



Cite this article: Sousa LM, Vilarinho LM, Ribeiro GH, Bogado AL, Dinelli LR. 2017 An electronic device based on gold nanoparticles and tetraruthenated porphyrin as an electrochemical sensor for catechol. *R. Soc. open sci.* **4**: 170675.

<http://dx.doi.org/10.1098/rsos.170675>

Received: 13 June 2017

Accepted: 16 November 2017

Subject Category:

Chemistry

Subject Areas:

inorganic chemistry/supramolecular chemistry

Keywords:

dihydroxybenzene, electrode modified, electropolymerization, cobalt porphyrin

Author for correspondence:

Luís R. Dinelli

e-mail: dinelli@ufu.br

This article has been edited by the Royal Society of Chemistry, including the commissioning, peer review process and editorial aspects up to the point of acceptance.



An electronic device based on gold nanoparticles and tetraruthenated porphyrin as an electrochemical sensor for catechol

Luana M. Sousa¹, Luana M. Vilarinho¹, Gabriel H. Ribeiro², André L. Bogado¹ and Luís R. Dinelli¹

¹Faculdade de Ciências Integradas do Pontal, Universidade Federal de Uberlândia, Rua vinte, 1600, CEP 38304-402, Ituiutaba, Minas Gerais, Brazil

²Departamento de Química, Universidade Federal de São Carlos, CP 676, CEP 13565-905, São Carlos, Sao Paulo, Brazil

LRD, 0000-0003-4199-6572

The aim of this study was to obtain an electrochemical device between the electrostatic interaction of the electropolymerized porphyrin {CoTPyP[RuCl₃(dppb)]₄}, where TPyP = 5,10,15, 20-tetrapyrrodimethylporphyrin and dppb = 1,4-bis(diphenylphosphino)butane, and gold nanoparticles (AuNPsⁿ⁻), to be used as a voltammetric sensor to determine catechol (CC). The modified electrode, labelled as [(CoTPRu₄)_n⁸⁺-BE]/AuNPsⁿ⁻ {where BE = bare electrode = glassy carbon electrode (GCE) or indium tin oxide (ITO)}, was made layer-by-layer. Initially, a cationic polymeric film was generated by electropolymerization of the {CoTPyP[RuCl₃(dppb)]₄} onto the surface of the bare electrode to produce an intermediary electrode [(CoTPRu₄)_n⁸⁺-BE]. Making the final electronic device also involves coating the electrode [(CoTPRu₄)_n⁸⁺-BE] using a colloidal suspension of AuNPsⁿ⁻ by electrostatic interaction between the species. Therefore, a bilayer labelled as [(CoTPRu₄)_n⁸⁺-BE]/AuNPsⁿ⁻ was produced and used as an electrochemical sensor for CC determination. The electrochemical behaviour of CC was investigated using cyclic voltammetry at [(CoTPRu₄)_n⁸⁺-GCE]/AuNPsⁿ⁻ electrode. Compared to the GCE, the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPsⁿ⁻ showed higher electrocatalytic activity towards the oxidation of CC. Under the optimized conditions, the calibration curves for CC were 21–1357 μmol l⁻¹ with a high sensitivity of 108 μA μmol l⁻¹ cm⁻². The detection limit was 1.4 μmol l⁻¹.

1. Introduction

Catechol (CC) is a phenolic compound commonly used in the cosmetic industry, for tanning, flavouring agents, pesticides, medicines, antioxidant, dye, photography, etc [1]. The lethal human dose of CC is 50–500 g kg⁻¹ or one teaspoon given once to a person weighing 70 kg and death results from pulmonary failure [2]. Therefore, it is important to establish a sensitive, fast and convenient method for the detection and quantification of CC. Various techniques including liquid chromatography [3], capillary electrochromatography [4], electrochemiluminescence [5] and spectrophotometry [6] have been used to detect polyphenols. As CC presents excellent electrochemical processes, there are various electrochemical methods for its determination which can be found in the literature [7–15]. Compared with other methods, electrochemical methods have advantages such as being simple, sensitive and selective for samples with small concentrations.

Many materials involving metals, various nanotubes and nanoparticles, graphene and fullerene and porphyrin and their derivatives have been extensively used to construct electrochemical sensors [16–30]. Considering this, porphyrin, metalloporphyrin and their derivatives, have received much attention as they show excellent electrocatalytic properties [31]. Moreover, porphyrins and their metal derivatives are potential candidates for the construction of electrochemical sensors and catalytic applications as they present an extensively conjugated two-dimensional π -system and special electrochemical properties [32,33].

In addition, gold nanoparticles (AuNPsⁿ⁻) have aroused considerable interest in applications as electrochemical sensors because they are easily synthesized and manipulated [34–38]. AuNPsⁿ⁻ has a unique optical and electrochemical property that significantly depends on its morphological and physiological characteristics, i.e. on its size, shape and aggregation state and can be easily controlled by the synthesis methodology [39,40]. In addition, because nanoparticles have a large volume/surface ratio, when placed on the surface of the electrode they can increase their effective area and, owing to the spherical morphology, the interstitial space allows for an efficient mass transfer. Regarding the metallic nature of the nanoparticles, the conductivity of the electrode can be increased, improving its sensitivity and selectivity [41,40]. In recent years, interest in the study of aggregates of AuNPsⁿ⁻ with ruthenium complex, labelled as [Ru]⁺/AuNPsⁿ⁻, which takes place through a self-assembly process owing to the electrostatic interaction between species, has increased. These aggregates have been used in electrochemical devices [42,43] and homogeneous catalysis [44]. As a result, improvements in electrochemical responses, such as sensibility, stability, selectivity and catalytic activity, have been observed in the presence of AuNPsⁿ⁻.

Many efforts have been made to develop modified electrodes using the electropolymerization of tetraruthenated porphyrin and use them as electrochemical sensors for phenolic compounds [45–47]. The biggest advantage of using electropolymerization is that it controls the number of layers deposited on the surface of a bare electrode (BE), such as glassy carbon electrode (GCE) or indium tin oxide (ITO). This process can be controlled by voltammetric cycles, and consequently the polymeric film thickness formed on the electrode. Recently, the simultaneous determination of CC and hydroquinone (HQ) was reported using a modified GCE with the {VOTPyP[RuCl₃(dppb)]₄} porphyrin [46]. In this study, it was demonstrated that the modified electrode (VOTPRu-GCE) improves the electrochemical response of CC and HQ when compared with the bare GCE. The electrochemical behaviours of the two isomers were reversible at the VOTPRu-GCE with two pairs of well-defined voltammetric peaks for the electrochemical processes of the analytes. Furthermore, the redox peak currents and sensitivity in the determination of CC and HQ were also greatly enhanced compared to the BE, 12.73 and 15.91 $\mu\text{A } \mu\text{mol L}^{-1} \text{cm}^{-2}$, respectively, at VOTPRu-GCE. These results suggest that the electropolymerized film on the GCE surface can accelerate the electron transfer between analytes and electrode, which is probably caused by a synergistic effect of the vanadium ion in the centre of a porphyrin ring with the complexes containing ruthenium at the peripheral position.

The interaction and synergistic effect with layer–layer assembly of [(CoTPRu₄)_n⁸⁺-BE]/AuNPsⁿ⁻ (where BE = bare electrode = GCE or ITO) and AuNPsⁿ⁻ = gold nanoparticles was investigated. These electronic devices were characterized by UV/Vis measurements and atomic force microscopy (AFM) images and they were used as an electrochemical sensor for CC. The [(CoTPRu₄)_n⁸⁺-GCE]/AuNPsⁿ⁻ showed an outstanding performance in the determination of CC in 0.1 mol l⁻¹ acetate buffer at pH 4.76. The viability of the sensor was evaluated, such as the linear range, detection limit, sensitivity, stability and reproducibility.

2. Material and methods

2.1. Materials

CC was purchased from Vetec and all the chemicals used were of reagent grade or comparable purity. Methanol, dichloromethane and ether were distilled prior to use. All the aqueous solutions were prepared with ultra-pure water (ASTM type I, 18 M Ω cm of resistivity). Ruthenium(III) chloride hydrate, 1,4-bis(diphenylphosphino)butane (dppb), chloroauric acidH [AuCl₄], tetrabutylammonium hexafluorophosphate (TBAH) and cobalt (II) acetate salt were purchased from Aldrich and they were used as received. Acetate buffer solutions (HAc-NaAc, 0.1 mol l⁻¹) with different pH values were prepared by mixing 0.1 mol l⁻¹ of acetic acid (HAc) and 0.1 mol l⁻¹ of sodium acetate (NaAc) and the pH was adjusted by NaOH or HCl. Purified argon atmosphere was used in all procedures described to remove the dissolved oxygen.

2.2. Apparatus

Voltammetric experiments were performed on a μ AUTOLAB type III potentiostat/galvanostat with a conventional three-electrode system. A bare or modified GCE (geometric diameter = 0.2 cm) or ITO electrode was used as a working electrode. A KCl saturated Ag/AgCl (AgCl_(sat)) electrode was used as a reference electrode and a platinum wire as a counter electrode. All the experiments were carried out at room temperature. The cyclic voltammetry (CV) was performed from -0.4 to 1.0 V versus Ag/AgCl_(s) with a scan rate of 100 mV s⁻¹, sample interval, 0.0001 V and quiet time of 2 s. Under the conditions used, E₀ for the one-electron oxidation of [Fe(η^5 -C₅H₅)₂], added to the test solutions as an internal standard, is +0.43 V.

UV/Vis spectra were recorded on a Shimadzu spectrophotometer, model UV-1800, coupled with a thermoelectrically temperature controlled cell TCC-100 (at 25 \pm 0.1°C), using a quartz cell (1 cm), between 200 and 800 nm. Elemental analyses were performed with a Thermo Scientific CHNS-O FLASH 2000 micro analyzer. AFM measurements were performed using a Nanoscope V Veeco/Bruker with a scan assist.

2.3. Preparation of gold nanoparticles

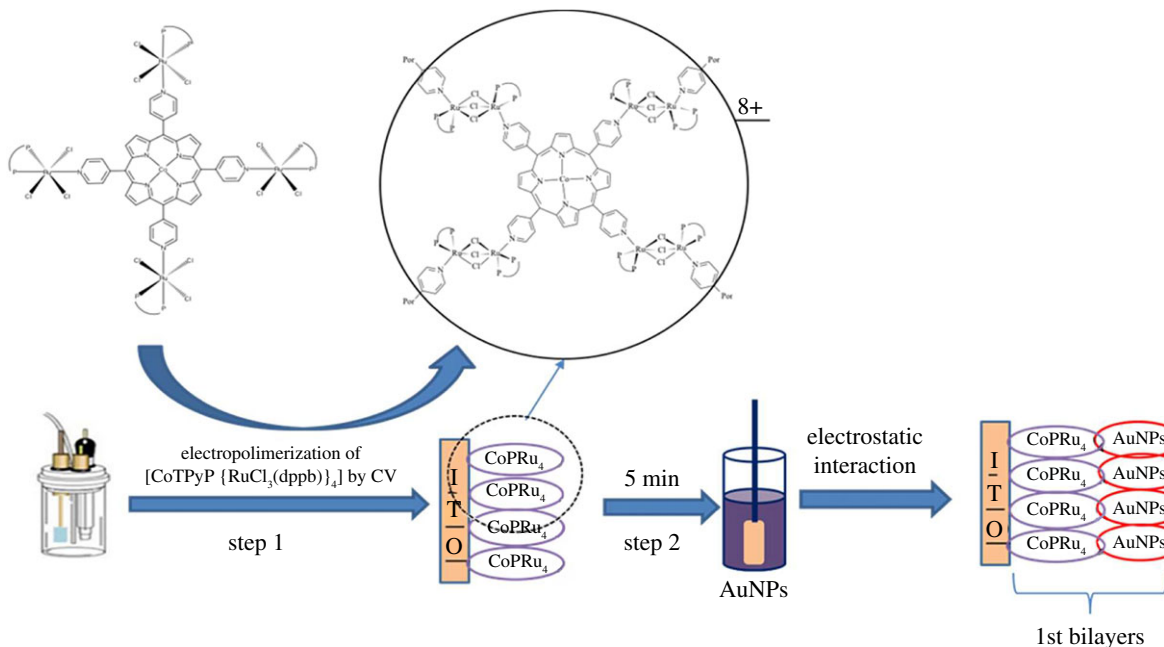
AuNPsⁿ⁻ with an average diameter of 5 nm were prepared by reduction of H[AuCl₄] in a biphasic mixture of water/toluene (1 : 1) with sodium citrate tribasic dihydrate and sodium tetrahydridoborate. Hexadecyltrimethylammonium bromide (HDTBr) was used as a transfer reagent phase by modifying a procedure described in the literature [48]. In brief, 1.0 ml of 1% H[AuCl₄] was added to 90 ml of H₂O at room temperature (20–23°C). After 1 min of stirring, 2.0 ml of sodium citrate tribasic dihydrate (38.8 mmol l⁻¹) was added. The solution was stirred for another 1 min, and then 1.0 ml of fresh 0.075% NaBH₄ was added to the solution of sodium citrate tribasic dihydrate (38.8 mmol l⁻¹). After this, 20 ml of HDTBr in toluene solution (0.13 mol l⁻¹) was mixed with aqueous AuNPsⁿ⁻. The resulting solution was kept under stirring for 20 min to move the AuNPsⁿ⁻ swiftly from an aqueous phase to an organic phase. Afterwards, the colloidal solution was allowed to stand for phase separation, and then the mixture was separated using an analytical funnel and the organic phase was stored in a dark bottle at 4°C.

2.4. Synthetic procedures

The 5,10,15,20-tetra(4-pyridyl)porphyrin (H₂TPyP) and 5,10,15,20-tetra(4-pyridyl)porphyrincobalt(II) [CoTPyP] were synthesized according to procedures reported in the literature [49]. The synthesis of the {CoTPyP[RuCl₃(dppb)]₄} complex was performed by modifying a procedure described in the literature [47]. Six milligrams (8.88 μ mol) of [CoTPyP] [50,51] and 23.1 mg (3.55 μ mol) of *mer*-[RuCl₃(dppb)H₂O] [52] were solubilized in a mixture of 27 ml of dichloromethane and 3 ml of methanol. The solution was stirred for 20 h and the resulting mixture was concentrated to about 3 ml. Then, Et₂O (20 ml) was added to precipitate a brown-red solid. The complex was collected, washed with Et₂O (three times, 10 ml) and dried under vacuum.

2.5. Preparation of the modified electrode

The modified electrode was obtained by modifying a procedure described in the literature [46]. Initially, the GCE surface was polished in alumina slurry (3 μ m) and washed with ultra-pure water in an ultrasonic



Scheme 1. Formation of film on the surface of the ITO electrode in two steps.

bath. The following procedure was performed to produce the first bilayer film: the electropolymerization of tetra-ruthenated porphyrin on the GCE surface was carried out in dichloromethane solution (0.1 mol l^{-1} TBAH used as a supporting electrolyte) of 0.1 mmol l^{-1} $\{\text{CoTPyP}[\text{RuCl}_3(\text{dppb})]_4\}$ by one voltammetric cycle in the range of potential between -0.4 and $+1.0 \text{ V}$ at a scan rate of 100 mV s^{-1} . The modified GCE $[(\text{CoTPRu}_4)_n]^{8+}$ -GCE was washed with acetone and ultra-pure water and was then added to a solution for 5 min containing 10 ml of gold nanoparticles (AuNPs^{n-} , 0.05 mol l^{-1} of Au). The new modified electrode, labelled as $[(\text{CoTPRu}_4)_n]^{8+}$ -GCE/ AuNPs^{n-} , was washed with water to remove the excess of non-immobilized AuNPs^{n-} . This procedure was repeated threefold before applying the $[(\text{CoTPRu}_4)_n]^{8+}$ -GCE/ AuNPs^{n-} as a work electrode. The $[(\text{CoTPRu}_4)_n]^{8+}$ -GCE/ AuNPs^{n-} was conditioned with 50 voltammetric cycles in NaAc solution (0.1 mol l^{-1}) to stabilize the current and potential. The same procedure was used to build the electrode on the ITO surface. The ITO electrode was not used for electrochemical measurements of CC because it has low electrical conductivity.

3. Results and discussion

3.1. Preparation and characterization of the modified electrode

The preparation of the modified electrode, labelled as $[(\text{CoTPRu}_4)_n]^{8+}$ -BE/ AuNPs^{n-} {where BE = bare electrode = GCE or ITO}, was done layer-by-layer in two basic steps (see scheme 1). The first step requires the electropolymerization, by CV, of the porphyrin $\{\text{CoTPyP}[\text{RuCl}_3(\text{dppb})]_4\}$ on the surface of the electrode, making a voltammetric cycle in the potential range between -0.4 and $+1.0 \text{ V}$ at a scan rate of 100 mV s^{-1} . The second step involves a coating of the electropolymerized porphyrin with a colloidal solution of AuNPs^{n-} .

The electropolymerized porphyrin produces a cationic species of mixed-valence $\{\text{CoTPyP}[\text{Ru}(\text{dppb})_4(\text{Cl}_3)_2]_{2n}^{8n+}\}$, which adsorbs on the electrode surface (figure 1). The mechanism of the electropolymerization, including all characterizations of the film, was previously published by our research group [52] and involves the reduction of 'RuCl₃(dppb)' moiety ($\text{Ru}^{\text{III}} \rightarrow \text{Ru}^{\text{II}}$) at 0 V . The quasi-reversible redox process of $\text{Ru}^{\text{II}}\text{Ru}^{\text{III}} \rightarrow \text{Ru}^{\text{III}}\text{Ru}^{\text{III}}$ couple is observed at $E_{1/2} = 0.55 \text{ V}$, assigned to the formation of the films of the mixed-valence complex $\{\text{CoTPyP}[\text{Ru}(\text{dppb})_4(\text{Cl}_3)_2]_{2n}^{8n+}\}$. The aggregation of AuNPs^{n-} on the cationic polymeric film occurs by physical adsorption using electrostatic interactions [42,44].

The two basic steps described above were carried out threefold, which produced a total of three bilayers, that presented the optimal condition for CC sensing. In figure 1, it can be observed that the peak currents increase with the number of cyclic voltammograms, owing to the increase in the immobilized

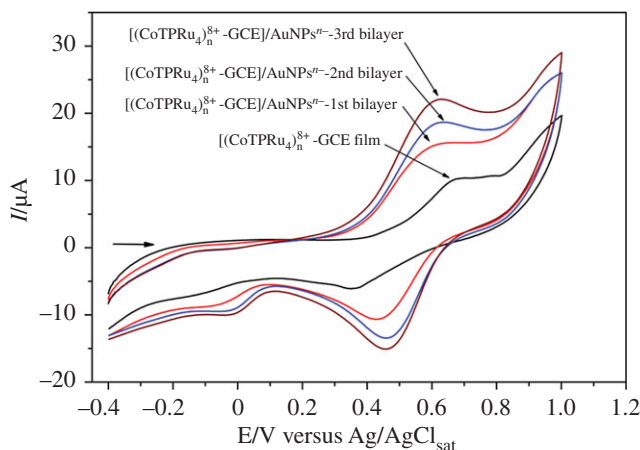


Figure 1. Electropolymerization process of $1 \text{ mmol l}^{-1} \{[\text{CoTPyP}(\text{RuCl}_3(\text{dppb}))_4]\}$ in dichloromethane (0.1 mol l^{-1} TBAH) on ITO electrode for each bilayer $[(\text{CoTPRu}_4)_n^{8+}\text{-ITO}]/\text{AuNPs}^{n-}$ by voltammetric cycles in the range of -0.4 to 1.0 V at 100 mV s^{-1} .

species on the electrode surface. Figure 2a shows the UV/Vis spectra of the AuNPs^{n-} in the toluene solution and $[\text{CoTPyP}(\text{RuCl}_3(\text{dppb}))_4]$ in the CH_2Cl_2 solution. The Soret band of porphyrin and Plasmon band of AuNPs^{n-} with a wavelength at 425 and 525 nm , respectively, can be observed. In figure 2b, the UV/Vis spectra after the formation of each bilayer reveal the presence of Soret and Q bands, related to the electropolymerized porphyrin and a Plasmon band centralized at 530 nm , related to the AuNPs^{n-} . These results show that the two species are immobilized onto the surface of the ITO electrode, producing the desired modified electrode labelled as $[(\text{CoTPRu}_4)_n^{8+}\text{-ITO}]/\text{AuNPs}^{n-}$.

AFM also characterized the modified electrode $[(\text{CoTPRu}_4)_n^{8+}\text{-ITO}]/\text{AuNPs}^{n-}$ (figure 3). The AFM image shows an average roughness of 54.5 nm in an area of $107 \mu\text{m}^2$, when three bilayers were bonded onto the ITO surface. This value is higher than the electropolymerized porphyrin $[\text{Mn}(\text{H}_2\text{O})_2\text{TPyP}(\text{RuCl}_3(\text{dppb}))_4](\text{PF}_6)$, which is a similar system without AuNPs^{n-} , with a roughness value of 4.5 nm , even after five cyclic voltammograms [45]. This result shows that the immobilization of the AuNPs^{n-} on the surface of the electrode causes an increase in the roughness of the film. Undoubtedly, the increasing electroactive area is directly associated with the presence of AuNPs^{n-} .

3.2. Electrochemical behaviour of catechol

Efforts were made in order to evaluate the $[(\text{CoTPRu}_4)_n^{8+}\text{-BE}]/\text{AuNPs}^{n-}$ (where BE = bare electrode = GCE or ITO) as a voltammetric sensor for CC. The best results were obtained when GCE was used as a holder electrode and the performance of $[(\text{CoTPRu}_4)_n^{8+}\text{-GCE}]/\text{AuNPs}^{n-}$ was compared with the $[(\text{CoTPRu}_4)_n^{8+}\text{-GCE}]$ and nude GCE. Figure 4 shows the cyclic voltammograms of CC (0.30 mmol l^{-1}), using the nude GCE as a work electrode, exhibiting the anodic peak potential (E_{pa}) at 516 mV and cathodic peak potential (E_{pc}) at -126 mV . The potential peak difference (ΔE_{p}) was 642 mV , which means that the oxidation of the CC is an irreversible process in the uncovered GCE. However, the work electrode was altered to the $[(\text{CoTPRu}_4)_n^{8+}\text{-GCE}]$, the electrochemical behaviour of the CC was changed from irreversible to quasi-reversible, with the E_{pa} at 251 mV and E_{pc} at 126 mV , and a significant decrease in the $\Delta E_{\text{p}} = 125 \text{ mV}$. Finally, in the third circumstance, when the $[(\text{CoTPRu}_4)_n^{8+}\text{-GCE}]/\text{AuNPs}^{n-}$ was used as a sensor for CC detection, well-resolved redox processes were also observed, $E_{\text{pa}} = 334 \text{ mV}$; $E_{\text{pc}} = 209 \text{ mV}$; and $\Delta E_{\text{p}} = 125 \text{ mV}$, related to the exposed GCE. The main advantage of AuNPs^{n-} on the electrode was a significant increase in the peak currents (I_{p}), which was approximately seven times higher than the I_{p} obtained with the $[(\text{CoTPRu}_4)_n^{4+}\text{-GCE}]$. These results show that the AuNPs^{n-} increases the sensitivity of the electrode.

3.3. Influence of pH and scan rate

The effect of the pH solution and scan rate on the electrochemical behaviour of CC with $[(\text{CoTPRu}_4)_n^{8+}\text{-GCE}]/\text{AuNPs}^{n-}$ as a work electrode was investigated by CV. Figure 5 shows the cyclic voltammograms of CC (0.33 mmol l^{-1}) performed at different pH solutions in the range of 2.0 – 10.3 . The acetate buffer solution (pH 4.76) provides the optimum anodic peak current (I_{pa}), $E_{1/2}$ and ΔE_{p} among the investigated

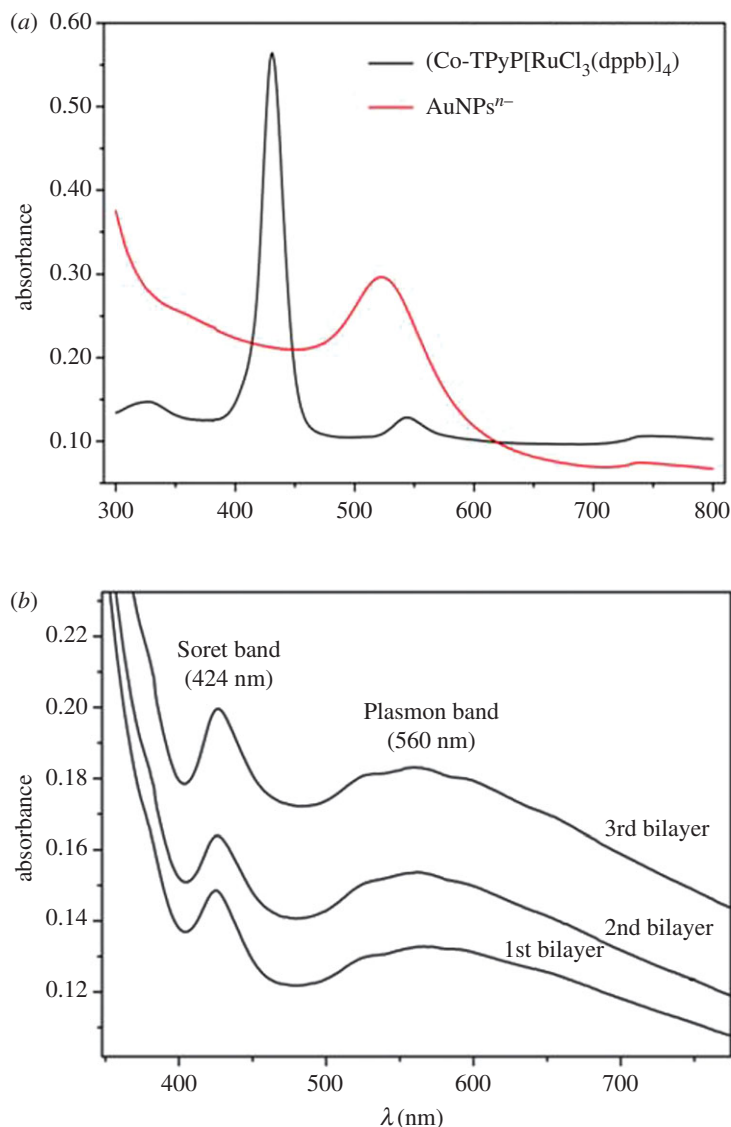


Figure 2. (a) UV/Vis spectrum of [CoTPyP{RuCl₃(dppb)}₄] in CH₂Cl₂ solution and AuNPsⁿ⁻ in toluene solution; (b) monitoring of film formation of [(CoTPRu₄)_n⁸⁺-ITO]/AuNPsⁿ⁻ by electronic spectrum onto ITO surface.

conditions. Consequently, the acetate buffer solution was chosen as the supporting electrolyte in the protocol of the quantitative analyses.

The influence of the pH solution on the peak potentials of CC was also studied. It was observed that the E_{pa} of CC decreases with the increase of the pH. A linear relationship can be noted between the E_{pa} and pH, described as follows (equation (3.1)):

$$E_{pa}(\text{mV}) = -52.5 \text{ pH} + 0.64 \quad (R^2 = 0.885). \quad (3.1)$$

A plot of E_{pa} versus pH from equation (3.1) (inside figure 5) provides a slope = -52.5 mV pH^{-1} , which is consistent with the theoretical value of -59 mV pH^{-1} [53], indicating that the redox processes of CC require the same number of protons and electrons. Figure 6 shows the result for CC detection, where the intensity of the redox peak currents increases linearly with the increase in the square root of the scan rate ($v^{1/2}$). Therefore, it indicates that the redox processes of CC on the surface of [(CoTPRu₄)_n⁸⁺-GCE]/AuNPsⁿ⁻ are controlled by diffusion, according to equations (3.2) and (3.3):

$$I_{pa} = (5.22 \pm 0.09)v^{1/2}(\text{mV}^{-2} \text{ s}^2) + (-6.69 \pm 0.95) \quad (R^2 = 0.9973), \quad (3.2)$$

$$I_{pc} = (-3.25 \pm 0.05)v^{1/2}(\text{mV}^{-2} \text{ s}^2) + (2.67 \pm 0.53) \quad (R^2 = 0.9978). \quad (3.3)$$

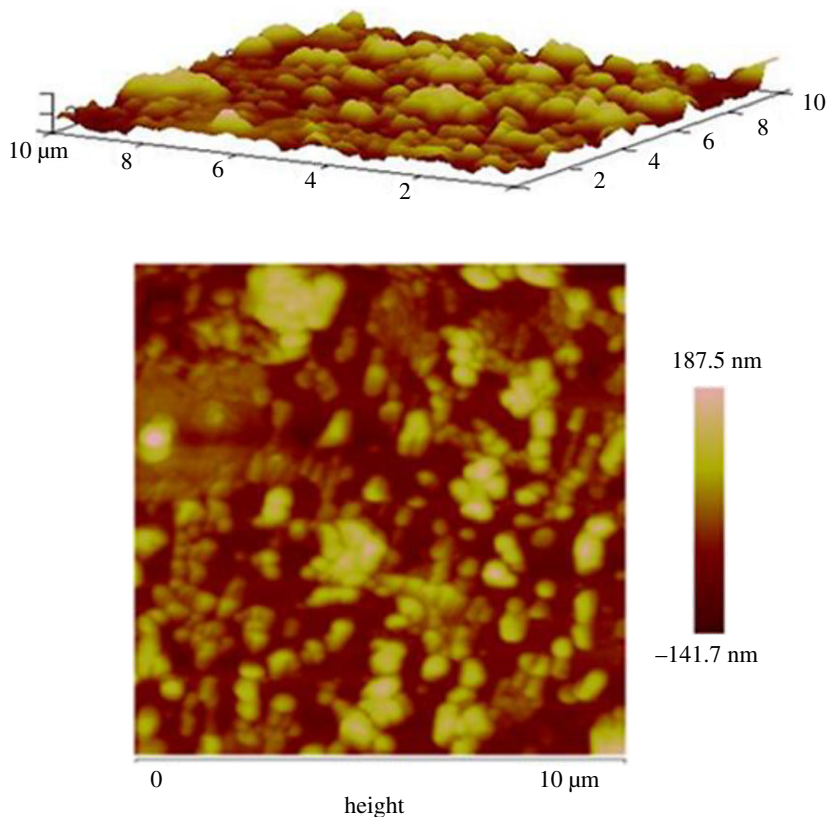


Figure 3. AFM image of a $[(\text{CoTPRu}_4)_n^{8+}\text{-ITO}]/\text{AuNPs}^{0-}$ film deposited onto ITO electrode surface.

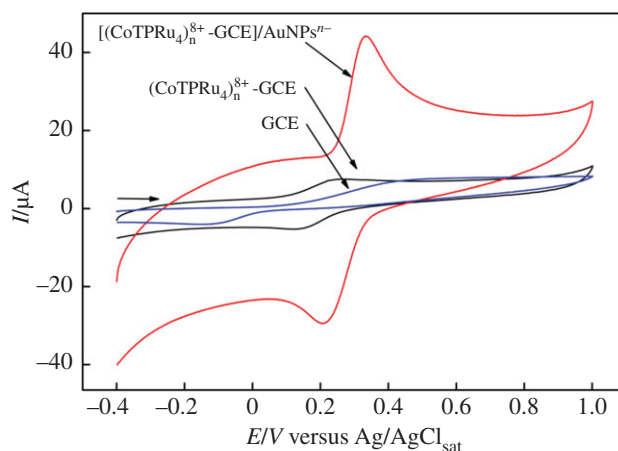


Figure 4. Cyclic voltammograms of 0.30 mmol l^{-1} CC in 0.1 mol l^{-1} acetate buffer solution (pH 4.76) at the $[(\text{CoTPRu}_4)_n^{8+}\text{-GCE}]/\text{AuNPs}^{0-}$; $(\text{CoTPRu}_4)_n^{8+}\text{-GCE}$ and GCE.

3.4. Determination of catechol

The determination of CC was carried out by CV using the $[(\text{CoTPRu}_4)_n^{8+}\text{-GCE}]/\text{AuNPs}^{0-}$. Figure 7 shows the cyclic voltammograms obtained in different concentrations of CC in 0.1 mol l^{-1} buffer acetate (pH 4.76), at a scan rate of 100 mV s^{-1} . The results show a linear relationship between the I_{pa} and the CC (equation (3.4)) in a wide range of $21\text{--}1357 \text{ } \mu\text{mol l}^{-1}$, with a detection limit of $1.4 \text{ } \mu\text{mol l}^{-1}$ ($S/N=3$) and the sensitivity of $108 \text{ } \mu\text{A } \mu\text{mol l}^{-1} \text{ cm}^{-2}$:

$$I_{\text{pa}}(\mu\text{A}) = 7.64 C_{\text{CC}}(\mu\text{mol l}^{-1}) + 21.84 \quad (R^2 = 0.998). \quad (3.4)$$

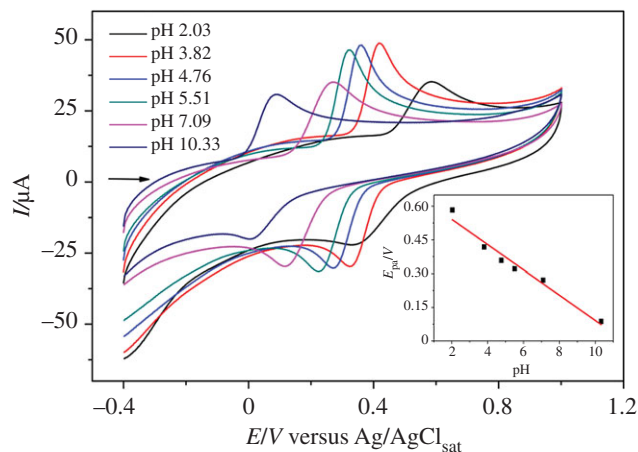


Figure 5. Cyclic voltammograms of 0.33 mmol l^{-1} CC at $[(\text{CoTPRu}_4)_n]^{8+}\text{-GCE}/\text{AuNPs}^{n-}$ in 0.1 mol l^{-1} acetate solution with different pH. Inset: effect of pH on the anodic peak potential.

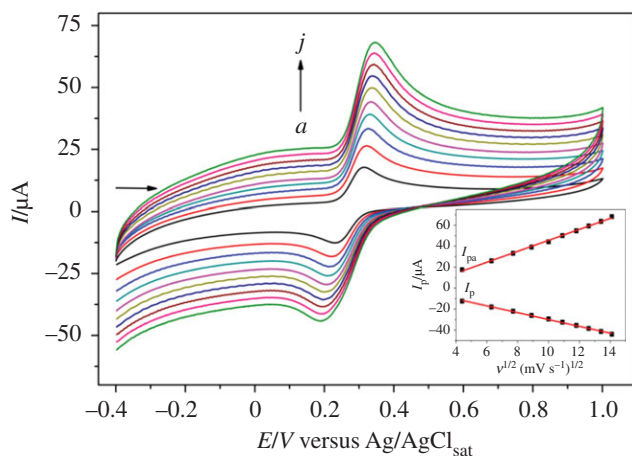


Figure 6. Cyclic voltammograms of 0.30 mmol l^{-1} CC in 0.1 mol l^{-1} acetate buffer (pH 4.76) at $[(\text{CoTPRu}_4)_n]^{8+}\text{-GCE}/\text{AuNPs}^{n-}$ at different scan rates in the range of $20\text{--}200 \text{ mV s}^{-1}$. *a-j*: 20; 40; 60; 80; 100; 120; 140; 160; 180; 200 mV s^{-1} . Inset: plot of anodic and cathodic peak currents of CC versus square root of the scan rate.

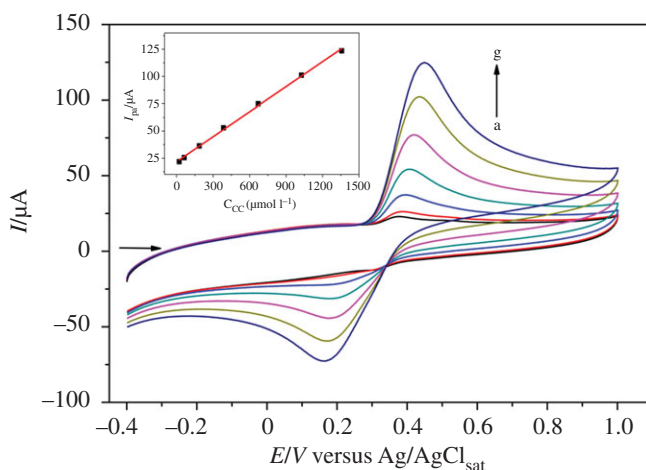


Figure 7. Cyclic voltammograms of CC as a function of concentration in the range of $21\text{--}1357 \text{ μmol l}^{-1}$ in 0.1 mol l^{-1} acetate buffer solution (pH 4.76) at the $[(\text{CoTPRu}_4)_n]^{8+}\text{-GCE}/\text{AuNPs}^{n-}$ electrode. Inset: anodic peak currents versus CC concentration.

Table 1. Performance comparison of [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ electrode for the determination of CC with other modified electrodes reported in the literature.

electrode	technique	linear range (μM)	detection limit (μM)	sensitivity (μA μM cm ⁻¹)	ref.
[(CoTPRu ₄) _n ⁸⁺ -GCE]/AuNP ⁿ⁻	CV	21–1357	1.4	108	this paper
VOTPRu-GCE	DPV	2–38	0.41	12.73	[46]
AuNPs-MPS/CPE	SWV	30–1000	1.1	—	[54]
GNPs/CNF/Au	DPV	5–350	0.36	—	[55]
Tyr-AuNPs-DHP/GCE	Amper.	2.5–95	0.17	115	[10]
Tyr-MWCNT-MNP/SPE	Amper.	10–80	7.61	4.8	[56]
Au-NP/HS(CH ₂) ₆ SH-Au electrode	CV	2–8	—	—	[57]
GCE/{Nf-fc}-MME	CV	250–2500	10.8	1.1	[58]
MWCNT-GCE	DPV	20–1200	10	—	[59]
Au-GQDs/GCE	DPV	2–50	0.87	—	[60]
GO@PDA–AuNPs	DPV	0.3–77.5	0.015	4.66	[61]
CS/MWCNTs/PDA/AuNPs/GCE	DPV	0.1–10	0.047	—	[62]
GCE/Pt–MnO ₂	DVP	3–481	—	0.25	[17]
Ag-PGly/GCE	CV	0.6–100	0.1	—	[11]

CV = cyclic voltammetry; DPV = differential pulse voltammetry; SWV = square wave voltammetry, Amper. = amperometry technique.

The results suggest that the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ electrode may be used for the determination of CC. The linear dynamic range (LDR), sensitivity (S) and detection limit (LD) of the proposed method were compared with other systems reported in the literature for the determination of CC (table 1). It can be observed that the proposed method shows good LD and an acceptable LDR, and the S of the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ was greater than most of the reported electrochemical methods.

3.5. Reproducibility and stability of the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻

The repeatability of [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ as a sensor to CC determination was evaluated by 20 successive cyclic voltammograms of a solution of CC (0.30 mmol l⁻¹). The relative standard deviations of the *I*_{pa} was 1.4% after 20 measurements, suggesting that the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ has a high level of reproducibility. Daily measurements were performed over a period of 15 days using the same electrode, conditioning the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ before each analysis (see details in §2.3). As a result, the response of the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ for CC detection was about 97% of its initial response obtained in the first day.

4. Conclusion

This work reports on the construction of an electronic device, using a bilayer based on AuNPⁿ⁻ and tetraruthenated porphyrin, with the aim of applying it as an electrochemical sensor. Having the electrostatic interaction between the cationic polymeric film of porphyrin and anionic surface of AuNPⁿ⁻, the modified electrode based on self-assembly bilayers could be built, labelled as [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻. Therefore, the number of bilayers and consequently the film thickness could be controlled. The performance of the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ was evaluated in the determination of CC, where it exhibited well-defined voltammetric peaks with enhanced peak currents for the electrochemical process of CC, compared to the uncovered GCE or [(CoTPRu₄)_n⁸⁺-GCE]. Under the optimized conditions, a wide linear range for the detection of CC was obtained, from 21 to 1357 μmol l⁻¹, and the detection limit was 1.4 μmol l⁻¹ (S/N = 3). As far as we know, the [(CoTPRu₄)_n⁸⁺-GCE]/AuNPⁿ⁻ showed an exceptional sensitivity of 108 μA μmol l⁻¹ cm⁻² for CC, much higher than that of [(CoTPRu₄)_n⁸⁺-GCE] or other similar electrochemical sensors described in the literature. This exceptional sensitivity allows an accurate determination of CC concentration, that is ideal for an

environmental samples analysis. In addition, it exhibits excellent stability over a period of 15 days, with a response of about 97%, owing to its initial response on the first day. The results suggest that the aggregation of AuNPsⁿ⁻ for the polymeric film of porphyrin provides a significant increase in the electroactive area of the electrode.

Data accessibility. All primary data collected and analysed for this study have been deposited with Figshare and can be accessed at <https://doi.org/10.6084/m9.figshare.5514994>.

Authors' contributions. L.M.S. conceived the experiments and performed the data analysis; L.M.V. carried out the synthesis of the H₂TPyP; G.H.R. contributed to writing and revising the article; A.L.B. designed the study and revised the article; L.R.D. designed and directed the study and also wrote the manuscript. All authors gave final approval for publication. Competing interests. We have no competing interests.

Funding. The authors gratefully acknowledge the financial support of CNPq, CAPES, FINEP (ctinfra 03/2007), FAPEMIG (APQ-00548-14, APQ-01867-13) and Rede Mineira de Química. The authors are thankful to the Grupo de Materiais Inorgânicos do Triângulo (GMIT) research group supported by FAPEMIG (APQ-00330-14).

Acknowledgements. The authors are grateful to Chem Axon for letting them use the academic licence from the Marvin Sketch programme.

References

- Wang J, Park JN, Wei XY, Lee CW. 2003 Room-temperature heterogeneous hydroxylation of phenol with hydrogen peroxide over Fe²⁺, Co²⁺ ion-exchanged Naβ zeolite. *Chem. Commun.* **5**, 628–629. (doi:10.1039/B22296K)
- Schweigert N, Zehnder AJB, Eggen RIL. 2001 Chemical properties of catechols and their molecular modes of toxic action in cells from microorganisms to mammals. *Environ. Microbiol.* **3**, 81–91. (doi:10.1046/j.1462-2920.2001.00176.x)
- Cui H, He C, Zhao G. 1999 Determination of polyphenols by high-performance liquid chromatography with inhibited chemiluminescence detection. *J. Chromatogr. A* **855**, 171–179. (doi:10.1016/S0021-9673(99)00670-6)
- Guan N, Zeng Z, Wang Y, Fu E, Cheng J. 2000 Open tubular capillary electrochromatography in fused-silica capillaries chemically bonded with macrocyclic dioxopolyamine. *Anal. Chim. Acta* **418**, 145–151. (doi:10.1016/S0003-2670(00)00951-X)
- Sun Y, Cui H, Li Y, Lin X. 2000 Determination of some catechol derivatives by a flow injection electrochemiluminescent inhibition method. *Talanta* **53**, 661–666. (doi:10.1016/S0039-9140(00)00550-6)
- Nagaraja P, Vasantha RA, Sunitha KR. 2001 A new sensitive and selective spectrophotometric method for the determination of catechol derivatives and its pharmaceutical preparations. *J. Pharm. Biomed. Anal.* **25**, 417–424. (doi:10.1016/S0731-7085(00)00504-5)
- Kong Y, Chen X, Wang W, Chen Z. 2011 A novel palygorskite-modified carbon paste amperometric sensor for catechol determination. *Anal. Chim. Acta* **688**, 203–207. (doi:10.1016/j.aca.2011.01.007)
- Han JT, Huang KJ, Li J, Liu YM, Yu M. 2012 β-cyclodextrin-cobalt ferrite nanocomposite as enhanced sensing platform for catechol determination. *Colloids Surf. B* **98**, 58–62. (doi:10.1016/j.colsurfb.2012.05.003)
- Mazloum-Ardakani M, Hosseinzadeh L, Taleat Z. 2015 Synthesis and electrocatalytic effect of Ag@Pt core-shell nanoparticles supported on reduced graphene oxide for sensitive and simple label-free electrochemical aptasensor. *Biosens. Bioelectron.* **74**, 30–36. (doi:10.1016/j.bios.2015.05.072)
- Vicentini FC, Garcia LLC, Figueiredo-Filho LCS, Janegitz BC, Fatibello-Filho O. 2016 A biosensor based on gold nanoparticles, dihexadecylphosphate, and tyrosinase for the determination of catechol in natural water. *Enzyme Microb. Technol.* **84**, 17–23. (doi:10.1016/j.enzmictec.2015.12.004)
- Chen M, Li X, Ma X. 2012 Selective determination of catechol in wastewater at silver doped polyglycine modified film electrode. *Int. J. Electrochem. Sci.* **7**, 2616–2622.
- Ahmad A et al. 2015 Size dependent catalytic activities of green synthesized gold nanoparticles and electro-catalytic oxidation of catechol on gold nanoparticles modified electrode. *RSC Adv.* **5**, 99 364–99 377. (doi:10.1039/CSRA20096B)
- Chen Y, Liu X, Zhang S, Yang L, Liu M, Zhang Y, Yao S. 2017 Ultrasensitive and simultaneous detection of hydroquinone, catechol and resorcinol based on the electrochemical co-reduction prepared Au-Pd nanoflower/reduced graphene oxide nanocomposite. *Electrochim. Acta* **231**, 677–685. (doi:10.1016/j.electacta.2017.02.060)
- Maikap A, Mukherjee K, Mondal BN, Mandal N. 2016 Zinc oxide thin film based nonenzymatic electrochemical sensor for the detection of trace level catechol. *RSC Adv.* **6**, 64 611–64 617. (doi:10.1039/C6RA09598D)
- Xiong W, Wu M, Zhou B, Liu S. 2014 The highly sensitive electrocatalytic sensing of catechol using a gold/titanium dioxide nanocomposite-modified gold electrode. *RSC Adv.* **4**, 32 092–32 099. (doi:10.1039/C4RA04256e)
- Yin H, Zhang Q, Zhou Y, Ma Q, Liu T, Zhu L, Ai S. 2011 Electrochemical behavior of catechol, resorcinol and hydroquinone at graphene-chitosan composite film modified glassy carbon electrode and their simultaneous determination in water samples. *Electrochim. Acta* **56**, 2748–2753. (doi:10.1016/j.electacta.2010.12.060)
- Unnikrishnan B, Ru PL, Chen SM. 2012 Electrochemically synthesized Pt-MnO₂ composite particles for simultaneous determination of catechol and hydroquinone. *Sens. Actuators B* **169**, 235–242. (doi:10.1016/j.snb.2012.04.075)
- Chen L, Tang Y, Wang K, Liu C, Luo S. 2011 Direct electrodeposition of reduced graphene oxide on glassy carbon electrode and its electrochemical application. *Electrochem. Commun.* **13**, 133–137. (doi:10.1016/j.elecom.2010.11.033)
- Du H, Ye J, Zhang J, Huang X, Yu C. 2011 A voltammetric sensor based on graphene-modified electrode for simultaneous determination of catechol and hydroquinone. *J. Electroanal. Chem.* **650**, 209–213. (doi:10.1016/j.jelechem.2010.10.002)
- Guo Q, Huang J, Chen P, Liu Y, Hou H, You T. 2012 Simultaneous determination of catechol and hydroquinone using electrospun carbon nanofibers modified electrode. *Sens. Actuators B* **163**, 179–185. (doi:10.1016/j.snb.2012.01.032)
- Zheng L, Xiong L, Li Y, Xu J, Kang X, Zou Z, Yang S, Xia J. 2013 Facile preparation of polydopamine-reduced graphene oxide nanocomposite and its electrochemical application in simultaneous determination of hydroquinone and catechol. *Sens. Actuators B* **177**, 344–349. (doi:10.1016/j.snb.2012.11.006)
- Yu J, Du W, Zhao F, Zeng B. 2009 High sensitive simultaneous determination of catechol and hydroquinone at mesoporous carbon CMK-3 electrode in comparison with multi-walled carbon nanotubes and Vulcan XC-72 carbon electrodes. *Electrochim. Acta* **54**, 984–988. (doi:10.1016/j.electacta.2008.08.029)
- Mazloum-Ardakani M, Khoshroo A. 2014 High performance electrochemical sensor based on fullerene-functionalized carbon nanotubes/ionic liquid: determination of some catecholamines. *Electrochem. Commun.* **42**, 9–12. (doi:10.1016/j.elecom.2014.01.026)
- Kalimuthu P, Sivanesan A, John SA. 2012 Fabrication of optochemical and electrochemical sensors using thin films of porphyrin and phthalocyanine derivatives. *J. Chem. Sci.* **124**, 1315–1325. (doi:10.1007/s12039-012-0330-5)
- Quintino MSM, Araki K, Toma HE, Angnes L. 2006 Amperometric quantification of sodium metabisulfite in pharmaceutical formulations utilizing tetraarthenated porphyrin film modified electrodes and batch injection analysis. *Talanta* **68**, 1281–1286. (doi:10.1016/j.talanta.2005.07.034)

26. Schaming D *et al.* 2011 Easy methods for the electropolymerization of porphyrins based on the oxidation of the macrocycles. *Electrochim. Acta* **56**, 10 454–10 463. (doi:10.1016/j.electacta.2011.02.064)
27. Dreyse P, Isaacs M, Calfumán K, Cáceres C, Aliaga A, Aguirre MJ, Villagra D. 2011 Electrochemical reduction of nitrite at poly-[Ru(5-NO₂-phen)2Cl] tetrapyrrolylporphyrin glassy carbon modified electrode. *Electrochim. Acta* **56**, 5230–5237. (doi:10.1016/j.electacta.2011.03.028)
28. Fan S, Zhu Y, Lui R, Zhang H, Wang ZS, Wu H. 2016 A porphyrin derivative for the fabrication of highly stable and sensitive electrochemical sensor and its analytical applications. *Sens. Actuators B* **223**, 206–213. (doi:10.1016/j.snb.2016.04.049)
29. Paolesse R, Nardis S, Monti D, Stefanelli M, Di Natale C. 2016 Porphyrinoids for chemical sensor applications. *Chem. Rev.* **117**, 2517–2584. (doi:10.1021/acs.chemrev.6b00361)
30. Fagadar-Cosma E, Sebarchievici I, Lascu A, Creanga I, Palade A, Birdeanu M, Taranu B, Fagadar-Cosma G. 2016 Optical and electrochemical behavior of new nano-sized complexes based on gold-colloid and Co-porphyrin derivative in the presence of H₂O₂. *J. Alloys Compd* **685**, 896–904. (doi:10.1016/j.jallcom.2016.06.246)
31. Sebarchievici I, Taranu BO, Birdeanu M, Rus SF, Fagadar-Cosma E. 2016 Electrocatalytic behaviour and application of manganese porphyrin/gold nanoparticle-surface modified glassy carbon electrodes. *Appl. Surf. Sci.* **390**, 131–140. (doi:10.1016/j.apsusc.2016.07.158)
32. Biesaga M, Pырzyska K, Trojanowicz M. 2000 Porphyrins in analytical chemistry. A review. *Talanta* **51**, 209–224. (doi:10.1016/S0039-9140(99)00291-X)
33. Toma HE, Araki K. 2000 Supramolecular assemblies of ruthenium complexes and porphyrins. *Coord. Chem. Rev.* **196**, 307–329. (doi:10.1016/S0010-8545(99)00041-7)
34. Priyadarshini E, Pradhan N. 2017 Gold nanoparticles as efficient sensors in colorimetric detection of toxic metal ions: a review. *Sens. Actuators B* **238**, 888–902. (doi:10.1016/j.snb.2016.06.081)
35. Khudaish EA, Rather JA. 2016 Electrochemical studies of dopamine under stagnant and convective conditions at a sensor based on gold nanoparticles hosted in poly(triaminopyrimidine). *J. Electroanal. Chem.* **776**, 206–212. (doi:10.1016/j.jelechem.2016.06.041)
36. Vinoth V, Wu JJ, Asiri AN, Anandan S. 2015 Simultaneous detection of dopamine and ascorbic acid using silicate network interlinked gold nanoparticles and multi-walled carbon nanotubes. *Sens. Actuators B* **210**, 731–741. (doi:10.1016/j.snb.2015.01.040)
37. Krajczewski J, Kotataj K, Kudelski A. 2017 Plasmonic nanoparticles in chemical analysis. *RSC Adv.* **7**, 17 559–17 576. (doi:10.1039/c7ra01034f)
38. Liana DD, Raguse B, Wiecek L, Baxter GR, Chuah K, Gooding JJ, Chow E. 2013 Sintered gold nanoparticles as an electrode material for paper-based electrochemical sensors. *RSC Adv.* **3**, 8683–8691. (doi:10.1039/c3ra00102d)
39. Brust M, Walker M, Bethell D, Schiffrin DJ, Whyman R. 1994 Synthesis of thiol-derivatised gold nanoparticles in a two-phase liquid-liquid system. *J. Chem. Soc. Chem. Commun.* **7**, 801–802. (doi:10.1039/C39940000801)
40. Eustis S, El-Sayed MA. 2006 Why gold nanoparticles are more precious than pretty gold: noble metal surface plasmon resonance and its enhancement of the radiative and nonradiative properties of nanocrystals of different shapes. *Chem. Soc. Rev.* **35**, 209–217. (doi:10.1039/B514191E)
41. Katz E, Willner I, Wang J. 2004 Electroanalytical and bioelectroanalytical systems based on metal and semiconductor nanoparticles. *Electroanalysis* **16**, 19–44. (doi:10.1002/elan.200302930)
42. De Oliveira KM, Dos Santos TCC, Dinelli LR, Marinho JZ, Lima RC, Bogado AL. 2013 Aggregates of gold nanoparticles with complexes containing ruthenium as modifiers in carbon paste electrodes. *Polyhedron* **50**, 410–417. (doi:10.1016/j.poly.2012.11.014)
43. Ferreira VF, Do Prado CRA, Rodrigues CM, Otubo L, Batista AA, Da Cruz Jr JW, Ellena J, Dinelli LR, Bogado AL. 2014 Modified glassy carbon electrode with AuNPs using cis-[RuCl(dppb)(bipy)(4-vpy)]⁺ as crossed linking agent. *Polyhedron* **78**, 46–53. (doi:10.1016/j.poly.2014.04.024)
44. Souza LCM *et al.* 2016 Influence of gold nanoparticles applied to catalytic hydrogenation of acetophenone with cationic complexes containing ruthenium. *RSC Adv.* **6**, 53 130–53 139. (doi:10.1039/C6RA05616D)
45. Da Silva MM, Ribeiro GH, Batista AA, De Faria AM, Bogado AL, Dinelli LR. 2013 Electropolymerized supramolecular tetra-ruthenated porphyrins applied as a voltammetric sensor. *J. Braz. Chem. Soc.* **24**, 1772–1780. (doi:10.5935/0103-5053.20130222)
46. Ribeiro GH, Vilarinho LM, Ramos TDS, Bogado AL, Dinelli LR. 2015 Electrochemical behavior of hydroquinone and catechol at glassy carbon electrode modified by electropolymerization of tetra-ruthenated oxovanadium porphyrin. *Electrochim. Acta* **176**, 394–401. (doi:10.1016/j.electacta.2015.06.139)
47. Dinelli LR, Poelsitz GV, Castellano EE, Ellena J, Galembeck SE, Batista AA. 2009 On an electrode modified by a supramolecular ruthenium mixed valence (Rull/RullII) diphosphine-porphyrin assembly. *Inorg. Chem.* **48**, 4692–4700. (doi:10.1021/ic702471d)
48. Brown KR, Walter DG, Natan MJ. 2000 Seeding of colloidal Au nanoparticle solutions. Improved control of particle size and shape. *Chem. Mater.* **12**, 306–313. (doi:10.1021/cm980065p)
49. Fleischer EB. 1962 α , β , γ , δ -Tetra-(4-pyridyl)-porphyrine. Some of its metal complexes. *Inorg. Chem.* **3**, 493–495. (doi:10.1021/ic50003a010)
50. Bonnett R, Brewer P, Noro K, Noro T. 1978 Chemistry of vanadyl porphyrins. *Tetrahedron* **34**, 379–385. (doi:10.1016/S0040-4020(01)93596-3)
51. Ghosh SK, Patra R, Rath SP. 2008 Axial ligand coordination in sterically strained vanadyl porphyrins: synthesis, structure, and properties. *Inorg. Chem.* **47**, 9848–9856. (doi:10.1021/ic800714w)
52. Dinelli LR *et al.* 1999. Synthesis and characterization of [RuCl₃(P-P)(H₂O)] complexes; P-P = achiral or chiral, chelating ditertiary phosphine ligands. *Inorg. Chem.* **38**, 5341–5345. (doi:10.1021/ic990130c)
53. Compton RG, Banks CE. 2011 *Understanding voltammetry*. London, UK: Imperial College Press.
54. Tashkhourian J, Daneshi M, Nami-Ana F, Behbahani M, Bagheri A. 2016 Simultaneous determination of hydroquinone and catechol at gold nanoparticles mesoporous silica modified carbon paste electrode. *J. Hazard. Mater.* **318**, 117–124. (doi:10.1016/j.jhazmat.2016.06.049)
55. Huo Z, Zhou Y, Liu Q, He X, Liang Y, Xu M. 2011 Sensitive simultaneous determination of catechol and hydroquinone using a gold electrode modified with carbon nanofibers and gold nanoparticles. *Microchim. Acta* **173**, 119–125. (doi:10.1007/s00604-010-0530-y)
56. Pérez-López B, Merkoçi A. 2011 Magnetic nanoparticles modified with carbon nanotubes for electrocatalytic magnetoswitchable biosensing applications. *Adv. Funct. Mater.* **21**, 255–260. (doi:10.1002/adfm.201001306)
57. Su L, Mao L. 2006 Gold nanoparticle/alkanedithiol conductive films self-assembled onto gold electrode: electrochemistry and electroanalytical application for voltammetric determination of trace amount of catechol. *Talanta* **70**, 68–74. (doi:10.1016/j.talanta.2006.01.015)
58. Kumar AS, Swetha P, Pillai KC. 2010 Enzyme-less and selective electrochemical sensing of catechol and dopamine using ferrocene bound Nafion membrane modified electrode. *Anal. Methods* **2**, 1962–1968. (doi:10.1039/C0AY00430H)
59. Xu Z, Chen X, Qu X, Dong S. 2004 Electrocatalytic oxidation of catechol at multi-walled carbon nanotubes modified electrode. *Electroanalysis* **16**, 684–687. (doi:10.1002/elan.200302843)
60. Zhao X, He D, Wang Y, Hu Y, Fu C. 2016 Au nanoparticles and graphene quantum dots Co-modified glassy carbon electrode for catechol sensing. *Chem. Phys. Lett.* **647**, 165–169. (doi:10.1016/j.cplett.2016.01.019)
61. Palanisamy S, Thangavelu K, Chen SM, Thirumalraj B, Liu XH. 2016 Preparation and characterization of gold nanoparticles decorated on graphene oxide@polydopamine composite: application for sensitive and low potential detection of catechol. *Sens. Actuators B* **233**, 298–306. (doi:10.1016/j.snb.2016.04.083)
62. Wang Y, Xiong Y, Qu J, Qu J, Li S. 2016 Selective sensing of hydroquinone and catechol based on multiwalled carbon nanotubes/polydopamine/gold nanoparticles composites. *Sens. Actuators B* **223**, 501–508. (doi:10.1016/j.snb.2015.09.117)