual specialization in the foraging and feeding strategies of seabirds: a review. *Mar. Biol.* **162**, 1923–1938. (doi:10.1007/s00227-015-2735-4)
34. Hamer K, Humphreys E, Garthe S, Hennicke J, Peters G, Grémillet D, Phillips R, Harris M, Wanless S. 2007 Annual variation in diets, feeding locations and foraging behaviour of gannets in the North Sea: flexibility, consistency and constraint. *Mar. Ecol. Prog. Ser.* **338**, 5–305. (doi:10.3354/meps338295)
35. Woo KJ, Elliott KH, Davidson M, Gaston AJ, Davoren GK. 2008 Individual specialization in diet by a generalist marine predator reflects specialization in foraging behaviour. *J. Anim. Ecol.* **77**, 1082–1091. (doi:10.1111/j.1365-2656.2008.01429.x)
36. Devictor V, Clavel J, Julliard R, Lavergne S, Mouillot D, Thuiller W, Venail P, Villéger S, Mouquet N. 2010 Defining and measuring ecological specialization. *J. Appl. Ecol.* **47**, 15–25. (doi:10.1111/j.1365-2664.2009.01744.x)
37. Colles A, Liow LH, Prinzing A. 2009 Are specialists at risk under environmental change? Neoecological, paleoecological and phylogenetic approaches. *Ecol. Lett.* **12**, 849–863. (doi:10.1111/j.1461-0248.2009.01336.x)
38. Bunce A, Norman FI, Brothers N, Gales R. 2002 Long-term trends in the Australasian gannet (*Morus serrator*) population in Australia: the effect of climate change and commercial fisheries. *Mar. Biol.* **141**, 263–269. (doi:10.1007/s00227-002-0838-1)
39. Srinivasan M, Dassis M, Benn E, Stockin KA, Martinez E, Machovsky-Capuska GE. 2015 Using non-systematic surveys to investigate effects of regional climate variability on Australasian gannets in the Hauraki Gulf, New Zealand. *J. Sea Res.* **99**, 74–82. (doi:10.1016/j.seares.2015.02.004)
40. Bunce A. 2001 Prey consumption of Australasian gannets (*Morus serrator*) breeding in Port Phillip Bay, southeast Australia, and potential overlap with commercial fisheries. *ICES J. Mar. Sci.* **58**, 904–915. (doi:10.1006/jmsc.2001.1083)
41. Ridgway KR. 2007 Long-term trend and decadal variability of the southward penetration of the East Australian Current. *Geophys. Res. Lett.* **34**, L13613. (doi:10.1029/2007GL030393)
42. Lough JM, Hobday AJ. 2011 Observed climate change in Australian marine and freshwater environments. *Mar. Freshwater Res.* **62**, 984–999. (doi:10.1071/MF10272)
43. Hobday AJ, Pecl GT. 2014 Identification of global marine hotspots: sentinels for change and vanguards for adaptation action. *Rev. Fish Biol. Fish.* **24**, 415–425. (doi:10.1007/s11160-013-9326-6)
44. Pecl GT, Hobday AJ, Frusher S, Sauer WHH, Bates AE. 2014 Ocean warming hotspots provide early warning laboratories for climate change impacts. *Rev. Fish Biol. Fish.* **24**, 409–413. (doi:10.1007/s11160-014-9355-9)
45. Johnson CR et al. 2011 Climate change cascades: shifts in oceanography, species' ranges and subtidal marine community dynamics in eastern Tasmania. *J. Exp. Mar. Biol. Ecol.* **400**, 17–32. (doi:10.1016/j.jembe.2011.02.032)
46. Schuckard R, Melville D, Cook W, Machovsky-Capuska G. 2012 Diet of the Australasian gannet (*Morus serrator*) at Farewell Spit, New Zealand. *Notornis* **59**, 66–70.
47. Machovsky-Capuska GE et al. 2016 Sex-specific macronutrient foraging strategies in a highly successful marine predator: the Australasian gannet. *Mar. Biol.* **163**, 75. (doi:10.1007/s00227-016-2841-y)
48. Angel LP, Wells MR, Rodríguez-Malagón MA, Tew E, Speakman JR, Arnould JPY. 2015 Sexual size dimorphism and body condition in the Australasian gannet. *PLoS ONE* **10**, e0142653. (doi:10.1371/journal.pone.0142653)
49. Wells MR, Angel LP, Arnould JPY. 2016 Habitat-specific foraging strategies in Australasian gannets. *Biol. Open*. bio018085. (doi:10.1242/bio.018085)
50. Besel D, Hauber ME, Hunter C, Ward-Smith T, Raubenheimer D, Millar CD, Ismar SMH. 2018 Multifactorial roles of interannual variability, season, and sex for foraging patterns in a sexually size monomorphic seabird, the Australasian gannet (*Morus serrator*). *Mar. Biol.* **165**, 72. (doi:10.1007/s00227-018-3332-0)
51. Machovsky-Capuska GE et al. 2014 Foraging behaviour and habitat use of chick-rearing Australasian gannets in New Zealand. *J. Ornithol.* **155**, 379–387. (doi:10.1007/s10336-013-1018-4)
52. Angel LP, Berlincourt M, Arnould JPY. 2016 Pronounced inter-colony variation in the foraging ecology of Australasian gannets: influence of habitat differences. *Mar. Ecol. Prog. Ser.* **556**, 261–272. (doi:10.3354/meps11845)
53. Lewis R. 1981 Seasonal upwelling along the southeastern coastline of South Australia. *Mar. Freshwater Res.* **32**, 843–854. (doi:10.1071/MF9810843)
54. Walker SJ. 1999 Coupled hydrodynamic and transport models of Port Phillip Bay, a semi-enclosed bay in southeastern Australia. *Mar. Freshwater Res.* **50**, 469–481. (doi:10.1071/MF98071)
55. Wingham E. 1982 Breeding biology of the Australasian gannet *Morus serrator* (Gray) at Motu Karamarama, Hauraki Gulf, New Zealand. II. Breeding success and chick growth. *Emu* **84**, 211–224. (doi:10.1071/MU9840211)
56. Garthe S, Guse N, Montevecchi WA, Rail JF, Groggier F. 2014 The daily catch: flight altitude and diving behavior of northern gannets feeding on Atlantic mackerel. *J. Sea Res.* **85**, 456–462. (doi:10.1016/j.seares.2013.07.020)
57. R Core Team. 2017 *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. See <https://www.R-project.org/>.
58. McConnell B, Chambers C, Fedak M. 1992 Foraging ecology of southern elephant seals in relation to the bathymetry and productivity of the Southern Ocean. *Antarct. Sci.* **4**, 393–398. (doi:10.1017/S0954102092000580)
59. Sumner MD, Sebastian L, Anthony F. 2016 trip: Tools for the analysis of animal track data. R package version 1.5.0.
60. Calenge C. 2006 The package 'adehabitat' for the R software: a tool for the analysis of space and habitat use by animals. *Ecol. Modell.* **197**, 516–519. (doi:10.1016/j.ecolmodel.2006.03.017)
61. Benhamou S. 2004 How to reliably estimate the tortuosity of an animal's path: straightness, sinuosity, or fractal dimension? *J. Theor. Biol.* **229**, 209–220. (doi:10.1016/j.jtbi.2004.03.016)
62. Calenge C, Dray S, Royer-Carenzi M. 2009 The concept of animals' trajectories from a data analysis perspective. *Ecol. Inform.* **4**, 34–41. (doi:10.1016/j.ecoinf.2008.10.002)
63. Warwick-Evans V, Atkinson PW, Gauvain RD, Robinson LA, Arnould JPY, Green JA. 2015 Time-in-area represents foraging activity in a wide-ranging pelagic forager. *Mar. Ecol. Prog. Ser.* **527**, 233–246. (doi:10.3354/meps11262)
64. Ropert-Coudert Y, Daunt F, Kato A, Ryan PG, Lewis S, Kobayashi K, Mori Y, Grémillet D, Wanless S. 2009 Underwater wingbeats extend depth and duration of plunge dives in northern gannets *Morus bassanus*. *J. Avian Biol.* **40**, 380–387. (doi:10.1111/j.1600-048X.2008.04592.x)
65. Ropert-Coudert Y, Grémillet D, Kato A, Ryan PG, Naito Y, Le Maho Y. 2004 A fine-scale time budget of Cape gannets provides insights into the foraging strategies of coastal seabirds. *Anim. Behav.* **67**, 985–992. (doi:10.1016/j.anbehav.2003.09.010)
66. Weimerskirch H, Le Corre M, Ropert-Coudert Y, Kato A, Marsac F. 2005 The three-dimensional flight of red-footed boobies: adaptations to foraging in a tropical environment? *Proc. R. Soc. B* **272**, 53–61. (doi:10.1098/rspb.2004.2918)
67. Sakamoto KQ, Sato K, Ishizuka M, Watanuki Y, Takahashi A, Daunt F, Wanless S. 2009 Can ethograms be automatically generated using body acceleration data from free-ranging birds? *PLoS ONE* **4**, e5379. (doi:10.1371/journal.pone.0005379)
68. Gleiss AC, Wilson RP, Shepard ELC. 2011 Making overall dynamic body acceleration work: on the theory of acceleration as a proxy for energy expenditure. *Methods Ecol. Evol.* **2**, 23–33. (doi:10.1111/j.2041-210X.2010.00057.x)
69. Qasem L, Cardew A, Wilson A, Griffiths I, Halsey LG, Shepard ELC, Gleiss AC, Wilson R. 2012 Tri-

- axial dynamic acceleration as a proxy for animal energy expenditure; should we be summing values or calculating the vector? *PLoS ONE* **7**, e31187. (doi:10.1371/journal.pone.0031187)
70. Angel LP. 2015 Factors influencing foraging behaviour in the Australasian gannet. Doctoral thesis, Deakin University, Melbourne, Australia.
71. Pinheiro J, Bates D, DebRoy S, Sarkar D, 2014 R Core Team. 2014 nlme: linear and nonlinear mixed effects models. R package version 3.1–117. See <http://CRAN.R-project.org/package=nlme>.
72. Cox NJ. 2011 Stata tip 96: cube roots. *Stata J.* **11**, 149–154. (doi:10.1177/1536867X1101100112)
73. Zuur AF, Ieno EN, Elphick CS. 2010 A protocol for data exploration to avoid common statistical problems. *Methods Ecol. Evol.* **1**, 3–14. (doi:10.1111/j.2041-210X.2009.00001.x)
74. Barton KJ. 2016 MuMIn: Multi-Model Inference. R package version 1.15.6, <https://CRAN.R-project.org/package=MuMIn>.
75. Burnham KP, Anderson DR. 2002 *Model selection and multimodel inference: a practical information-theoretic approach*. New York, NY: Springer Science & Business Media.
76. Symonds ME, Moussalli A. 2011 A brief guide to model selection, multimodel inference and model averaging in behavioural ecology using Akaike's information criterion. *Behav. Ecol. Sociobiol.* **65**, 13–21. (doi:10.1007/s00265-010-1037-6)
77. Paradis E, Claude J, Strimmer K. 2004 APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)
78. Comrasse ECM, Sutton GJ, Berlincourt M, Arnould JPY. 2017 Changing with the times: little penguins exhibit flexibility in foraging behaviour and low behavioural consistency. *Mar. Biol.* **164**, 169. (doi:10.1007/s00227-017-3193-y)
79. Harris S, Raya Rey A, Zavalaga C, Quintana F. 2014 Strong temporal consistency in the individual foraging behaviour of imperial shags *Phalacrocorax atriceps*. *Ibis* **156**, 523–533. (doi:10.1111/ibi.12159)
80. Bell AM, Hankson SJ, Laskowski KL. 2009 The repeatability of behaviour: a meta-analysis. *Anim. Behav.* **77**, 771–783. (doi:10.1016/j.anbehav.2008.12.022)
81. Wolf M, Weissing FJ. 2012 Animal personalities: consequences for ecology and evolution. *Trends Ecol. Evol.* **27**, 452–461. (doi:10.1016/j.tree.2012.05.001)
82. Carneiro APB, Bonnet-Lebrun AS, Manica A, Staniland IJ, Phillips RA. 2017 Methods for detecting and quantifying individual specialisation in movement and foraging strategies of marine predators. *Mar. Ecol. Prog. Ser.* **578**, 151–166. (doi:10.3354/meps12215)
83. Moseley C *et al.* 2012 Foraging ecology and ecophysiology of Cape gannets from colonies in contrasting feeding environments. *J. Exp. Mar. Biol. Ecol.* **422–423**, 29–38. (doi:10.1016/j.jembe.2012.04.002)
84. Pettex E, Lorentsen SH, Gremillet D, Gimenez O, Barrett RT, Pons JB, Le Bohec C, Bonadonna F. 2012 Multi-scale foraging variability in northern gannet (*Morus bassanus*) fuels potential foraging plasticity. *Mar. Biol.* **159**, 2743–2756. (doi:10.1007/s00227-012-2035-1)
85. Baylis AMM, Page B, Goldsworthy SD. 2008 Colony-specific foraging areas of lactating New Zealand fur seals. *Mar. Ecol. Prog. Ser.* **361**, 279–290. (doi:10.3354/meps07258)
86. Falk K, Benvenuti S, Dall'Antonia L, Gilchrist G, Kamp K. 2002 Foraging behaviour of thick-billed mures breeding in different sectors of the North Water polynya: an inter-colony comparison. *Mar. Ecol. Prog. Ser.* **231**, 293–302. (doi:10.3354/meps231293)
87. Angel LP, Barker S, Berlincourt M, Tew E, Warwick-Evans V, Arnould JP. 2015 Eating locally: Australasian gannets increase their foraging effort in a restricted range. *Biol. Open* **4**, 1298–1305. (doi:10.1242/bio.013250)
88. Kai ET, Benhamou S, van der Linden CD, Coetzee JC, Pichegru L, Ryan PG, Gremillet D. 2013 Are Cape gannets dependent upon fishery waste? A multi-scale analysis using seabird GPS-tracking, hydro-acoustic surveys of pelagic fish and vessel monitoring systems. *J. Appl. Ecol.* **50**, 659–670. (doi:10.1111/1365-2664.12086)
89. Cleasby IR, Wakefield ED, Bodey TW, Davies RD, Patrick SC, Newton J, Votier SC, Bearhop S, Hamer KC. 2015 Sexual segregation in a wide-ranging marine predator is a consequence of habitat selection. *Mar. Ecol. Prog. Ser.* **518**, 1–12. (doi:10.3354/meps11112)
90. Evans K, Bax N, Smith DC. 2017 *Australia state of the environment 2016: marine environment*. Independent report to the Australian Government Minister for the Environment and Energy, Australian Government Department of the Environment and Energy, Canberra.
91. Ito M, Minami H, Tanaka Y, Watanuki Y. 2009 Seasonal and inter-annual oceanographic changes induce diet switching in a piscivorous seabird. *Mar. Ecol. Prog. Ser.* **393**, 273–284. (doi:10.3354/meps08192)
92. Jakubas D, Wojczulanis-Jakubas K, Iliszko L, Darecki M, Stempniewicz L. 2014 Foraging strategy of the little auk *Alle alle* throughout breeding season – switch from unimodal to bimodal pattern. *J. Avian Biol.* **45**, 551–560. (doi:10.1111/jav.00303)
93. Shoji A *et al.* 2016 Foraging flexibility and search patterns are unlinked during breeding in a free-ranging seabird. *Mar. Biol.* **163**, 72. (doi:10.1007/s00227-016-2826-x)
94. Weimerskirch H, Le Corre M, Ropert-Coudert Y, Kato A, Marsac F. 2006 Sex-specific foraging behaviour in a seabird with reversed sexual dimorphism: the red-footed booby. *Oecologia* **146**, 681–691. (doi:10.1007/s00442-005-0226-x)
95. Stauss C *et al.* 2012 Sex-specific foraging behaviour in northern gannets *Morus bassanus*: incidence and implications. *Mar. Ecol. Prog. Ser.* **457**, 151–162. (doi:10.3354/meps09734)
96. Sampson J, Easton A, Singh M. 2014 Port Phillip bay. In *Estuaries of Australia in 2050 and beyond*. *Estuaries of the world* (ed. E Wolanski), pp. 49–68. Dordrecht, The Netherlands: Springer.
97. Wakefield ED, Cleasby IR, Bearhop S, Bodey TW, Davies RD, Miller PI, Newton J, Votier SC, Hamer KC. 2015 Long-term individual foraging site fidelity – why some gannets don't change their spots. *Ecology* **96**, 3058–3074. (doi:10.1890/14-1300.1)
98. Hamer K, Phillips R, Hill J, Wanless S, Wood A. 2001 Contrasting foraging strategies of gannets *Morus bassanus* at two North Atlantic colonies: foraging trip duration and foraging area fidelity. *Mar. Ecol. Prog. Ser.* **224**, 283–290. (doi:10.3354/meps224283)
99. Ceia FR *et al.* 2014 Annual and seasonal consistency in the feeding ecology of an opportunistic species, the yellow-legged gull *Larus michahellis*. *Mar. Ecol. Prog. Ser.* **497**, 273–284. (doi:10.3354/meps10586)
100. Schreiber EA, Burger J. 2001 *Biology of marine birds*. Boca Raton, FL: CRC Press.
101. Bost CA, Cotté C, Bailleul F, Chérel Y, Charrassin JB, Guinet C, Ainley DG, Weimerskirch H. 2009 The importance of oceanographic fronts to marine birds and mammals of the southern oceans. *J. Mar. Syst.* **78**, 363–376. (doi:10.1016/j.jmarsys.2008.11.022)
102. Sherley RB, Ludynia K, Dyer BM, Lamont T, Makhado AB, Roux JP, Scales KL, Underhill LG, Votier SC. 2017. Metapopulation tracking juvenile penguins reveals an ecosystem-wide ecological trap. *Curr. Biol.* **27**:S63–S68. (doi:10.1016/j.cub.2016.12.054)
103. McIntyre T, Bester MN, Bornemann H, Tosh CA, de Bruyn PJN. 2017 Slow to change? Individual fidelity to three-dimensional foraging habitats in Southern elephant seals *Mirounga leonina*. *Anim. Behav.* **127**:91–99. (doi:10.1016/j.anbehav.2017.03.006)
104. Weimerskirch H. 2007 Are seabirds foraging for unpredictable resources? *Deep Sea Res. Part II. Topical Stud. Oceanogr.* **54**, 211–223. (doi:10.1016/j.dsr2.2006.11.013)
105. Patrick SC, Weimerskirch H. 2014 Consistency pays: sex differences and fitness consequences of behavioural specialization in a wide-ranging seabird. *Biol. Lett.* **10**, 20140630. (doi:10.1098/rsbl.2014.0630)
106. Blandon A, zu Ermgassen PSE. 2014 Quantitative estimate of commercial fish enhancement by seagrass habitat in southern Australia. *Estuar. Coast. Shelf Sci.* **141**, 1–8. (doi:10.1016/j.ecss.2014.01.009)
107. Jenkins GP, May HMA, Wheatley MJ, Holloway MG. 1997 Comparison of fish assemblages associated with seagrass and adjacent unvegetated habitats of Port Phillip Bay and Corner Inlet, Victoria, Australia, with emphasis on commercial species. *Estuar. Coast. Shelf Sci.* **44**, 569–588. (doi:10.1006/ecss.1996.0131)
108. Comrasse ECM, Chérel Y, Arnould JPY, Hoskins AJ, Bost CA. 2017 Combined bio-logging and stable isotopes reveal individual specialisations in a benthic coastal seabird, the Kerguelen shag. *PLoS ONE* **12**, e0172278. (doi:10.1371/journal.pone.0172278)
109. Ratcliffe N, Takahashi A, O'Sullivan C, Adlard S, Trathan PN, Harris MP, Wanless S. 2013 The roles of sex, mass and individual specialisation in partitioning foraging-depth niches of a pursuit-diving predator. *PLoS ONE* **8**, e79107. (doi:10.1371/journal.pone.0079107)
110. Middleton JF, Bye JA. 2007 A review of the shelf-sole circulation along Australia's southern shelves: Cape Leeuwin to Portland. *Prog. Oceanogr.* **75**, 1–41. (doi:10.1016/j.pocean.2007.07.001)

111. Sandery PA, Kämpf J. 2007 Transport timescales for identifying seasonal variation in Bass Strait, southeastern Australia. *Estuar. Coast. Shelf Sci.* **74**, 684–696. (doi:10.1016/j.ecss.2007.05.011)
112. Camprasse EC, Chereil Y, Bustamante P, Arnould JP, Bost CA. 2017. Intra- and inter-individual variation in the foraging ecology of a generalist subantarctic seabird, the gentoo penguin. *Mar. Ecol. Prog. Ser.* **578**, 227–242 (doi:10.3354/meps12151)
113. Dingemans NJ, Wolf M. 2013 Between-individual differences in behavioural plasticity within populations: causes and consequences. *Anim. Behav.* **85**, 1031–1039. (doi:10.1016/j.anbehav.2012.12.032)