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Synthesis of adenosine analogues with indole moiety as human adenosine A₃ receptor ligands

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Adenosine is an endogenous modulator exerting its functions through the activation of four adenosine receptor (AR) subtypes, termed A₁, A_{2A}, A_{2B} and A₃, which belong to the G-protein-coupled receptor superfamily. The human A₃AR (hA₃AR) subtype is implicated in several cytoprotective functions. Therefore, hA₃AR modulators, and in particular agonists, are sought for their potential application as anti-inflammatory, anti-cancer and cardioprotective agents. Here, we prepared novel adenosine derivatives with indole moiety as hA₃AR ligands. According to the biological assay, we found that 2-substituents **11** were critical structural determinants for A₃AR ligands ($K_i = 111$ nM). The observed structure–affinity relationships of this class of ligands were also exhaustively rationalized using the molecular modelling approach. This allows the investigation on the binding mode of the potential compound in the ligand-binding pocket of the human A₃ receptor. The results demonstrated that **11** can interact with the ASN250, GLN167, PHE168 and VAL178 through hydrogen bonding, which are shown to be important for ligand–receptor interaction.

1. Introduction

Adenosine is an endogenous purine nucleoside that modulates many physiological processes, which is composed of a molecule of adenine attached to a ribose sugar molecule (ribofuranose) moiety

via a β - N_9 -glycosidic bond [1–3]. Adenosine is widely found in nature and plays an important role in biochemical processes, such as energy transfer as adenosine triphosphate (ATP) and adenosine diphosphate (ADP), as well as in signal transduction as cyclic adenosine monophosphate (cAMP). It is also a neuromodulator, believed to play a role in promoting sleep and suppressing arousal. Adenosine also plays a role in the regulation of blood flow to various organs through vasodilation [4–6]. Cellular signalling by adenosine occurs through four known adenosine receptor (AR) subtypes (A_1 , A_{2A} , A_{2B} and A_3) [7]. All AR subtypes are G-protein-coupled receptors. The four receptor subtypes are further classified based on their ability to either stimulate or inhibit adenylate cyclase activity. They have long been considered to be promising therapeutic targets in a wide range of conditions, ranging from cerebral diseases to cancer, including inflammatory and immunological disorders [8,9]. Adenosine contributes in a significant manner to the maintenance of tissue integrity by modulating the immune system. Encouraging results have emerged with AR ligands for the management of several physiological conditions in preclinical and clinical settings [7,10].

Among the four AR subtypes, the A_3 AR, probably the most studied AR subtype, is also ubiquitously expressed [11]. The distribution of A_3 AR is species-dependent, and in humans, this subtype is expressed in the lungs, liver, heart, kidneys and brain [12–14]. The widespread distribution in different cells and tissues of the A_3 AR suggests a potential involvement in various pathologies and the possible use as a selective pharmacological target [15]. This subtype of ARs is involved in a variety of important pathophysiological processes, ranging from modulation of cerebral and cardiac ischaemic damage to regulation of immunosuppression and inflammation [16].

The increasing knowledge about A_3 ARs, in particular regarding the molecular biology of this subtype, has provided important evidence to consider this receptor as a novel therapeutic target. In addition, it enables rational design and the development of potent and selective A_3 AR ligands as promising therapeutic options for a variety of diseases [17,18]. Therefore, small molecule modulators targeting the A_3 AR have been sought for their potential application as anti-inflammatory, anti-cancer and cardioprotective agents [19–21].

Indoles probably represent one of the most important structural classes in drug discovery [22]. The indole substructure is a basic element for a number of biologically active natural and synthetic products. Indoles are found in a wide range of therapeutically important drugs [23,24]. Over the years, a considerable amount of effort has been made to find indoles with various biological activities and select certain agents for leads in drug discovery research. On account of their potent biological activities, indoles have continued to attract the interest of chemists and biologists alike in drug discovery.

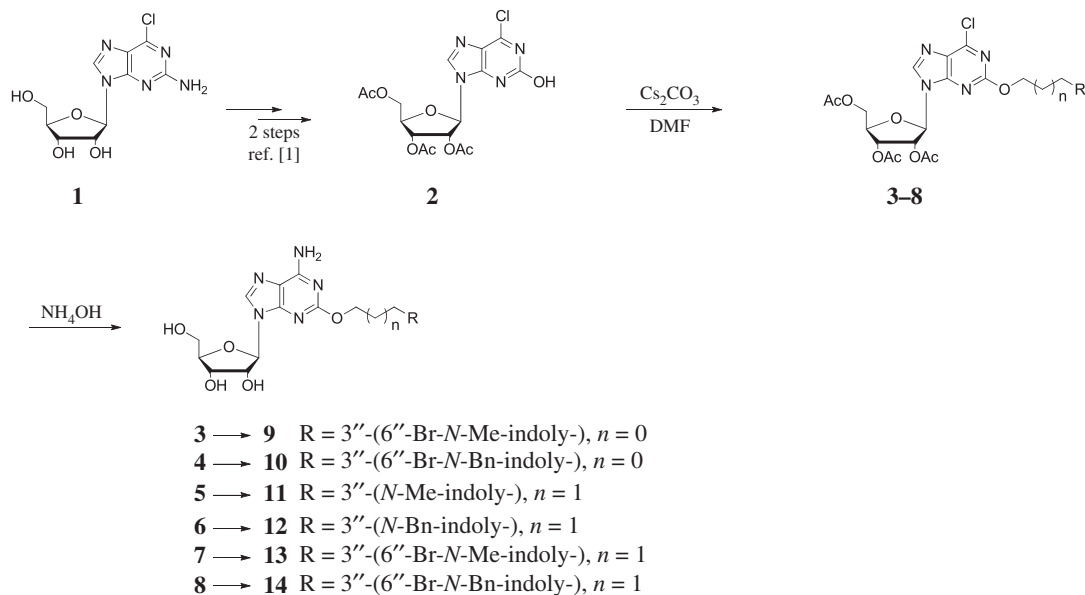
In addition, the scientific community is making intensive efforts to design AR ligands endowed with greater selectivity or to develop innovative compounds acting as receptor modulators. To further explore the importance of indole framework as the basis for AR ligands, we investigate the receptor subdomain that binds the purine moiety by the study of 2-*O*-alkyl derivatives of 2-chloroadenosine. The present paper reports on the synthesis and binding studies of these compounds as well as of adenosine. We describe the lengths of alkyl group in terms of potency at the A_3 receptor as well as receptor subtype selectivity. Here, we use the conversion of the 2-alkynyl group into flexible 2-*O*-alkyl between purine and indole moiety.

2. Results and discussion

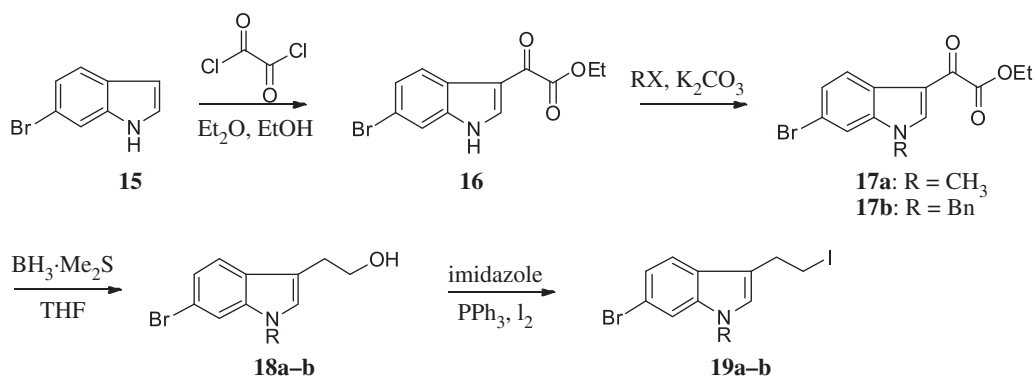
2.1. Synthesis

In an effort to discover new nucleoside analogues as potential A_3 AR ligands, we pursued the 2-oxypurine nucleoside using indole alkyl iodides as shown in scheme 1. The synthesis of the 5'-CH₂OH analogues started from 2-amino-6-chloropurine riboside **1**, which was converted into 6-chloro-2-hydroxy-9-(2,3,5-tri-*O*-acetyl- β -D-ribofuranosyl)purine **2**, as reported [25]. The reaction of the hydroxyl group at the 2-position of **2** with various indole iodides was conducted in the presence of caesium carbonate to affect compounds **3–8**. Simultaneous removal of the acetyl group and amination at the 6-position of **3–8** using ammonium hydroxide solution yielded compounds **9–14**.

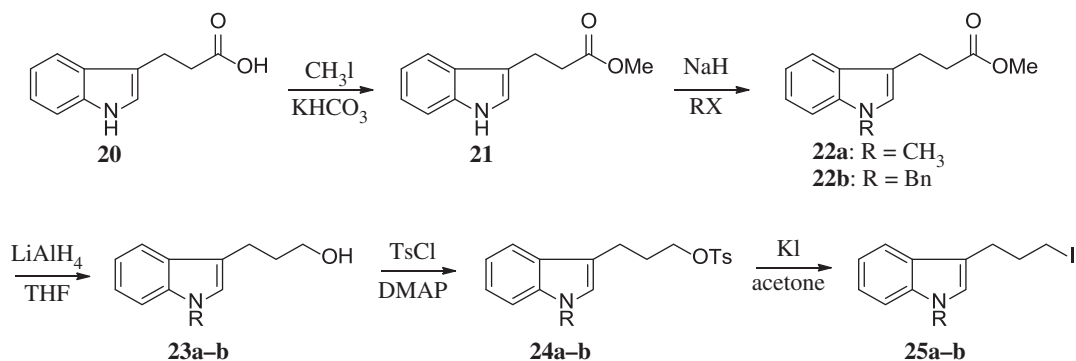
For synthesizing the key compounds, we should synthesize intermediates for the 2-ether component depicted in schemes 2–4. Scheme 2 illustrates the synthesis of two carbons as linker indole iodides. The indole 2-oxoacetate **16** was accessed by the treatment of 6-bromoindole **15** to oxalyl chloride, followed by a reaction with ethanol. Alkylation of indole nitrogen by CH₃I and BnBr provided *N*-alkyl **17a** and **17b**, respectively. The following exposure to BH₃-SMe₂ provided corresponding alcohols **18a** and



Scheme 1. Synthesis of compounds 9–14.



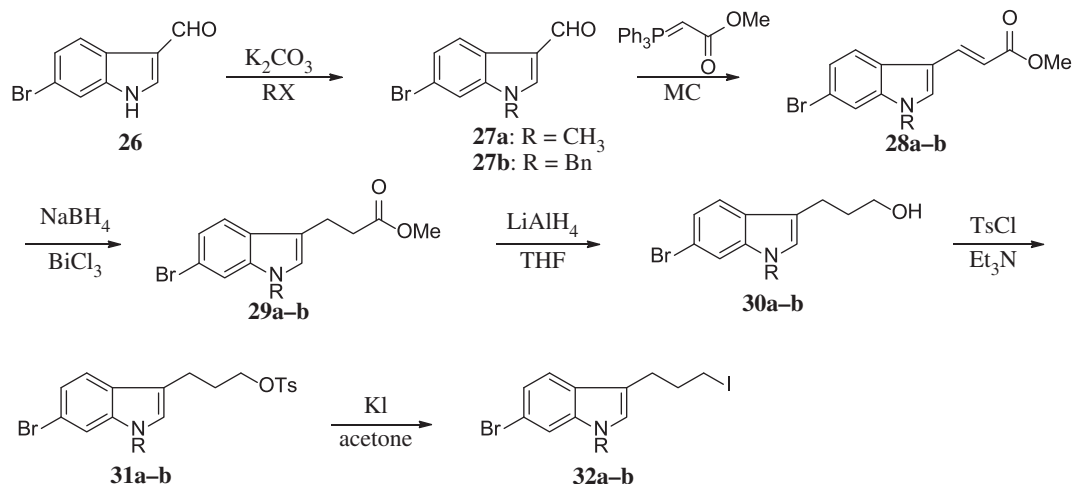
Scheme 2. Synthesis of intermediates 19a–b.



Scheme 3. Synthesis of intermediates 25a–b.

18b. The alcohols were transformed to corresponding iodides **19a–b** by iodine, triphenylphosphine and imidazole.

The three carbons used as the linker of indole iodides shown in scheme 3 were obtained from the commercially available 3-indolepropionic acid **20**. Propionate **21** was prepared by the conversion of carboxylic acid into methyl ester by treatment with CH₃I and KHCO₃. Subsequent N-alkylation of the indole nitrogen provided **22a** and **22b** followed by reduction of carboxylic methyl ester to produce the



Scheme 4. Synthesis of intermediates **32a–b**.

corresponding alcohols **23a** and **23b**. Finally, iodides **25a** and **25b** were derived from **23a–b** via tosylate alcohol, which underwent substitution with iodine.

The synthetic strategy for the preparation of substituted indole derivatives is described in scheme 4. The commercially available 4-bromoindole-3-carboxaldehyde **26** was reacted with CH_3I and BnBr to give rise to *N*-alkylation **27a–b**, respectively. The Wittig reaction of aldehyde provided *trans*-indole acrylate **28a–b**. The following treatment with NaBH_4 and BiCl_3 yielded propionates **29a–b**. LiAlH_4 reduction of the carboxylic ester produced intermediates **30a–b** followed by treatment with tosyl chloride to afford the corresponding **31a–b**, and the tosylate moiety was displaced with good yield with iodine to incorporate the iodo functionality. Detailed synthetic procedures including the yield of reactions and characteristic data can be found in the electronic supplementary material.

2.2. Binding affinity studies

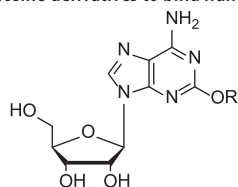
The aim of the present study is to expand knowledge of the structure–activity relationships at the A_3AR and at other subtypes, both in relation to binding affinity and intrinsic efficacy, of adenosine derivatives modified in the 2-position. A screening campaign to discover new scaffolds for A_3AR inhibition yielded the moderate-potency lead **11** (table 1). Among 2-substituted derivatives, 2-ethers were more potent than the corresponding amines or thioethers [26]. The effect of bromine substitution of the phenyl ring was evaluated. This series of 5'-bromo analogue showed a tendency towards increased K_i values with A_1 and A_3ARs , depending on the bulkiness of the bromine atom. Of these analogues, compound **10** was equipotent to **13** with A_1AR . However, its selectivity to binding A_3AR was improved. Unexpectedly, **14** was threefold less potent than **13** in binding to A_3AR and was tolerated by $\text{A}_{2\text{A}}\text{AR}$. Although the 5'-bromo derivative **13** was somewhat equipotent to **11** with A_3AR , its selectivity was reduced.

Interestingly, the 3-indolyl analogue **11** was sevenfold more potent than **12** with A_3AR . The increased size or steric hindrance of the *N*-substituent markedly decreases A_3AR potency. Nevertheless, **12** was less potent than **13** in binding to $\text{A}_{2\text{A}}\text{AR}$. The effect of the space of the alkyl chain between the 2-ethers and the indole moiety was tested. Elongation increased the affinity for A_3AR . Compound **11** showed a fivefold increased potency with A_3AR , while it was somewhat tolerated by $\text{A}_{2\text{A}}\text{AR}$. The affinity of **9** to all three AR subtypes showed low potency compared with that of **13**, similar to the results with **10**.

However, the corresponding 2-indolyl derivative **13** was more potent than compound **11** in affinity for A_1 and $\text{A}_{2\text{A}}\text{Rs}$. The bulkiness of the *N*-substituent may be related to the increased affinity. Compound **14** was more potent than **12** in binding to all three ARs. Meanwhile, compound **11** displayed a fivefold potency enhancement over **9** with A_3AR . The *N*-Bn derivative **10** with potency close to that of **9** was invariant in affinity for A_1 and $\text{A}_{2\text{A}}\text{Rs}$.

2.3. Molecular docking analysis

Driven by docking of several derivatives with hA_3R , we performed the molecular modelling studies [27] to explore the binding modes of all six aforementioned compounds as shown in table 2. Among

Table 1. Potency of 2-alkoxyadenosine derivatives to bind human A₁, A_{2A} and A₃ARs expressed in CHO cells.^a

entry	R	K_i (nM \pm SEM) or % inhibition at 10 μ M		
		hA ₁ AR ^b	hA _{2A} AR ^b	hA ₃ AR ^b
9		(47 \pm 5%)	2770 \pm 500	679 \pm 149
10		217 \pm 57	380 \pm 81	532 \pm 144
11		300 \pm 70	880 \pm 200	111 \pm 30
12		(34 \pm 2%)	(56 \pm 5%)	731 \pm 209
13		230 \pm 26	262 \pm 192	177 \pm 43
14		152 \pm 49	371 \pm 79	491 \pm 143
	adenosine	—	—	290

^aAll experiments were performed on CHO cells stably expressing one of three subtypes of human ARs. The binding affinities for A₁, A_{2A} and A₃ARs were expressed as K_i values and were determined using agonist radioligands ([³H]CCPA), ([³H]CGS21680) and [¹²⁵I]I-AB-MECA, respectively. Values in parentheses are for weak binding, corresponding to an IC₅₀ \geq 10 μ M. Data are expressed as mean \pm s.e.

^b K_i in binding, unless noted.

Table 2. The predicted free energy and affinity compared with experimental values.

entry	experimental K_i (hA ₃ AR, nM)	predicted free energy (kcal mol ⁻¹)	predicted affinity (nM)
11	111 \pm 30	-9.08	222.18
13	177 \pm 43	-8.73	398.82
9	679 \pm 149	-8.38	719.10

these compounds, the compounds cpd10, 12 and 14 with the N-Bn substituent failed to dock into the ligand-binding site of the protein. The current docking results are consistent with the biological studies mentioned above and serve as an explanation for the sharply reduced affinity. On the contrary, all other three compounds (cpd9, 11 and 13) with N-methyl substituent were found to dock into the binding pocket of the protein of interest and further interact with the amino acids GLN167, PHE168, ASN250, etc., through hydrogen bonding or hydrophobic interactions (figures 1 and 2). Detailed information for docking experiment can be found in the electronic supplementary material. In particular, the common interactions for these three compounds were the hydrogen bonding with the amino acid ASN250 of the protein, which is shown to be closely relevant to antagonist interaction [28,29]. Furthermore, the docking

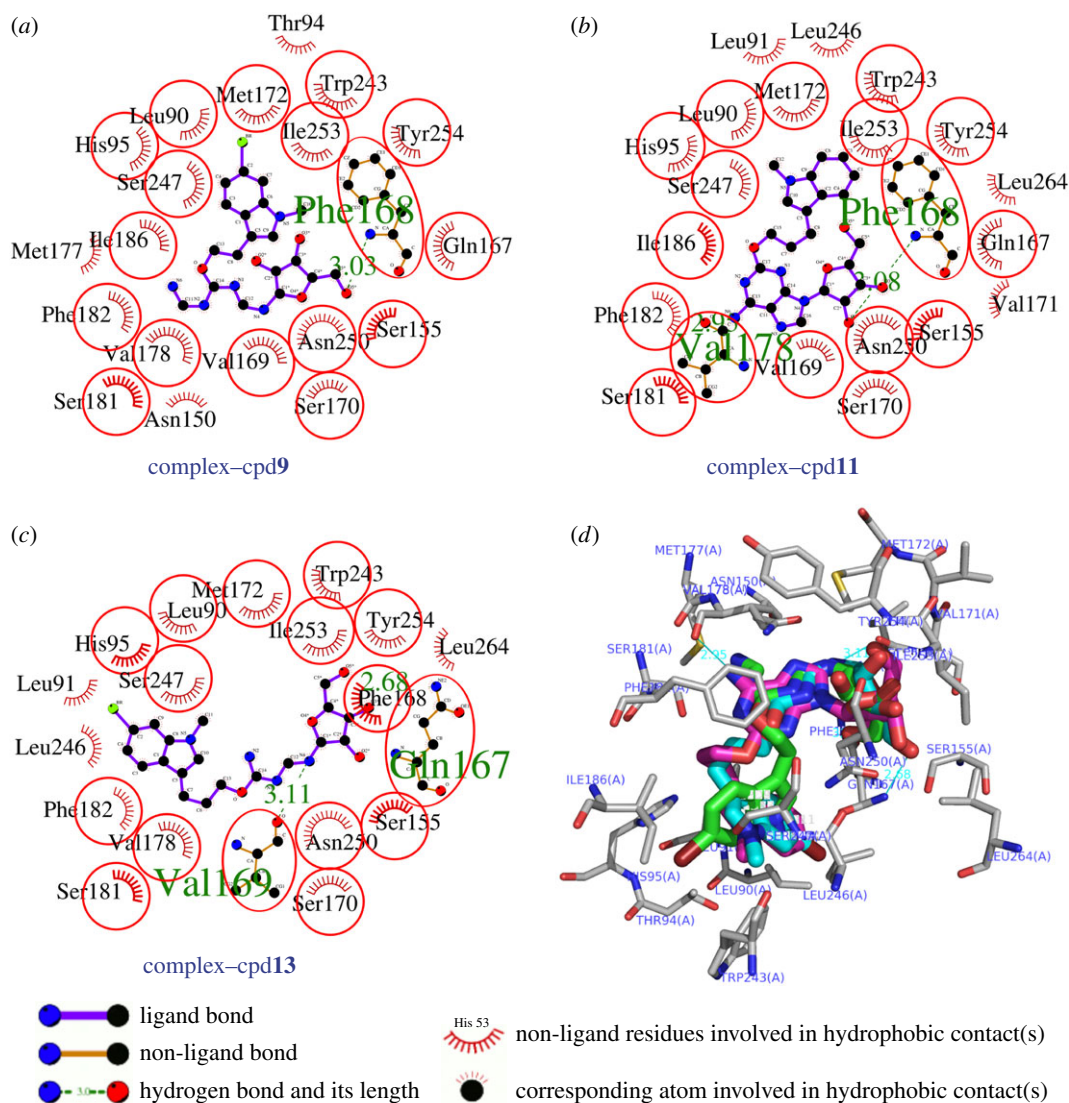


Figure 1. The two-dimensional image of ligand-hA₃AR interactions with the hydrogen bonds and hydrophobic interactions (a–c); (d) the cpd₉ is coloured green, the cpd₁₁ is coloured magenta, the cpd₁₃ is coloured cyan, respectively. The surrounding residues interacting with these ligands are also shown and labelled. The meaning of the items on the two-dimensional plot above is listed in the bottom panel.

results showed that the compound cpd₁₁ with the highest K_i value has the highest predicted affinity ($-9.08 \text{ kcal mol}^{-1}$, table 2).

Figure 2 depicts the binding mode of the compound cpd₁₁ in the ligand-binding pocket of the protein in details. Cpd₁₁ can be well docked into the binding site of interest. In the most potent molecule cpd₁₁, the adenosine core contributed strongly to the binding affinity, which also demonstrates the rationality of the core as a key scaffold for the further chemical modifications. The hydroxyl oxygen of furan ring and the 6-position *N* atom of adeny group interact with the ASN250, GLN167, PHE168 and VAL178 through hydrogen bonding. On the other hand, the high affinity also requires the presence of the 2-*O*-alkyl-substituted groups for producing the strong hydrophobic interactions.

From these results, we have illustrated the interactions of our newly synthesized 2-*O*-alkyl-substituted adenosine analogues with a ligand-binding site of hA₃AR from the molecular modelling point of view. The results also showed the different roles of the substitutions which are strongly linked to the increased or decreased affinity. The exploration for these interactions can provide us the guidance for the future chemical modifications.

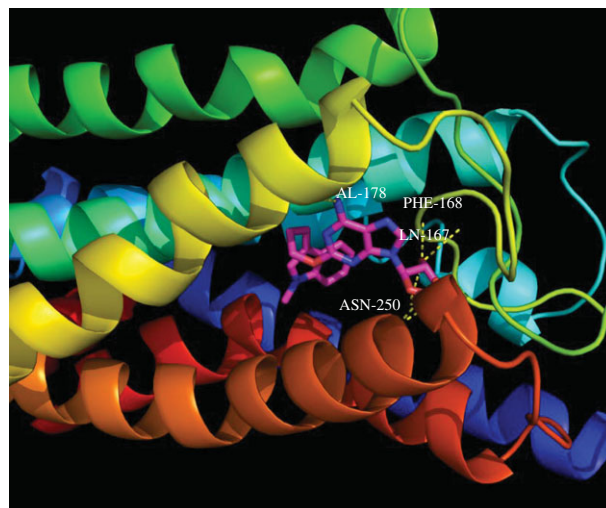


Figure 2. The most active compound cpd11 (magenta colour) in the active site of human adenosine A_3 receptor showing hydrogen bonding interactions.

3. Conclusion

In this work, we designed and synthesized a series of 2-*O*-alkyl-substituted adenosine analogues with indole moiety. The 2-substituents **11** was the most potent among the series, and it was confirmed to be a modulator in a functional assay measuring its capacity to bind receptors in CHO cells expressing the hA₃A receptor. We found that 2-substituents **11** were critical structural determinant for A₃AR ligands ($K_i = 111$ nM). The promising compound can be considered a valuable seed for the design and development of new and even more selective and potent compounds. The molecular modelling studies have also been performed to investigate the binding mode of the potential compound in the ligand-binding pocket of human A₃ receptor. Here, this study provides useful foundations for the attainment of a detailed pharmacological and physiological characterization of the adenosine A₃ receptor.

Data accessibility. The detailed experimental synthetic procedures and spectra of the final compounds are provided in the electronic supplementary material.

Authors' contributions. Y.X. carried out the synthetic work and performed the NMR experiments. X.Z. carried out the biological screening. R.H. helped analyse and interpret the data. J.W. conceived the study, designed it and drafted the manuscript. All authors gave final approval for publication.

Competing interests. The authors declare that they have no competing interests.

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References

- Huang ZL, Zhang Z, Qu WM. 2014 Roles of adenosine and its receptors in sleep wake regulation. *Int. Rev. Neurobiol.* **119**, 349–371. (doi:10.1016/B978-0-12-801022-8.00014-3)
- Fredholm BB, Abbracchio MP, Burnstock G, Daly JW, Harden TK, Jacobson KA, Leff P, Williams M. 1994 Nomenclature and classification of purinoceptors. *Pharmacol. Rev.* **46**, 143–156.
- Sawynok J. 2007 Adenosine and ATP receptors. *Handb. Exp. Pharmacol.* **177**, 309–328.
- Sato A. 2005 Mechanism of vasodilation to adenosine in coronary arterioles from patients with heart disease. *Am. J. Physiol. Heart Circ. Physiol.* **288**, H1633–H1640. (doi:10.1152/ajpheart.00575.2004)
- Costa F, Biaggioni I. 1998 Role of nitric oxide in adenosine-induced vasodilation in humans. *Hypertension* **31**, 1061–1064. (doi:10.1161/01.HYP.31.5.1061)
- Morgan JM, McCormack DG, Griffiths MJ, Morgan CJ, Barnes PJ, Evans TW. 1991 Adenosine as a vasodilator in primary pulmonary hypertension. *Circulation* **84**, 1145–1149. (doi:10.1161/01.CIR.84.3.1145)
- Haskó G, Linden J, Cronstein B, Pacher P. 2008 Adenosine receptors: therapeutic aspects for inflammatory and immune diseases. *Nat. Rev. Drug Discov.* **7**, 759–770. (doi:10.1038/nrd2638)
- Hilaire CS, Carroll S, Chen H, Ravid K. 2009 Mechanisms of induction of adenosine receptor genes and its functional significance. *J. Cell Physiol.* **218**, 35–44. (doi:10.1002/jcp.21579)
- Baraldi P, Tabrizi M, Gessi S, Borea P. 2008 Adenosine receptor antagonists: translating medicinal chemistry and pharmacology into clinical utility. *Chem. Rev.* **108**, 238–263. (doi:10.1021/cr0682195)
- Antonioni L, Csoka B, Fornai M, Colucci R, Kokai E, Blandizzi C, Haskó G. 2014 Adenosine and inflammation: what's new on the horizon? *Drug Discov. Today* **19**, 1051–1068.

11. Müller CE. 2007 Adenosine receptor ligands-recent developments part I. Agonists. *Curr. Top. Med. Chem.* **7**, 1269–1288. (doi:10.2174/0929867003374101)
12. Linden J. 1994 Cloned adenosine A₃ receptors: pharmacological properties, species differences and receptor functions. *Trends Pharmacol. Sci.* **15**, 298–306. (doi:10.1016/0165-6147(94)90011-6)
13. Fredholm BB, Ilzerman AP, Jacobson KA, Klotz KN, Linden J. 2001 International union of pharmacology. XXV. Nomenclature and classification of adenosine receptors. *Pharmacol. Rev.* **53**, 527–552.
14. Jacobson KA, Gao ZG. 2006 Adenosine receptors as therapeutic targets. *Nat. Rev. Drug Discovery* **5**, 247–264. (doi:10.1038/nrd1983)
15. Borea PA, Varani K, Vincenzi F, Baraldi PG, Tabrizi MA, Merighi S, Gessi S. 2015 The A₃ adenosine receptor: history and perspectives. *Pharmacol. Rev.* **67**, 74–102. (doi:10.1124/pr.113.008540)
16. Gaspar A, Reis J, Kachler S, Paoletta S, Uriarte E, Klotz KN, Moro S, Borges F. 2012 Discovery of novel A₃ adenosine receptor ligands based on chromone scaffold. *Biochem. Pharmacol.* **84**, 21–29. (doi:10.1016/j.bcp.2012.03.007)
17. Luan F, Borges F, Cordeiro MN. 2012 Recent advances on A₃ adenosine receptor antagonists by QSAR tools. *Curr. Top. Med. Chem.* **12B**, 78–894.
18. Melani A, Pugliese AM, Pedata F. 2014 Adenosine receptors in cerebral ischemia. *Int. Rev. Neurobiol.* **119**, 309–348. (doi:10.1016/B978-0-12-801022-8.00013-1)
19. Samsel M, Dzierzbicka K. 2011 Therapeutic potential of adenosine analogues and conjugates. *Pharmacol. Rep.* **63**, 601–617. (doi:10.1016/S1734-1140(11)70573-4)
20. Samsel M, Dzierzbicka K, Trzonkowski P. 2013 Adenosine, its analogues and conjugates. *Postepy Hig. Med. Dosw.* **67**, 1189–1203. (doi:10.5604/17322693.1078588)
21. Samsel M, Dzierzbicka K, Trzonkowski P. 2014 Synthesis and antiproliferative activity of conjugates of adenosine with muramyl dipeptide and nor-muramyl dipeptide derivatives. *Bioorg. Med. Chem. Lett.* **24**, 3587–3591. (doi:10.1016/j.bmcl.2014.05.043)
22. Lalit K, Shashi B, Kamal J. 2012 The diverse pharmacological importance of indole derivatives: a review. *Intl. J. Res. Pharm. Sci.* **2**, 23–33.
23. de Sá Alves FR, Barreiro EJ, Fraga CA. 2009 From nature to drug discovery: the indole scaffold as a 'privileged structure'. *Mini Rev. Med. Chem.* **9**, 782–793. (doi:10.2174/138955709788452649)
24. Horton DA, Bourne GT, Smythe ML. 2003 The combinatorial synthesis of bicyclic privileged structures or privileged substructures. *Chem. Rev.* **103**, 893–930. (doi:10.1021/cr020033s)
25. Moorman AR. 2003 WO patent, 2003035662.
26. Gao ZG, Mamedova L, Chen P, Jacobson KA. 2004 2-Substituted adenosine derivatives: affinity and efficacy at four subtypes of human adenosine receptors. *Biochem. Pharmacol.* **68**, 1985–1993. (doi:10.1016/j.bcp.2004.06.011)
27. Goodsell DS, Morris GM, Olson AJ. 1996 Automated docking of flexible ligands: applications of AutoDock. *J. Mol. Recognit.* **9**, 1–5. (doi:10.1002/(SICI)1099-1352(199601)9:1<1::AID-JMR241>3.0.CO;2-6)
28. Wei J, Li H, Qu W, Gao Q. 2009 Molecular docking study of A₃ adenosine receptor antagonists and pharmacophore-based drug design. *Neurochem. Int.* **55**, 637–642. (doi:10.1016/j.neuint.2009.06.006)
29. Laskowski RA, Swindells MB. 2011 LigPlot+ : multiple ligand–protein interaction diagrams for drug discovery. *J. Chem. Inf. Model.* **51**, 2778–2786. (doi:10.1021/ci200227u)